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1 **Is carotid body physiological O₂ sensitivity determined by a unique mitochondrial**
2 **phenotype?**

3
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18

1 **Abstract**

2 The mammalian carotid body (CB) is the primary arterial chemoreceptor that responds to acute
3 hypoxia, initiating systemic protective reflex responses that act to maintain O₂ delivery to the
4 brain and vital organs. The CB is unique in that it is stimulated at O₂ levels above those that
5 begin to impact on the metabolism of most other cell types. Whilst a large proportion of the CB
6 chemotransduction cascade is well defined, the identity of the O₂ sensor remains highly
7 controversial. This short review evaluates whether the mitochondria can adequately function as
8 acute O₂ sensors in the CB. We consider the similarities between mitochondrial poisons and
9 hypoxic stimuli in their ability to activate the CB chemotransduction cascade and initiate rapid
10 cardiorespiratory reflexes. We evaluate whether the mitochondria are required for the CB to
11 respond to hypoxia. We also discuss if the CB mitochondria are different to those located in
12 other non-O₂ sensitive cells, and what might cause them to have an unusually low O₂ binding
13 affinity. In particular we look at the potential roles of competitive inhibitors of mitochondrial
14 complex IV such as nitric oxide in establishing mitochondrial and CB O₂-sensitivity. Finally,
15 we discuss novel signaling mechanisms proposed to take place within and downstream of
16 mitochondria that link mitochondrial metabolism with cellular depolarization.

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1 **1 Introduction-the carotid body and hypoxia**

2 The mammalian carotid body (CB) is a highly specialized sensory organ derived from the
3 neural crest. The sensory units of the CB are the ‘glomus’ or ‘type I’ cells that respond to a
4 variety of stimuli including hypoxia, hypercapnia, acidosis and hormones thereby allowing the
5 CB to function as a polymodal receptor (Kumar and Bin-Jaliah, 2007; Ribeiro et al., 2013;
6 Thompson et al., 2016). Type I cell activation leads to stimulation of adjacent chemoafferent
7 fibers that relay sensory information into the central nervous system. The physiological
8 consequence of CB stimulation is therefore the initiation of series of systemic protective
9 reflexes characterized by increased ventilation, tachycardia, systemic vasoconstriction and
10 adrenaline release from the adrenal medulla (Kumar, 2009).

11
12 There is now a general consensus that a series of key processes contribute to the CB hypoxic
13 chemotransduction cascade. These include attenuation of outward K^+ current, type I cell
14 depolarization, Ca^{2+} influx through L-type Ca^{2+} channels, neurosecretion and chemoafferent
15 excitation (Kumar and Prabhakar, 2012; Lopez-Barneo et al., 2016). Single type I cells are
16 exquisitely sensitive to O_2 and rapid (within ms) activation of the hypoxic chemotransduction
17 cascade initiates at PO_2 s of between 20-40mmHg (Biscoe and Duchon, 1990; Buckler and
18 Vaughan-Jones, 1994; Buckler and Turner, 2013); a level considerably greater than that at
19 which cell metabolism is affected in O_2 -insensitive cells.

20
21 What remains highly controversial is the molecular identity of the specific O_2 sensor within the
22 type I cell. We would argue that the physiological O_2 sensor should exhibit certain key features:
23 1. expression in the type I cell permitting intrinsic O_2 sensitivity; 2. the ability to bind O_2 ; 3. its
24 binding of O_2 occurs over the physiological range at which the type I cell is stimulated; 4. it is
25 required for the CB to be stimulated by hypoxia; and 5. it is able to activate the CB transduction
26 cascade within milliseconds. Many proposed sensors fit one or two of these criteria but few
27 have been shown to adequately comply with all five.

28
29 In mammalian cells, O_2 is the terminal electron acceptor in the mitochondrial respiratory chain.
30 Continuous binding and reduction of O_2 in the CuB/haem a_3 (cytochrome a_3) binuclear center of
31 complex IV drives mitochondrial electron transport and promotes activation of the
32 mitochondrial ATP synthase. The long-established, mitochondrial hypothesis for
33 chemoreception proposes that CB excitation induced by hypoxia is initiated by a reduction in
34 O_2 dependent mitochondrial energy respiration. This review will briefly critique the current
35 evidence as to whether the mitochondria can be considered the functionally relevant O_2 sensors
36 within the CB.

37 38 **2 Mitochondrial inhibitors mimic all aspects of the carotid body hypoxic** 39 **chemotransduction cascade**

40 All mitochondrial poisons induce chemoafferent excitation (Krylov and Anichkov, 1968;
41 Mulligan et al., 1981; Obeso et al., 1989; Donnelly et al., 2014; Holmes et al., 2016; Holmes et
42 al., 2017) leading to rapid increases in ventilation (Owen and Gesell, 1931), heart rate and
43 arterial blood glucose (Alvarez-Buylla and de Alvarez-Buylla, 1988). Chemoafferent responses
44 are rapid, dose dependent and reversible (Donnelly et al., 2014; Holmes et al., 2016) and the
45 magnitude of the rise in chemoafferent frequency caused by saturating concentrations is
46 consistent with those evoked by severe hypoxia or anoxia (Krylov and Anichkov, 1968;
47 Mulligan et al., 1981; Obeso et al., 1989). Furthermore, mitochondrial inhibitors and uncouplers
48 augment neurotransmitter secretion, confirming an action through the type I cell rather than the
49 afferent nerve endings (Obeso et al., 1989; Rocher et al., 1991). Despite the strong consistency
50 between all of the different types of mitochondrial poisons (both in the older and more recent

1 studies), it should be noted that some of the pharmacological agents used may not have acted
2 selectively on the mitochondria and so the conclusions should be viewed with a certain degree
3 of caution.

4
5 Mitochondrial poisons cause fast (within ms) type I cell depolarization and increases in $[Ca^{2+}]_i$.
6 The size and timing of the $[Ca^{2+}]_i$ rise observed using many different mitochondrial inhibitors or
7 uncouplers closely resembles that seen in hypoxia (Biscoe et al., 1989; Biscoe and Duchen,
8 1990; Wyatt and Buckler, 2004; Buckler, 2011). As with hypoxia (Buckler and Vaughan-Jones,
9 1994; Urena et al., 1994), the increases in $[Ca^{2+}]_i$ are dependent on cellular depolarization and
10 extracellular Ca^{2+} influx through voltage gated Ca^{2+} channels (Wyatt and Buckler, 2004).
11 TASK1/3, TREK-1, BK_{Ca} , $K_v4.1$ and $K_v4.3$, have all been shown to be expressed in the CB and
12 inhibited by hypoxia (Buckler et al., 2000; Sanchez et al., 2002; Williams et al., 2004;
13 Yamamoto and Taniguchi, 2006; Kim et al., 2009; Kreneisz et al., 2009; Turner and Buckler,
14 2013; Wang et al., 2017b). Of these, TASK1/3 and TASK-like K^+ currents are diminished by a
15 multitude of mitochondrial inhibitors leading to membrane depolarization (Barbe et al., 2002;
16 Wyatt and Buckler, 2004; Buckler, 2011; Turner and Buckler, 2013; Kim et al., 2015a).

17
18 Recent evidence has revealed the presence of TRP and other non-selective Ca^{2+} -activated cation
19 currents in type I cells that are activated by hypoxia (Kumar et al., 2006; Kang et al., 2014; Kim
20 et al., 2015b; Wang et al., 2017a). Although intriguing, the full functional relevance of these
21 currents in type I cell O_2 -sensing remains to be further characterized, and in particular whether
22 these currents can be upregulated to preserve O_2 sensing in the absence of TASK channels
23 (Turner and Buckler, 2013). Current evidence suggests that mitochondrial inhibitors are also
24 capable of increasing these inward depolarizing currents (Kim et al., 2015b; Wang et al.,
25 2017a).

26 27 **3 Mitochondria are necessary for the carotid body to respond to hypoxia**

28 The presence of functional mitochondria does appear necessary for the CB to respond fully to
29 hypoxia. For instance, cyanide, rotenone and FCCP all attenuate TASK channel currents in such
30 a way that prevents any further reduction by hypoxia (Wyatt and Buckler, 2004). In intact CB
31 preparations oligomycin, cyanide and azide all reduce or abolish subsequent chemoafferent
32 responses to hypoxia (Mulligan et al., 1981; Donnelly et al., 2014). Some of the attenuation
33 observed in these experiments may have been due to impairment of oxidative phosphorylation
34 in the chemoafferent fibers, limiting their excitability. Any such impairment is not apparent in
35 response to physiological levels of hypoxia, since the PO_2 that activates the type I cells occurs
36 at a much higher level than those which would decrease mitochondrial function in the
37 chemoafferent fibers. As such, chemoafferent responses to sustained hypoxia are better
38 maintained than those in response to sustained high doses of mitochondrial inhibitors (Mulligan
39 et al., 1981).

40
41 In a recent study the importance of mitochondrial complex I was tested by developing mice
42 deficient in *Ndufs2* (a gene coding for NADH dehydrogenase [ubiquinone] iron-sulfur protein
43 2- a component of complex I that participates in ubiquinone binding) in tyrosine hydroxylase
44 positive cells (Fernandez-Aguera et al., 2015). Type I cells isolated from these mice were
45 insensitive to hypoxia; they lacked any hypoxia-induced K^+ current attenuation, $[Ca^{2+}]_i$
46 elevation or neurotransmitter release. Furthermore, these mice failed to increase respiratory
47 frequency when breathing 10% O_2 . This work supported a previous study in which type I cell
48 hypoxic chemosensitivity was abolished in the presence of rotenone (Ortega-Saenz et al., 2003).
49 The authors propose a mechanism whereby exposure to hypoxia promotes reverse electron

1 transport and ROS/NADH generation via complex I which is driven by a high rate of succinate
2 oxidation at complex II. Accordingly, they have recently shown that genetic and
3 pharmacological deactivation of complex II completely blocks type I cell hypoxic sensitivity
4 (Gao et al., 2017).

5
6 This intriguing and elegant hypothesis does however show some discrepancies with evidence
7 from earlier reports. For instance, similar experiments performed on CBs with heterozygous
8 *Sdhb* knock out displayed an augmented, rather than depressed basal activity and had a
9 completely preserved hypoxic response (Piruat et al., 2004). Furthermore, when rat type I cells
10 were exposed to tetramethyl-p-phenylenediamine (TMPD) and ascorbate in the presence of
11 rotenone, there was still a robust elevation in Ca^{2+} upon hypoxic stimulation (Wyatt and
12 Buckler, 2004). This would suggest that feeding electrons into cytochrome c is sufficient to
13 sustain type I cell hypoxic sensitivity even when complex I activity (and ROS generation) is
14 inhibited. Complex IV activity rather than complex I and II may therefore be necessary for
15 hypoxic chemotransduction. The same report also showed that application of H_2O_2 was unable
16 to excite the type I cell directly. This observation is consistent with the lack of effect of multiple
17 anti-oxidants used in other CB preparations and animal species (Sanz-Alfayate et al., 2001;
18 Agapito et al., 2009; Gomez-Nino et al., 2009). Interestingly, using novel *ex vivo* CB culture
19 techniques combined with FRET based ROS sensors, Bernardini and co-workers deduced that
20 type I cell ROS actually decreases in hypoxia due to reduction in NADPH oxidase activity (an
21 alternative ROS source) (Bernardini et al., 2015). Clearly there is a need for reconciliation
22 between these findings.

23 24 **4 The carotid body mitochondria are unique and have a low threshold for O_2**

25 The evidence that mitochondria are required for CB O_2 chemotransduction and that
26 mitochondrial inhibitors can cause chemoexcitation, is not enough to define them as the O_2 -
27 sensors in the CB. Clearly, mitochondria are able to bind O_2 . However, the K_m of the
28 cytochrome a_3 for O_2 is reported to be $<1\text{mmHg}$ in isolated mitochondria and between 1-
29 5mmHg in dissociated cells and tissue preparations, with little variation existing between
30 different cell types (Wilson et al., 1988; Tamura et al., 1989). This is far lower than the PO_2 at
31 which the CB type I cells begin to be activated and, for this reason, is a common argument
32 against the mitochondrial hypothesis.

33
34 However, there is now a substantial body of evidence indicating that the CB type I cell
35 mitochondria are unique. Experiments performed by Mills & Jöbsis, were the first to identify an
36 unusually low affinity cytochrome a_3 within the CB (Mills and Jobsis, 1970; 1972). Using
37 absorbance spectra, they estimated that 43-67.5% of total cytochrome a_3 within the intact CB
38 preparation had a remarkably low O_2 affinity. This fraction was reported to be almost 100%
39 reduced at PO_2 s between 7-9mmHg and 50% reduced at a PO_2 as high as 90mmHg. In contrast,
40 the remaining fraction was only 50% reduced at a PO_2 of approximately 0.8mmHg, comparable
41 to cytochrome a_3 found in other tissues (Gnaiger, 2001). Thus, the CB appeared to express both
42 low and high affinity subtypes of cytochrome a_3 . At that time, the specific cellular location(s) of
43 each was unclear. Later experiments utilized the photolabile binding of CO, to deduce that
44 saturation of cytochrome a_3 with CO prevented any additional chemoafferent excitation during
45 hypoxia, implying that not only was the cytochrome a_3 in the CB unusual, it was also required
46 for O_2 -sensing (Wilson et al., 1994; Lahiri et al., 1999). It should be pointed out that the
47 concentrations of CO used in these studies could have directly modified the activity of the BK_{Ca}
48 channel (Williams et al., 2004; Williams et al., 2008) and the generation of H_2S (Yuan et al.,
49 2015) and as such some of the observations could be related to mechanisms independent of the
50 mitochondria.

1
2 In dissociated rabbit type I cell clusters, mitochondrial electron transport begins to be inhibited
3 at a high PO₂ value of approximately 40mmHg (Duchen and Biscoe, 1992a). PO₂-NADH
4 response curves demonstrate a significant ‘right shift’ in type I cells compared to sensory
5 neurons, indicative of a heightened and distinctive O₂ sensitivity. In addition, mitochondrial
6 depolarization occurs at higher PO₂s compared to O₂-insensitive cells (Duchen and Biscoe,
7 1992b). More recent work has verified the “right shifts” in both PO₂-NADH and PO₂-rh-123
8 response curves in rat type I cells, confirming that the unusually low mitochondrial O₂ affinity
9 is conserved in multiple species (Turner and Buckler, 2013). By isolating complex IV activity
10 with a cocktail of mitochondrial inhibitors plus TMPD and ascorbate, the authors were able to
11 reveal that complex IV activity is a component of the mitochondria with the exceptionally low
12 O₂ affinity. Importantly, type I cell hypoxic response curves for electron transport inhibition,
13 mitochondrial depolarization and complex IV run-down display considerable overlap with the
14 rise in Ca²⁺, indicating that these processes are intimately linked. Therefore, it does appear that
15 type I cell mitochondria have a highly specialized *low affinity* for O₂ due to an altered
16 function/subtype of cytochrome a₃ in complex IV that predisposes CB energy metabolism to
17 being impaired at high O₂ tensions. It is likely that the high affinity cytochrome a₃ in the CB
18 described by Mills & Jöbsis is located in the non-O₂ sensing tissue such as the, nerve endings,
19 blood vessels and type II cells.

20
21 Understanding the mechanism linking a fall in mitochondrial O₂ consumption with K⁺ channel
22 inhibition (or cation channel activation) is contentious. As previously mentioned, there could be
23 a role for elevated mitochondrial ROS generation but this is still to be validated (Fernandez-
24 Aguera et al., 2015). Another possibility is an alteration in cytosolic nucleotides. Switching
25 from a cell attached to inside-out patch configuration diminishes background K⁺ channel
26 activity, suggesting that a basal level of an intracellular factor(s) activates TASK channels in
27 normoxia (Varas et al., 2007). Addition of 5mM MgATP in the inside-out configuration can
28 restore about 50% of this background K⁺ channel activity. Both mitochondrial inhibition and
29 hypoxia also significantly elevate free Mg²⁺, consistent with a decrease in MgATP. Thus, the
30 fall in MgATP during hypoxia is likely to attenuate a significant proportion of TASK-current
31 leading to depolarization. However, the remaining modulators that account for the other 50% of
32 TASK current are still to be identified.

33
34 Another proposed mediator of TASK channel activity that is sensitive to changes in cytosolic
35 nucleotide concentrations is AMPK (Wyatt et al., 2007). However, initial favorable studies
36 based on pharmacological evidence have since been challenged by the finding that the AMPK-
37 α₁α₂ deficient CB retains complete O₂-sensitivity (Mahmoud et al., 2016). Other groups have
38 also shown that pharmacological targeting of AMPK does not impact on the hypoxia-induced
39 K⁺ channel inhibition (Kim et al., 2014). Discrepancies may arise from the non-selectivity of
40 the drugs used to evaluate AMPK function and potential redundancy mechanisms known to
41 develop in genetic animal models (Nowak et al., 1997). A final hypothesis is that a build-up in
42 lactate upon mitochondrial inhibition in hypoxia activates the olfactory receptor Olfr78 (Chang
43 et al., 2015). However, the concentration of lactate necessary to elevate Ca²⁺ in an intact CB
44 preparation appears to be quite high (30mM) and whether local levels reach this threshold
45 during hypoxia is uncertain. We await a mechanism demonstrating how activation of Olfr78 (a
46 G-protein-coupled receptor) modulates TASK or cation channel activity. A summary of the
47 proposed O₂-sensitive mitochondrial signaling pathways in the CB is presented in Figure 1.

48
49 **5 What determines the low O₂ affinity of the carotid body mitochondria?**

1 We propose that there are 2 potential means to account for the extraordinary O₂ sensitivity of
2 type I cell mitochondria. First, there could be a high level of production of a cytosolic factor
3 that is able to freely diffuse into the mitochondria and then compete with O₂ binding in complex
4 IV (Figure 2). We predict that this competition would render mitochondrial electron transport
5 more susceptible to subsequent falls in O₂. We recently tested this by applying exogenous nitrite
6 to CBs and subsequently measuring hypoxic sensitivity (Holmes et al., 2016). Nitrite is reduced
7 within the mitochondria to generate local NO, a recognized competitive inhibitor of complex IV
8 (Brown and Cooper, 1994; Cleeter et al., 1994; Castello et al., 2006; Basu et al., 2008; Castello
9 et al., 2008). Moderate basal inhibition of the CB mitochondria by nitrite exaggerated the
10 subsequent chemoafferent excitation during hypoxia signifying an increase in CB O₂ sensitivity.
11 Therefore, we validated the idea that CB hypoxic sensitivity could be adjusted by a factor
12 capable of competing with O₂ in the mitochondria and suggested a physiological role for
13 endogenous NO in establishing type I cell mitochondrial O₂-sensitivity. Measurable amounts of
14 NO have been detected in mitochondrial of type I cells (Yamamoto et al., 2006). A possible
15 source is nitric oxide synthase 3 (NOS-3) given its location within the type I cell (Yamamoto et
16 al., 2006). Interestingly, mice with reduced NOS-3 have a dampened hypoxic ventilatory
17 response and a depressed CB function (Kline et al., 2000). One explanation for this is an
18 adaptation to chronic hypoxia brought about by reduced CB blood flow. However, this is
19 unlikely as there is no significant type I cell hyperplasia/hypertrophy (McGregor et al., 1984;
20 Tatsumi et al., 1991). Instead, the blunted CB activity could be due to the lack of NO acting on
21 the CB mitochondria. Consideration of the precise compartmentalization of NO should also be
22 taken into account. Whilst NO in the mitochondria induces chemostimulation, its action in other
23 regions is likely to have opposing effects via modulation of soluble guanylate cyclase and L-
24 type Ca²⁺/BK_{Ca} channels (Summers et al., 1999; Iturriaga et al., 2000; Silva and Lewis, 2002;
25 Valdes et al., 2003). In addition, other diffusible cytosolic factors have been implicated in CB
26 O₂ sensing including H₂S and CO (Peng et al., 2010; Yuan et al., 2015). Both of these gases are
27 capable of inhibiting type I cell mitochondria (Wilson et al., 1994; Lahiri et al., 1999; Buckler,
28 2011). Future experiments are required to evaluate if these substances act by setting type I cell
29 mitochondrial O₂ sensitivity.

30
31 A second explanation for the low O₂-affinity of the type I cell mitochondria is that it has a
32 unique gene expression profile (Figure 2). Exploring gene expression in the CB is challenging
33 due to its relatively small size and heterogeneity. However, advances in molecular biology
34 techniques now make it possible to perform whole genome analysis using just micrograms of
35 tissue or even single cells. RNA-sequencing analysis has now revealed a number of
36 mitochondrial related genes that have a particularly high expression in the type I cell (Zhou et
37 al., 2016). Of these *Ndufa4l2* and *Cox4i2* have been shown to be more highly expressed in the
38 CB compared with tissue from the superior cervical ganglion (Gao et al., 2017). Whether these
39 two genes contribute to the low mitochondrial O₂ affinity remains to be determined but these
40 findings do support the idea that CB mitochondria have a unique genetic signature encoding
41 their mitochondrial complexes. We would expect many more genetic studies to probe this
42 further until the type I cell mitochondria can be accurately modelled to pinpoint the structural
43 conformation underlying its low O₂ affinity. An interesting comparator may be the
44 mitochondria isolated from guinea pig CB which does not appear to have any inherent O₂
45 sensitivity (Gonzalez-Obeso et al., 2017).

46

47 **7 Conclusion**

48 On current evidence, it is very hard to disprove the mitochondrial hypothesis of CB O₂ sensing.
49 The mitochondria seem to fulfil all five criteria that we have proposed for adequate O₂-sensors.
50 What is less clear is a mechanistic understanding of how the low O₂ sensitivity of the CB

1 mitochondria is achieved and if mitochondria are involved in establishing pathological changes
2 in CB function.

3

4

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8

1 **Figure Legends**

2

3 **Figure 1: Carotid body mitochondrial signaling mechanisms activated during hypoxia.**

4 Hypoxia-induced mitochondrial inhibition is proposed to increase lactate generation (Chang et
5 al., 2015), augment mitochondrial complex I reactive oxygen species (ROS) production
6 (Fernandez-Aguera et al., 2015) or reduce mitochondrial ATP synthesis (Buckler and Turner,
7 2013). These changes are proposed to directly or indirectly (e.g. via Olf78 receptor activation,
8 reduced MgATP concentration or stimulation of AMP-activated protein kinase; AMPK) modify
9 ion channel function leading to resting membrane potential (RMP) depolarization. This causes
10 opening of L-type Ca²⁺ channels, neurosecretion and an increase in discharge frequency of the
11 adjacent chemoafferent fibers. TRP- transient receptor potential channel; NSC- non-selective
12 cation channel; BK_{Ca}- large conductance Ca²⁺-activated K⁺ channel; TASK-Twik-related acid-
13 sensitive K⁺ channel; TREK- TWIK-related K⁺ channel; AP-action potential.

14

15 **Figure 2: Potential mechanisms underlying the unique low O₂ affinity mitochondria in the**

16 **carotid body.** Inhibition of mitochondrial function in the carotid body type I cell occurs at PO₂
17 values well above those that inhibit metabolism in other O₂ insensitive cells. This activates a
18 number of proposed signaling pathways (shown in blue). The unique low O₂ affinity of the
19 carotid body mitochondria could be due to 1. a unique mitochondrial gene expression profile or
20 2. the presence of a competitive inhibitor such as NO (Holmes et al., 2016). The impact of
21 lower mitochondrial O₂ affinity is to cause a right-shift in the PO₂-chemoafferent response
22 curve, thereby enhancing the functional O₂ sensitivity and allowing the carotid body to respond
23 over a more physiological range of arterial PO₂s. RET-reverse electron transport.

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29 NA

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31 **Author contributions**

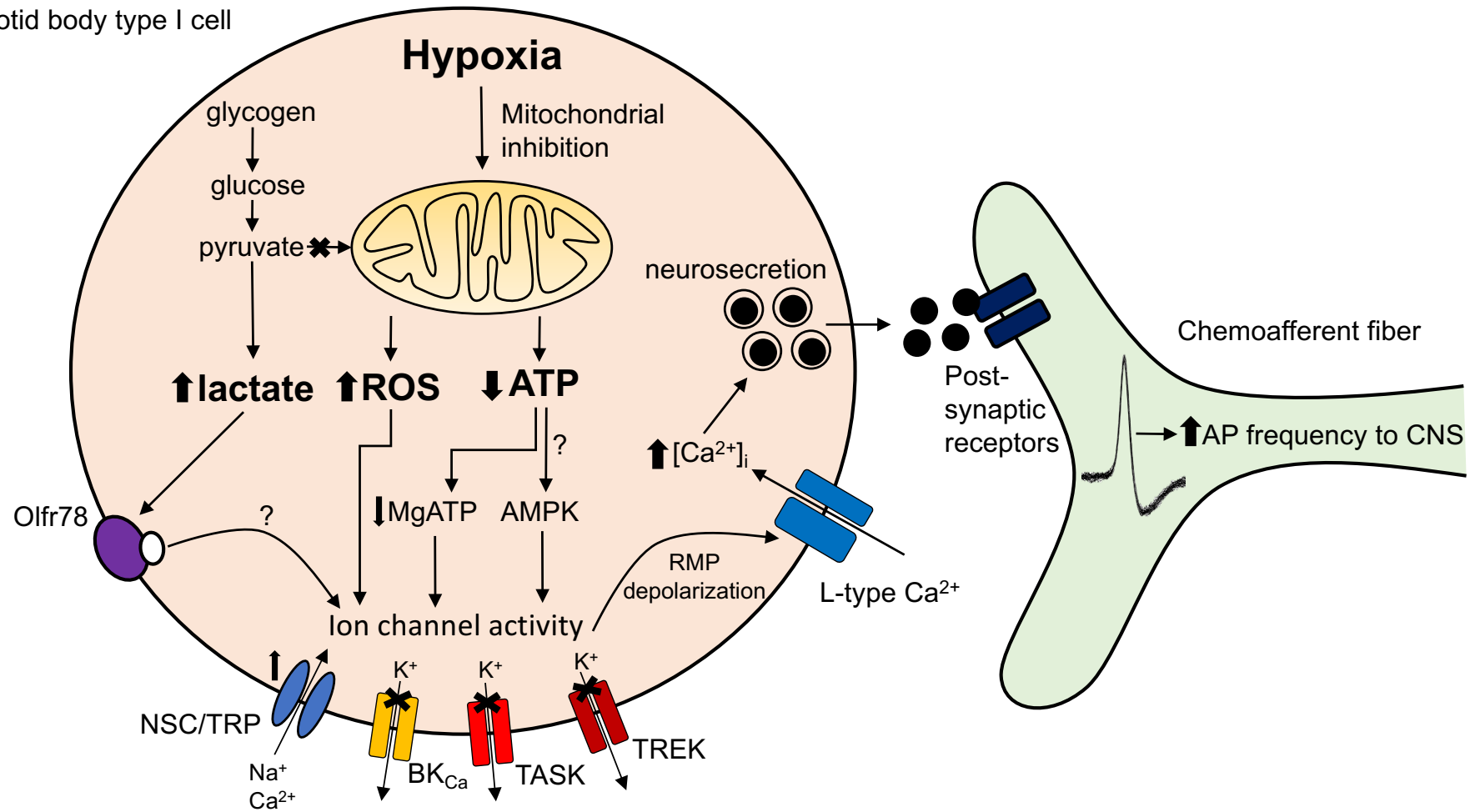
32 APH, CJR, AMC and PK all contributed to the writing and editing of the manuscript.

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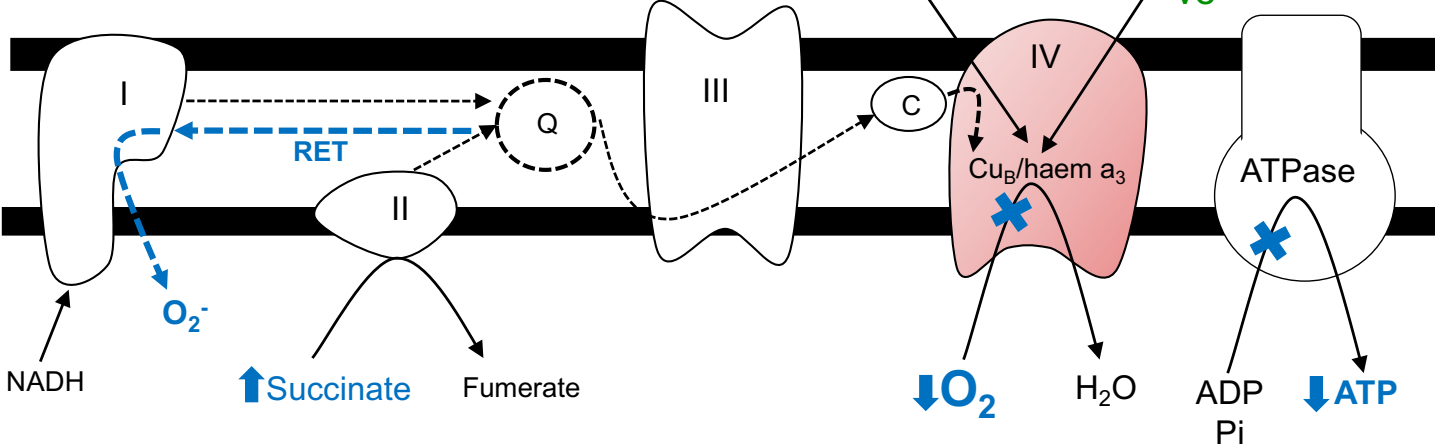
Carotid body type I cell



1. Unique 'low O₂ affinity' mitochondrial gene expression profile

2. Presence of a competitive inhibitor e.g. NO

Type I cell mitochondrial electron transport chain



Whole carotid body chemoafferent O₂ sensitivity

