# UNIVERSITY<sup>OF</sup> BIRMINGHAM University of Birmingham Research at Birmingham

# Lessons to cancer from studies of leukemia and hematopoiesis

Brown, Geoff

DOI: 10.3389/fcell.2022.993915

License: Creative Commons: Attribution (CC BY)

Document Version Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Brown, G 2022, 'Lessons to cancer from studies of leukemia and hematopoiesis', Frontiers in cell and developmental biology, vol. 10, 993915. https://doi.org/10.3389/fcell.2022.993915

Link to publication on Research at Birmingham portal

#### **General rights**

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

•Users may freely distribute the URL that is used to identify this publication.

•Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.

•User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?) •Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

#### Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

#### ( Check for updates

#### OPEN ACCESS

EDITED BY Shyamala Maheswaran, Massachusetts General Hospital, Harvard Medical School, United States

REVIEWED BY Neslihan Meriç, Istanbul Health and Technology University, Turkey Yiran Li, Tongji University, China

\*CORRESPONDENCE Geoffrey Brown, g.brown@bham.ac.uk

SPECIALTY SECTION

This article was submitted to Cancer Cell Biology, a section of the journal Frontiers in Cell and Developmental Biology

RECEIVED 14 July 2022 ACCEPTED 05 September 2022 PUBLISHED 20 September 2022

#### CITATION

Brown G (2022), Lessons to cancer from studies of leukemia and hematopoiesis. *Front. Cell Dev. Biol.* 10:993915. doi: 10.3389/fcell.2022.993915

#### COPYRIGHT

© 2022 Brown. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Lessons to cancer from studies of leukemia and hematopoiesis

### Geoffrey Brown\*

School of Biomedical Sciences, Institute of Clinical Sciences, College of Medical and Dental Sciences, University of Birmingham, Birmingham, United Kingdom

The starting point to describing the origin and nature of any cancer must be knowledge about how the normal counterpart tissue develops. New principles to the nature of hematopoietic stem cells have arisen in recent years. In particular, hematopoietic stem cells can "choose" a cell lineage directly from a spectrum of the end-cell options, and are, therefore, a heterogeneous population of lineage affiliated/biased cells. These cells remain versatile because the developmental trajectories of hematopoietic stem and progenitor cells are broad. From studies of human acute myeloid leukemia, leukemia is also a hierarchy of maturing or partially maturing cells that are sustained by leukemia stem cells at the apex. This cellular hierarchy model has been extended to a wide variety of human solid tumors, by the identification of cancer stem cells, and is termed the cancer stem cell model. At least, two genomic insults are needed for cancer, as seen from studies of human childhood acute lymphoblastic leukemia. There are signature mutations for some leukemia's and some relate to a transcription factor that guides the cell lineage of developing hematopoietic stem/progenitor cells. Similarly, some oncogenes restrict the fate of leukemia stem cells and their offspring to a single maturation pathway. In this case, a loss of intrinsic stem cell versatility seems to be a property of leukemia stem cells. To provide more effective cures for leukemia, there is the need to find ways to eliminate leukemia stem cells.

#### KEYWORDS

leukemia, leukemia stem cells, cancer stem cells, oncogenes, therapies

### Introduction

There are very many different types of leukemia. Since the late 19th century, their diagnosis and subtyping were underpinned by the leukemia cells' appearance, including their relationship to a cell lineage, maturation status, and the rapidity of proliferation. Based on these features, the four major subtypes of leukemia are chronic lymphocytic leukemia (CLL), chronic myeloid leukemia (CML), acute lymphoblastic leukemia (ALL), and acute myeloid leukemia (AML). The acute leukemias were subdivided from examination of a combination of morphologic, immunotypic, and cytogenetic features. For example, the French-American-British classification of AML recognized eight different subtypes, designated as M0 through to M7. In 2008, the WHO updated the classification of the malignancies of hematopoietic and lymphoid tissues incorporating layer upon layer of complexity, including morphogenic, immunotypic, cytogenetic, and

molecular genetics findings. The WHO guide to clinicians listed 94 disease entities (Swerdlow et al., 2008).

A precise diagram for how each blood cell type develops from HSCs is central to resolving how this is distorted in the different leukemias. Hematopoietic stem cells (HSCs) sit at the apex of blood cell development, and for over 30 years they were regarded as a uniform cohort of cells that is uniquely able to self-replicate indefinitely. In classic models, HSC developmental progression is depicted in a branching tree-like manner (Spangrude et al., 1988). Lineage options are restricted stepwise via a series of binary decisions and compartmentalized intermediate hematopoietic progenitor cells (HPCs) that ultimately give rise to lineage-restricted HPCs. These cells have a lesser ability to selfrenew. In addition to undergoing expansion, lineage-restricted HPCs differentiate further to give rise to mature blood cells that are functional and predominantly short-lived. (Kondo et al., 2003).

From around 2009, new principles to the nature of HSCs arose leading to continuum (Ceredig et al., 2009) and diffusion map (Nestorowa et al., 2016) models for hematopoiesis. In these models, HSCs "choose" a cell lineage directly from a spectrum of end-cell options; they are, therefore, a consortium of HSC subtypes with distinct characteristics. Known subsets include HSCs that are megakaryocyte (Sanjuan-Pla et al., 2013), lymphoid-, myeloid- and dendritic cell lineage-biased (Challen et al., 2010; Gekas and Graf, 2013; Shimazu et al., 2012; Naik et al., 2013), and erythroid and macrophage affiliated (Mooney et al., 2017). The identification of these lineage-biased/affiliated cells is important because they, like multipotent HSCs, are primarily at risk of leukemogenesis by virtue of their ability to self-renew. HPCs normally undergo a limited number of cell divisions. There are not findings to support the view that an oncogene can endow a committed HPC with the capacity to selfrenew which is required to sustain leukemia. In continuum and diffusion map models, lineage imprinting occurs earlier than previously thought, and the differentiation of HSCs and their downstream HPCs is a progressive process with broad developmental trajectories (Nestorowa et al., 2016). This allows cells to move to the left or right of a chosen pathway (Ceredig et al., 2009; Nestorowa et al., 2016); in other words, HSCs and HPCs having chosen a pathway remain versatile (Brown and Ceredig, 2019). Therefore, an important consideration in the transformation of multipotent HSCs is whether the transformed cell remains versatile to generate leukemia cells belonging to several cell lineages.

In this review, I focus on how a clearer picture of the involvement of HSCs in leukemogenesis has emerged from advances in our understanding of hematopoiesis, the influence of oncogenes on HSCs, and that the human leukemias are sustained by leukemia stem cells (LSCs) at their apex. In this regard, the leukemic cell hierarchy is similar to that of normal hematopoiesis. However, unlike normal stem cells, the progeny of LSCs seems to be restricted to a cell lineage. The principle of LSCs extends to solid cancers from the identification of cancer stem cells (CSCs), and consideration is given to finding ways to eliminate these cells.

# The stem cell origin of some leukemias

CML and some AMLs originate in an HSC. In 1978, the expression of the glucose-6-phosphate dehydrogenase (G-6-PD) isoenzymes A and B were examined for cells from three women with CML. From X chromosome inactivation, only one G-6-PD gene is active in each female cell which is a mixture of cells that express either the A or B isoenzyme. One enzyme type was found in the blood granulocytes, erythrocytes, platelets, monocytes/ macrophages, and B lymphocytes of the patients. The investigators concluded that the clonal origin of CML is a multipotent HSC, and coined the term LSC. Their findings also revealed that the above blood cell types arise from a common HSC (Fialkow et al., 1978). For the subtypes of AML analyzed in 1997, the investigators concluded that they had arisen from a primitive hematopoietic cell. The cells that initiated human AML in non-obese diabetic-severe combined immunodeficient (NOD/SCID) mice were CD34++, CD38- and able to self-renew, which are attributes of a normal HSC (Bonnet and Dick, 1997).

The advent of panels of monoclonal antibodies facilitated and strengthened the sub-typing of the leukemias regarding their affiliation to a cell lineage. Some leukemias were assigned as arising in a committed HPC from the well-described phenotypes of normal HPC sub-populations. However, further analyses of leukemias that were thought to originate in a committed HPC have revealed a more primitive origin. In acute promyelocytic leukemia (APL), there is a differentiation block at a promyeloid stage of myeloid differentiation, leading to an abundance of promyelocytic blasts. The hallmark of APL is the chromosomal translocation t (Edwards et al., 1999; Adolfsson et al., 2005) which gives rise to the fusion protein PML/RARa. The presence of translocation and fusion protein has not been reported in any other malignancy, and the protein plays a crucial role in APL leukemogenesis. The translocation and oncoprotein were observed to be present in CD34<sup>+</sup> CD38<sup>-</sup> HSC-like cells from patients with APL, supporting a disease origin in an HSC rather than a committed myeloid HPC (Edwards et al., 1999). Pro-B cells accumulate in the bone marrow of infant t (Ceredig et al., 2009; Mooney et al., 2017) MLL-AT4 B-cell precursor ALL. A new view is that the cell of origin is a fetal liver lymphoid-primed multipotent progenitor (LMPP) (Malouf and Ottersbach, 2018). These stem-like cells generate neutrophils, monocytes, B cells, and T cells, but do not produce significant erythroid cells and megakaryocytes (Adolfsson et al., 2005). The cells in childhood B-ALL express the early stage B-cell antigen CD19 and that a B-cell committed progenitor is the cell of origin is a longstanding

notion (Greaves, 1981). There are findings to support a more primitive HSC-like cell as the cell of origin. For children with B-ALL, cytogenetically aberrant cells are present in the CD34<sup>+</sup>, CD33<sup>-</sup>, CD38<sup>-</sup>, CD19<sup>-</sup> primitive cell compartment in the bone marrow (BM), and engraftment of B-ALL into NOD/SCID mice was achieved with CD34<sup>+</sup>, CD10<sup>-</sup> cells or CD34<sup>+</sup>, CD19<sup>-</sup> cells (Quijano et al., 1997; Cox and Blair, 2005; Hirt et al., 2011). A surprising finding was that transformation of a self-renewing HSC gives rise to CLL, as opposed to the CLL subtypes arising from the transformation of an antigen-experienced B-cell (Alizadeh and Majeti, 2011; Kikushige et al., 2011). A striking feature of the above LSCs is that they, unlike normal HSCs, have given rise to offspring belonging to just one cell lineage. There is excessive production of neutrophils in CML, promyelocytic blasts predominate in APL, and B-cell lineage cells prevail in infant B-cell precursor ALL, childhood ALL, and CLL.

### The cancer stem cell model

The CSC model of cancer captures the importance of stem cells to the genesis of cancer. The proposal arose from studies of AML and, as mentioned above, the NOD/SCID leukemiainitiating cells were rare, on average 1 per 10<sup>6</sup> (Dick, 2008). These are LSCs that sustain a hierarchy of maturing or partially maturing leukemia cells. This cellular hierarchy model was extended to solid cancers, and universality is important because CSCs are also largely responsible for disease relapse and metastasis. Studies of different human solid cancers have led to the conclusion that the cells that give rise to solid tumors when transplanted into NOD/SCID mice are also often quite rare (0.1%-0.0001%). For breast cancer, CSCs are a small fraction of tumor cells and are viewed as seeding the tumor. The population of breast cancer CSCs is heterogeneous; the two subgroups, namely a mesenchymal and quiescent type and an epithelial and proliferative type, are two dynamic states of breast cancer CSCs (Zhou et al., 2019). For colorectal cancer, CSCs are central to tumor initiation because when isolated from primary tumors they give rise to tumors in mice. The development of a better understanding of biomarkers is assisting the clarification of colorectal CSC phenotypes (Munro et al., 2018). CSCs have been identified and isolated for other solid cancers, including bladder (She et al., 2008), head and neck squamous cell carcinoma (Prince et al., 2007), lung (Eramo et al., 2008), pancreatic (Hermann et al., 2007), prostate (Collins et al., 2005), and sarcoma (Wu et al., 2007). For these diseases, CSCs play a role in disease initiation and metastasis.

A low frequency of CSCs is not always the case because around 27% of unselected melanoma cells from four patients gave rise to a tumor in single cell transplants when a modified xenotransplantation assay was used. The investigators concluded that melanoma follows the principle of the cancer stem cell model, and that CSCs are common in some tumors (Quintana et al., 2008). For some solid tumors there is still a lack of information about the frequency of CSCs. For example, glioblastoma is the most malignant primary brain tumor. It contains CSCs that have the ability to generate neurospheres and that are viewed as contributing to tumor initiation and therapeutic resistance. The frequency of CSCs within glioblastoma is uncertain because the linkage of markers between normal stem cells and CSCs is still controversial (Lathia et al., 2015).

# At least two "hits" are needed for leukemia

Studies in 1983 showed that at least two genomic insults are needed to transform primary embryo fibroblasts so that these cells give rise to large tumors in mice. Cooperating oncogenes included, for example, activation of a *RAS*-like and a *MYC*-like gene. (Land et al., 1983). Of importance was whether the same is true for cancer in humans, and, at least, two insults are needed as seen for the genesis of leukemia in children.

Twins with concordant leukemia provided a unique opportunity to unravel the etiology of leukemia's. Embryos that split after day 3 and before day 7 develop as separate embryos with a single (monochromic) placenta. This is the case for 60% of monozygotic twins and there is blood cell chimerism, via vascular anastomosis. Some years ago, it was suggested that concordant leukemia in twins has a common, and often prenatal, origin. In this case, leukemia is initiated in one of the twins, and the progeny of the clone transfers to the other twin via a shared placental (Wolman, 1962). A number of studies of concordant leukemia have provided evidence to support the sharing of a leukemia initiating clone from one twin to the other (reviewed in 37) An identical DNA rearrangement to the immunoglobulin heavy chain was seen to be shared by B-cell lineage ALL cells from a pair of infant Siamese twin boys who were separated 42 days after birth and both diagnosed with ALL 7 months later. The investigators concluded for twins diagnosed in the first 1-2 years of life with leukemia that a single neoplastic transformation had occurred in utero, rather than there was a strong genetic susceptibility (Pombode de Oliveira et al., 1986). Cytogenetic studies in 1998 showed that the leukemia cells from pediatric monozygotic twins with concordant AML had identical clones by virtue of the shared karyotype +8, inv16, and +21. The twins developed AML at 3 and 4 years. At the time of the bone marrow transplant of the first twin, the abnormal clone was seen to be present in the asymptomatic twin. This finding again pointed to in utero twin-twin transfusion, and that a preleukemic clone had evolved years before the onset of overt disease. The investigators proposed that information about an acquired genetic change(s) in one twin may be useful to assessing disease in an asymptomatic twin (Richkind et al., 1998).

Molecular analyses of twins with concordant leukemia have substantiated sharing of a leukemic clone from one twin to the other. Eighty percent of cases of infant ALL (<1 year of age) have a reciprocal chromosome translocation involving the MLL gene (at 11q23). The MLL gene fuses principally with the AF4 gene, and various other genes. Southern blots of MLL, with multiple enzyme digests, revealed that the ALL cells from three pairs of infant identical twins had identical restriction fragments. The restriction fragments from the DNA of a pair of twins comigrated, and the migration patterns were different between the pairs of twins and for the controls. This study also showed that the MLL rearrangement had occurred in utero (Ford et al., 1993). Twenty-five percent of patients with childhood B-cell precursor ALL have the chromosome translocation t (12; 21) resulting in the TEL(ETV6)-AML1(RUNX1) fusion gene. ALL cells from twins with concordant leukemia were characterized by cloning (Ford et al., 1998) or the use of the polymerase chain method and sequencing (Wiemels et al., 1999a; Wiemels et al., 1999b; Maia et al., 2001). The latter method was used to track leukemogenesis in a triplets pregnancy, and two of the three triplets developed ALL (Maia et al., 2001). The same TEL and AML1 breakpoints were observed to be present within the ALL cells for each of four pairs of twins.

There was often a latency to the development of TEL-AML1 ALL in one of the twins. In one study, monozygotic twins were diagnosed with TEL-AML1 ALL at 3 years, 6 months and 4 years, 10 months (Ford et al., 1998). The latency can be very protracted as in a different study the diagnoses of TEL-AML1 ALL were at ages 5 and 14 (Wiemels et al., 1999a). Examination of an archived slide of BM cells from the twin who developed leukemia later showed that the TEL-AML1 fusion sequence was present in the archived marrow. At the time, the marrow was haematologically normal. The TEL-AML fusion gene rearrangement has, therefore, a fetal origin. Moreover, the most plausible explanation for the above findings is that a leukemia-initiating mutation had occurred in utero, this clone was shared between the children, and a second, or more, insult(s) was needed for full-blown childhood ALL (Ford et al., 1998). Evidence from epidemiological studies of older children with TEL-AML1-positive or hyperdiploid ALL favours an abnormal response to infection playing a causal role regarding the second insult (Greaves et al., 2003). The rationale is that the naïve immune network is modulated by exposure to common infections, and a lack of exposure in the first year of life leads to a risk of leukemia by virtue of a dysregulated immune response. It may then be possible to develop a prophylactic to prevent ALL in children.

# The pre-leukemia stem cells arising from hematopoietic stem cells

HSCs may give rise to many leukemias and the two-hit model of leukemogenesis states that an HSC with a stable and preleukemia lesion must only progress to full-blown leukemia after a second hit. A pre-LSC clone having received just one insult is likely, therefore, to be able to undergo normal hematopoiesis. There is good evidence to support this view. Five percent of healthy newborns have a *TEL-AML1* fusion, as shown by DNAbased GIPFEL screening. This is compatible with normal hematopoietic development in the majority of the cases because ninety-nine percent of newborns will never develop B-cell precursor ALL (Schafer et al., 2018). Therefore, the TEL-AML1 transcription factor has a very low oncogenic potential, which strengthens the importance of the need for a second hit that is either spontaneous or environmental.

As mentioned above, blood granulocytes, erythrocytes, platelets, monocytes/macrophages, and B lymphocytes are all part of the BCR-ABLp210+ pre-LSC and HSC-derived CML clone with pre-LSCs differentiating into mature blood cells. AML1-ETO fusion transcripts are detectable in most patients with t (8; 21) AML, and were also found in various colonyforming cells, including mixed (CFU-Mix), erythroid (BFU-E), granulocyte-macrophage (CFU-GM), and megakaryocyte (CFU-Meg), when BM cells were examined for patients who had been in remission for 12-150 months (Miyamoto et al., 1996). Later studies showed that transcripts were detectable within HSCs, monocytes, and a fraction of B cells, but not T cells, and again in up to 60% of CFU-Mix BFU-E, CFU-GM, and CFU-Meg. The morphologies and compositions of the AML1-ETO1+ and AML1-ETO - myeloid colonies were the same. (Miyamoto et al., 2000). Hence, the t(8; 21) acquisition had occurred at the level of an HSC, and these cells can differentiate into various myeloid and B-cells. The investigators concluded that AML1-ETO is an early step in the process that leads to transformation and that a further insult might involve the machinery for myeloid lineage development.

### Signature mutations in leukemia

As considered above, analyses of genetic abnormalities have greatly assisted the sub-typing of leukemias. Moreover, whole genome sequencing has now transformed our understanding of the various types and the pathogenesis of hematopoietic malignancies. For example, the Pediatric Cancer Genome Project looked to sequence 600 pediatric cancers and matched germline cancers (Downing et al., 2012). Aims were to fully explore the mutations that drive cancer and their frequency within specific cancers. As to frequency and from earlier studies of leukemias, some types of leukemia are linked to a specific oncogenic event (Table 1). The overt leukemia cells TABLE 1 Signature mutations in leukemia.

Leukemia	Oncogenic event	Frequency
Lymphoid		
Infant B-cell ALL	MLL-AF4 fusion gene	80%
Childhood B-cell ALL	TEL-AML1 fusion gene	25%
Adult B-cell ALL	PAX5 mutations	34%
T-cell leukemia	LMO2 gene activation	Typifies
Extranodal MALT lymphomas	MLT/MALT1 involvement	26%
Myeloid		
APL	PML-RARA fusion gene	Typifies
M1, M2, and M4 AML	CEBPA mutations	7%-20%
Adult AML	NPM1 mutations	25%-41%
t (8; 21) AML	AML1-ETO fusion gene	Typifies
CML	BCR-ABLp210 oncoprotein	Typifies
Ph+ precursor B-cell ALL	BCR-ABLp190 oncoprotein	Typifies

closely resemble a normal differentiated or partially differentiated cell of a particular lineage. One interpretation of these findings is that the signature mutation plays a role in specifying the lineage fate of the LSCs and their offspring.

Some of the genes that are involved in the genetic abnormalities shown in Table 1 encode transcription factors (TFs) that are important to HSC development towards an end cell type. The PAX5 gene is frequently mutated in human B-cell leukemia, and encodes a transcription factor that regulates the early stages of B-cell development (Fuxa and Skok, 2007). The LM02 gene encodes a TF that is activated by chromosomal translocations in T-cell leukemia. It is expressed within immature CD4/CD8 double negative thymocytes but does not have an obligatory role in T-cell development, as revealed by studies of knockout mice (McCromak et al., 2003). However, the LMO2 TF may still have a regulatory role. The PML-RARA fusion gene typifies APL and the RARA gene encodes the TF retinoic acid receptor (RAR) a. Like LMO2, RARa does not have an obligatory role in myelopoiesis but is a key regulator of the differentiation of granulocyte-macrophage progenitor cells (Kastner and Chan, 2001). CEBPA is mutated in M1, M2, and M4 AML, and encodes the TF C/EBPa which is a regulatory switch that induces granulocyte lineage commitment in CFU-GM (Radomska et al., 1998).

The rest of the genetic abnormalities shown in Table 1 have an indirect influence on gene transcription. The main player in mucosa-associated lymphoid tissue (MALT) lymphomas is MALT1. MALTI is a paracaspase and a complex of BCL10, CAMA1, and MALT1 activates the transcription factor NF- $\kappa$ B, *via* IKK $\gamma$  degradation (Bertoni and Zucca, 2006). The AML1-ETO protein is viewed as regulating gene expression *via* epigenetic mechanisms. The fusion protein recruits multiple chromatin-modifying enzymes to target genes, to alter chromatin marks and transcription factor binding (Rejeski et al., 2021). The *MLL1* gene that is rearranged in infant B-cell ALL encodes the histone lysine-specific methyltransferase 2A which plays a role in gene expression (Winters and Bernt, 2017). Mutation of nucleophosmin (NPM1) is a common driver in adult AML. NPM1 is present in a small region inside the nucleus and is a multifunctional protein that has roles in the maintenance of genomic stability and chromatin remodeling, the biogenesis of ribosomes, the p53dependent stress response, and the modulation of pathways that suppress cell growth (Box et al., 2016; Heath et al., 2017).

## Oncogene-mediated setting of hematopoietic stem cell fate

In keeping with the link between signature mutations and TFs that play a role during hematopoietic cell development along a pathway, some oncogenes can set the cell lineage of HSCs/ HPCs and their offspring. This was revealed by targeting the expression of particular oncogenes to HSCs/HPCs in transgenic mice (Table 2).

As mentioned above, *LMO2* is expressed within immature thymocytes in the thymus, with aberrant expression in these cells leading to T-ALL (Chambers and Rabbitts, 2015). LMO2 expression was initiated and maintained in only HSCs/ HPCs by placing expression under the control of the stem cells antigen-1 (Sca1) promoter. These mice developed an aggressive T-cell ALL. Within HSCs/HPCs, the *LM O 2* oncogene, therefore, imposes aggressive T-cell ALL. T-cell ALL also developed when LMO2 expression was targeted, *via* appropriate Cre-mediated activation, to pro-B and germinal center B-cells. *LMO2* oncogene expression had, therefore, reprogrammed the B-cell lineage cells to favor the initiation of T-cell ALL and/or imposed the characteristics of T-cell ALL (Garcia-Ramirez et al., 2018).

Oncogenic event(s)	Targeted cells	Transgenic mice outcome
LMO2 gene activation	HSCs/HPCs, B-cells	T-cell ALL
BCR-ABLp210 fusion	HSCs/HPCs	CML
BCR-ABLp190 fusion	HSCs/HPCs	Precursor B-cell ALL at 13%
BCR-ABLp190 & PAX5 loss	HSCs/HPCs	Precursor B-cell ALL at 90%
TEL/AML1 fusion	HSCs/HPCs	B- and T-cell leukemias
TEL/AML1 fusion & KDM5C loss	HSCs/HPCs	B-ALL at 22%

TABLE 2 Targeting of oncogenic events in transgenic mice and the leukemia outcomes.

LMO2, when transfected with five other TFs (RUNX1T1, HLF, PRDM5, PBX1, and ZFP37), has also been shown to impart multilineage potential onto committed myeloid and lymphoid HPCs and myeloid effector cells (Riddell et al., 2014). To allow the LMO2 reprogramming of B-cell lineage, and other cells, to take place the cells must be inherently plastic. This is in keeping with the above view on the versatility of normal HSCs/HPCs.

BCR-ABLp210 was targeted to HSCs/HPCs in transgenic mice via Sca1-BCR-ABLp210. The mice developed leukemia resembling human CML (Perez-Caro et al., 2009), revealing that this oncogene alone can impose CML within HSCs/ HPCs. In this case, BCR-ABLp210 seems to be less reliant on a cooperating secondary hit. DNA methylation at CpG islands was examined for HSCs/HPCs from the Sca1-BCR-ABLp210 transgenic and the wild-type mice as to whether there had been a change to the epigenetic landscape. DNA hypomethylation was observed for the HSCs/HPCs from the Sca1-BCR-ABLp210 mice. This was a lasting change because hypomethylation was also observed for the mature leukemic myeloid cells from the Sca1-BCR-ABLp210 mice, despite an absence of oncogene expression in the mature cells. DNA methyltransferase 1 (Dnmt1) was upregulated within HSCs/ HPCs from the Sca1-BCR-ABLp210 mice. Expression of Dnmt1 in HSCs/HPCs, under control of the endogenous Sca1 promoter, led to DNA hypomethylation that was similar to that observed for the cells from the Sca1-BCR-ABLp210 mice. The Sca1-Dnmt1 transgenic mice developed malignancies that were mostly myeloid with a marked expansion of granulocytes in the bone marrow and blood. The investigators concluded that BCR-ABLp210 had reprogrammed the epigenome of HSCs/HPCs to lead to CML (Vicente-Duenas et al., 2019).

In contrast to the influence of *BCR-ABLp210*, targeting of *BCR-ABLp190* to HSCs/HPCs in transgenic mice, *via Sca1-BCR-ABLp190*, led to precursor B-cell ALL resembling the human disease. In an aging mouse colony, the penetrance was only 13%. In transgenic mice that were double *Sca1-BCR-ABLp190* and *Pax5<sup>+/-</sup>* (a B-cell TF), precursor B-cell ALL developed with much shorter latency and a 90% incidence, and there were alterations in the remaining wild-type *Pax5* allele. Therefore, *BCR-ABLp190* together with the loss of *Pax5* can confer the changes that are needed for precursor B-cell ALL, whereby *Sca1-BCR-ABLp190* 

has set the cell lineage of HSCs/HPCs. Regarding the role of Pax5, metabolic genes were upregulated and glucose uptake and energy metabolism were increased within the pro-B cells from the *Pax5* deficient mice (Martin-Lorenzo et al., 2018).

As considered above, the TEL/AML1 fusion gene confers a low risk of developing childhood B-ALL, and an abnormal reaction to infection has been postulated to trigger the overt disease. Transgenic mice failed to develop B-ALL when TEL/ AML1 expression was targeted to pro-B cells and the mice were exposed to natural infections. T-ALL and B-ALL were triggered when TEL/AML1 expression was initiated in HSCs/HPCs and the mice exposed to infections. The H3K4me3 and H3K4me2 demethylase KDM5C is missense mutated in mouse TEL/AML1 B-ALL and human relapse TEL/AML1 B-ALL (Rodriguez-Hernandez et al., 2017). For mice that expressed TEL/AML1, introducing a loss of KDM5C into the B-cell compartment did not give rise to B-ALL. By contrast, loss of KDM5C in HSCs/HPCs that expressed TEL/AML1 led to the development of B-ALL in mice (22%) that were kept in a special pathogen-free environment. Therefore, TEL/AML1 can trigger T-cell and B-cell leukemias, and the second "hit" determines the lineage fate of TEL/AML1+ leukemia, with both "hits" having to occur in an HSC/early HPC (Rodriguez-Hernandez et al., 2021).

Different mutant RUNX1 oncoproteins have been shown to program alternative trajectories for hematopoietic cell differentiation. Four types of RUNX1 oncoproteins were assessed by their induced expression in embryonic stem cells. RUNX1-ETO reduced the expression and binding of PU.1 and C/EBPa (both TFs regulate myeloid differentiation) and led to a bias towards a B cell identity. Expression of R2010, which has a mutation in the DNA binding domain, reduced the interaction of wild-type RUNX1 with CBFB (a master regulator of hematopoiesis) and increased GATA1 binding (a TF for erythropoiesis), leading to a bias away from megakaryocyte differentiation. RUNX1-EV11 reduced differentiation towards myeloid and erythroid cells (Kellaway et al., 2021). RUNX1 plays a role in the organization of the chromatin landscape for hematopoiesis (Lichtinger et al., 2012), and expression of the mutant proteins perturbed chromatin priming of lineage-specific sites, in addition to the regulatory TF network (Kellaway et al., 2021).

For the leukemias in the transgenic mice, LSCs appear to have been programmed by the "hit" to adopt a particular maturation pathway. For BCR-ABLp210 and RUNX1, investigators have shown that there was disruption to the epigenome/chromatin priming within stem cells, and the demethylase KDM5C is missense mutated in human relapse TEL/AML1 B-ALL. Dysregulation of epigenetics is viewed as an overarching hallmark of cancer. Therefore, it is perhaps not too surprising that a "hit" dysregulates the epigenetics to cell lineage commitment. A second "hit" might include lesions in genes such as MYC and RAS that highjack the control on cell proliferation leading to hyper-proliferation, and/or in genes that, for example, lead to loss of p53 function and BCL2 overexpression leading to inhibition of apoptosis. These lesions are forces in driving diverse human cancers and, for example, MYC and RAS are viral oncogenes within the highly oncogenic retroviruses.

# Conventional therapies spare leukemia stem cells

Chemotherapy and radiotherapy target cells that are cycling and are often very effective against the bulk of cancer cells. An important finding, in 1999, was that when quiescent LSCs were isolated from patients with CML they were found to be insensitive to high doses of chemotherapy (Holyoake et al., 1999). Second-generation drugs, e.g., dasatinib, nilotinib, and bosutinib, that inhibit the tyrosine activity of the BCR-ABLp220 protein are the main line treatment for CML and are used also to maintain remission. CML-LSCs are resistant to tyrosine kinase inhibitors, despite the expression of BCR-ABLp220 protein (Graham et al., 2002; Corbin et al., 2011). Tyrosine kinase inhibitors are also used to treat other cancers, including breast, kidney, liver, lung, pancreatic, prostate, and thyroid. They have been heralded as a breakthrough targeted therapy, as to having also a low toxicity profile even in older patients. Even so, the effectiveness of tyrosine kinase inhibition has been limited by the emergence of resistant clones (Pottier et al., 2020), and the second-generation tyrosine kinase inhibitors have only partially overcome the resistance conferred by the T3151 mutation.

It becoming clear that CSCs are resistant to conventional therapies. For example, the most common cancer globally is lung cancer and lung CSCs, which are responsible for relapse posttreatment, are resistant to many of the conventional therapies (Prbavathy et al., 2018). Similarly, the existence of a small population of breast CSCs is well proven, and these cells are thought to contribute to metastatic lesions. Breast CSCs cycle slowly, and there is accumulating evidence that they are the leading cause of resistance to conventional treatments. Various intracellular events govern CSC drug resistance and miRNAs have been proposed as playing a central role in governing the behavior of breast CSCs (Song and Farzaneh, 2012).

# How might we eliminate cancer stem cells?

The identification of LSCs, and subsequently of CSCs, has steered investigators towards searching for the means to eliminate these cells. The anticipated benefits would be the prevention of disease relapse and metastasis, and perhaps the provision of a bono fide cure for aggressive cancers that are hard to treat. Eliminating CSCs is a formidable and ongoing challenge because there is the need to spare normal stem cells as much as possible. As a starting point, targeting signaling events that play a crucial and selective role in governing the behavior of stem cells would spare their maturing and mature offspring. Activation of the RARs by all-trans retinoic acid (ATRA), the active metabolite of vitamin A, is well known to play a central role in controlling organogenesis during early embryonic development. The three main types of RAR, namely RARa, RARB, and RARy, form dimers with the retinoic acid X receptor. ATRA binding leads to the release of co-repressors of gene expression, the binding of coactivators, and gene expression.

Importantly, RARy is selectively expressed by HSCs and their immediate offspring, and ATRA activation supports HSC selfrenewal and maintenance as seen from a reduction in the number of HSCs in RARy knockout mice (Purton et al., 2006). In contrast, it is well known that activation of RARa drives the differentiation of myeloid progenitor cells and promyeloid cell lines. From studies of early zebrafish embryos, RARy expression is also restricted to primitive cells. They include mesodermal and neural crest stem/progenitor cells, in the head area, in the lateral plate mesoderm, and in the presomitic mesoderm of the tail bud (Hale et al., 2006). Treatment of embryos with a RARy agonist blocked stem cell differentiation, preventing the formation of bone in the head area, neural ganglia, and fins. Tbx-5immunopositive cells, for pectoral fin growth, were present in the appropriate region, and reversal of the action of the RARy agonist, by the use of a RARy antagonist or washout, restored fine growth (Wai et al., 2015). Taken together, the studies of hematopoiesis and zebrafish have shown that the activity statuses of RARy and RAR $\alpha$  mediate a balance between stem cell self-renewal and differentiation, playing opposing roles to promote stem cell maintenance and proliferation versus differentiation, respectively.

Hence, RAR $\gamma$  provides a putative target for eliminating CSCs, but further consideration is whether CSCs are more dependent than normal stem cells on the activation status of RAR $\gamma$  for their survival and/or proliferation. Activation of RAR $\gamma$  is vital to the survival and proliferation of prostate cancer (PCa) cells for three reasons. First, ATRA is at the limit of detection (~1 ng/g tissue) within patients' PCa tissue; adjacent normal cells and benign hyperplasia cells see ~8 times more (Pasquali et al., 1996). PCa cells appear, therefore, to be adapted to survive and grow in an exceedingly low ATRA environment. Secondly, PCa cells and normal prostate epithelium express RAR $\alpha$  and RAR $\gamma$ . For LNCap PCa cells, a sub nM level of ATRA (0.24 nM) transactivates RAR $\gamma$ , whereas 19.3 nM is needed to activate RAR $\alpha$  (Brown et al., 2017). PCa cells appear, therefore, to be dependent on active RAR $\gamma$  for survival and/or proliferation. Finally, to test this assumption, the DU145, LNCaP, and PC3 cell line cells were treated with 10<sup>-10</sup> to 10<sup>-9</sup> M ATRA, to activate just RAR $\gamma$ , or a RAR $\gamma$  agonist; this enhanced cell survival and proliferation (Petrie et al., 2020).

In keeping with active RARy plays a role in PCa survival is that antagonism of RARy and all RARs eliminated the CSC-like cells that formed colonies when the DU145, LNCaP, and PC3 PCa cell line cells were plated into dishes (Petrie et al., 2020). The RARy and pan-RAR antagonists killed the PCa cell line cells and primary PCa patients' cells when grown in flasks. Normal prostate epithelial cells were 50% less sensitive than PCa cells to both of the antagonists, and blood mononuclear cells and fibroblasts were insensitive to the pan-RAR antagonist. It is important to note that the RARy antagonist killed PCa CSClike cells and all of their offspring and all of the cells within cultures of primary patients' cells (Keedwell et al., 2004; Petrie et al., 2020). Therefore, there seems to be a failure to switch off RARy expression when PCa CSCs mature to give rise to the population of overt cancer cells. As above, activation of RARy promotes cell expansion, and in this way sustained RARy expression within the offspring of CSCs might contribute to disease burden to adversely affect patient prognosis. Further support for the use of a RARy antagonist to treat PCa is that a RARy network governs androgen signaling within and disease progression of PCa, as several of the target genes (e.g., SOX15) are associated with worse disease-free survival (Long et al., 2019). Antagonism of RARs was effective against other cancers. Antagonizing all RARs killed MCF7 and MDA-MB-231 breast cancer cell line cells and pediatric patients' primitive neuroectodermal tumor and astrocytoma cells, including elimination of the CSCs that gave rise to neurospheres (Brown and Petrie, 2021).

The three well-described modes of cell death are apoptosis (known as type I), autophagic (type II), and necrosis (type III). When Jurkat T-cell leukemia cells were deprived of ATRA they died via a caspase-independent and, therefore, non-apoptotic, process. They died via necroptosis which is a programmed form of necrosis involving the activation of poly-(ADP-ribose) polymerase 1 (PARP1) (Chiu et al., 2008). In turn, this has been associated with mitochondrial dysfunction and the release of ATP, NAD<sup>+</sup>, and the caspase-independent nucleases AIF and endonuclease G. RARy antagonist treated PCa cells growth arrested in G1 of cell cycle and cell death was mitochondria depolarisation-dependent and cellular DNA fragmentation occurred that was caspase-independent (Keedwell et al., 2004; Petrie et al., 2020). PARP1 inhibition, by 1,5dihydroisoquinoline, blocked the RARy antagonist's action, and, therefore, the antagonist-mediated cell death is via necroptosis.

The cytosolic death complex ripoptosome drives necroptosis. It comprises caspase 8, the kinases receptor-interacting protein 1 (RIPK1) and RIPK3 and the adaptor proteins FAS-associated DEATH domain (FADD) and TNFR1-associated DEATH domain (TRADD) (Wrighton, 2011). Cytosolic RARy plays a pivotal role in driving necroptosis that is provoked by DNAdamaging compounds which led to RARy being released from the nucleus to the cytosol. Loss of RARy abolished DNA damageinduced necroptosis, as seen from studies of knockout mice and primary squamous cell carcinoma cells. These studies showed that RARy is essential to RIPK1 and RIPK3 forming a complex, and the recruitment of the mixed lineage kinase domain-like protein (pMLKL) which is activated by RIP3 (Kadigamuwa et al., 2019). Cytosolic RARy has also been shown to be pivotal to tumor necrosis factor (TNF)-induced cell death of HT-29 colorectal adenocarcinoma cells (Xu et al., 2017). Caspase 8, a component of the ripoptosome, is negatively regulated by the cellular inhibitor of apoptosis 1 (cIAP1) and cIAP2. For TNFdriven necroptosis of HT-29 cells, it has been shown that RARy controls RIPK1-initiated cell death when the activity of cIAP was inhibited. RARy mediated dissociation of RIPK1 from TNF receptor 1 to facilitate necroptosis. Antagonism of RARy within PCa cells might, therefore, have led to RARy accumulating in the cytoplasm.

Further support to targeting RARy to eliminate CSCs is that RARy is an oncogene for some cancers. Some AML patients' cells harbour RARy fusion proteins (Conserva et al., 2019). RARy is often over expressed in colorectal carcinoma, and knockdown of RARG within the colorectal carcinoma cell lines HT29, HCT116, RK0 and SW480 enhanced their sensitivity to the chemotherapeutic drugs 5-fluorouracil, oxaliplatin and vincristine sulfate (Huang et al., 2017). RARy overexpression in the chemo-resistant bile duct carcinoma cholangiocarcinoma is associated with a poor prognosis and resistance to 5-flurouracil (Huang et al., 2013). Overexpression of RARy promotes the growth of hepatocellular carcinoma xenografts in mice (Yan et al., 2010) and occurs in around 50% of cases of clear cell renal carcinoma (Kudryavtseva et al., 2016). For gastric cancer, RARy has been identified as an important regulatory TF from mRNA microarray studies of differentially expressed genes (Wang, 2017). RARy overexpression within colorectal carcinoma, cholangiocarcinoma, and clear cell renal and hepatocellular carcinomas were measured for the cancer cell populations as a whole, rather than just for the CSC component. As above for PCa, the bulk of the above carcinoma cells seem to express RARy and active RARy maintains a stem cell phenotype. The presence of immature cells is a feature of cholangiocarcinoma and hepatocellular carcinoma. Progenitor cells give rise to these carcinomas with markers of progenitor cells. A very different view is that hepatocytes are the cell-of-origin with the transformed cells dedifferentiating into cells that express progenitor cell markers for hepatocellular carcinoma or transdifferentiating into bile duct-like cells for

cholangiocarcinoma (Sia et al., 2017). Aside from this uncertainty, an inappropriate expression of RAR $\gamma$  may underlie a precursor cell phenotype.

There are efforts to develop immunotherapeutic approaches to eradicating CSCs. Chimeric antigen receptor T-cells (CAR T) have engineered synthetic receptors that direct these cells to recognize cells that express the target antigen at their surface. The aim is to target molecules that are selectively expressed by cancer cells. For B-ALL, the target antigens used for CAR T design have included, for example, CD19 and CD20, whereby CD19 is expressed throughout B-cell development and CD20 is a B-cell-specific antigen. CD20-based CAR T therapy, alone or in combination with CD19-based therapy, was highly efficacious in animal models of B-ALL (Riaz et al., 2017). Accordingly, CAR T therapy has led to effective and durable clinical responses for hematological malignancies, providing effective and safe treatment for ALL in phase I/II clinical trials (Sheykhasan et al., 2022). Even so, successes from the use of CAR T-cells are still rare and limited for solid tumors by the physical barrier of the niche the cancer cells reside in (Liu et al., 2019). A challenge to preclinical studies is that there is a lack of translational animal models that are relevant. Antibodies that target a cell surface antigen also provide a route to immunotherapy of CSCs, and, for example, there are endeavors to target epidermal growth factor receptor and human epidermal growth factor receptor three in the case of triple-negative breast cancer (Rau et al., 2020). For both CAR T and antibody targeting, there is still often the need to identify a proper target, including even for patients with the same type of cancer, and moreover, antigen expression by cancer cells is generally heterogeneous.

The major intracellular signaling pathways within CSCs also provide avenues that are being explored to target CSCs. Their importance to the control of the proliferation, apoptosis, and metastasis of CSCs and endeavors to target are reviewed extensively elsewhere (Chen et al., 2013; Yang et al., 2020). In brief, the signaling pathways include Wnt, NF-KB, Notch, Hh, JAK-STAT, PI3K/AKT/mTOR, TGF/Smad, and PPAR. For example, Wnt signaling has a role in regulating the differentiation status of CSCs and apoptosis of these cells, and studies of PI3K/AKT/mTOR signaling within cancer cells have revealed roles in the maintenance of stemness and cell survival. Some of the proposed strategies to targeting signaling pathways kill CSCs. Sulphoraphane and curcumin inhibit the NF-κB pathway and regulate the self-renewal of breast cancer CSCs and the apoptosis of hepatocellular carcinoma CSCs, respectively. Disulfiram, which is used to treat alcoholism, inhibits the metastasis of breast cancer CSCs via interfering with NF-κB/ERK/Snail pathway signalling. The adenosine 5'-monophosphate-activated protein kinase signaling pathway (AMPK) is important to glucose balance within CSCs. The antidiabetic drug metformin targets AMPK signalling and inhibits the proliferation of colorectal and hepatocellular carcinoma CSCs. Small molecule antagonists of Hh signalling that inhibit tumor cell growth have been described, such as

GANT-61 (Lauth et al., 2007). Other efforts to target signaling pathways within CSCs are still under preclinical evaluation. And, it is important to note that there is cross-talk between pathways leading to the promotion of CSC maintenance and expansion, and, in turn, metastasis. A hurdle to the effective elimination of CSCs is the need to identify novel signalling within CSCs because they share many signaling pathways with normal stem cells.

In 2012, ferroptosis, a type of autophagy-dependent cell death, was described for NRAS-mutant HT-1080 fibrosarcoma cells treated with RAS-selective lethal compounds (e.g. erastin) (Dixon et al., 2012). It involves the iron-dependent accumulation of lipid hydroperoxides and is an emerging focus regarding the selective targeting of CSCs (Elgendy et al., 2020; Wu et al., 2021). In particular, there is growing evidence to support the view that oncogenic signaling renders cells susceptible to ferroptosis (Angeli et al., 2019). Medicinal chemistry has long been a dominant approach to the development of anticancer drugs, and investigators discovered small molecules with the potential to selectively kill CSCs when they developed new open-chain epothilone analogues. They killed mesenchymal breast cancer cells with CSC properties by harnessing reactive oxygen species to induce ferroptosis (Taylor et al., 2019).

Eliminating CSC may provide a cure for disease relapse. The frequency of CSCs within a tumor varies and does this affect the risk of relapse? For adenocarcinoma and squamous cell carcinoma of the lung, an observational prospective cohort study assessed the influence of the frequency of CSCs on relapse. CSCs were present at around three percent within primary cell cultures. Primary analyses showed that the risk of relapse was not affected by the frequency of CSCs. For patients with locally advanced disease, the risk of relapse was increased by a higher frequency of CSCs (Masciale et al., 2022). The findings for other tumors will be of interest regarding whether the frequency of CSCs is an important consideration to relapse. Other factors are important. Longitudinal analysis of cell-free DNA from patients with small cell lung cancer has brought to attention the importance of treatment efficacy and the frequencies of mutant alleles and copy number alterations. Disease-associated mutations were observed in eight-five percent of patient samples and mutant allele frequencies ranged from very low (0.1%) to eighty-seven percent. The response to treatment tracked closely with mutant allele frequencies and alterations to copy number (Almodovar et al., 2018). For breast cancer, the recurrence of metastatic disease is varied and unpredictable with investigators commenting that there are "lingering mysteries of disease recurrence" (Riggio et al., 2021).

### Perspectives and conclusions

Advancing knowledge about the different types of leukemias coupled to an understanding of the architecture of the hematopoietic cell system brought leukemia to the forefront as a model system for cancer. The relative ease of accessibility of patients' cells contributed, and, from the 1980s, hematopoietic cell lines were generated that typified almost all of the subsets of the lymphoid and myeloid leukemias (except for CML chronic phase) (Drexler et al., 1999). It might be argued that leukemia is an inappropriate mode for cancer because metastasis of prevalent carcinomas is a longstanding and major challenge to treating cancer successfully. However, leukemia is not exceptional because, from studies of the anatomy of the leukemias, it has been argued that their dissemination is a metastatic process (Whiteley et al., 2012).

Findings from studies of AML markedly shaped our view of the nature of cancer. They were that a rare subpopulation of cells, namely LSCs, sustains the hierarchy of leukemia cells and that LSCs share unique biological properties with HSCs. An attractive hypothesis, from consideration of the cell of origin of some leukemias, is that many cancers may arise directly from a tissuespecific stem cell. Seminal studies of B-ALL have shown that, at least, two genetic insults are required for leukemia. Some leukemias arise from an HSC and there is still the need to unravel how each of the insults contributes to an alteration to cell behavior. The targeting of some oncogenes to HSCs/HPCs restricted LSCs to one differentiation pathway. Mathematical modeling has revealed a high order of multi-stability for HSCs, rather than bi-stability, with noise and bursting of gene expression playing a role in HSCs veering towards a lineage (Wou et al., 2022). This plasticity is essential to the stem cell state, and HSCs may shift towards a lineage option by guidance from the environment. The hematopoietic cytokines erythropoietin and macrophage colony-stimulating factors guide erythroid and myeloid fate within HSCs, respectively (Mossadegh-Keller et al., 2013; Grover et al., 2014). As to the bursting of gene expression, the transformation of an HSC by some oncogenes may enforce a lineage affiliation at a moment in time of a particular burst of gene expression.

Conventional treatments that are often highly effective against the bulk of cancer cells spare LSCs/CSCs. Agents that drive apoptosis are less effective against CSCs because many cancers are intrinsically resistant or acquire resistance to the induction of apoptosis (Fulda, 2009), and necroptosis is considered as a "fail-safe" death pathway for apoptosisresistant cells (Gong et al., 2019). Pan-RAR and RARy antagonists were potent inducers of necroptosis of PCa, breast cancer, and pediatric patients' brain tumor CSCs and their offspring, and normal prostate epithelium cells were less sensitive than PCa cells. A number of other compounds are known to drive necroptosis of cancer cells, the list is growing, and it includes compounds that drive necroptosis of apoptosisresistant cancer cells (Fulda, 2013).

The stem cell theory of cancer is still a work in progress. Though CSCs have been identified for a good number of human solid tumors, whether the risk of relapse is increased by a higher frequency of CSCs needs further investigation for patients at presentation and with locally advanced disease. As for all cancer treatments, the aim is to kill or control cancer cells avoiding damage to normal tissues. This issue is highly pertinent to endeavors to kill CSCs when the normal counterpart cell is a tissue-specific stem cell because of its importance to replenishing worn out cells. These two cell populations share many characteristics, as seen for signaling pathways. Finding specific attributes of CSCs, for targeting, requires a much clearer picture of their nature versus their counterpart tissue-specific stem cell. Whether most, if not all, malignancies arise directly from a tissue-specific stem cell is still debatable. If so, how the progeny of LSCs/CSCs become restricted to a cell lineage is, as yet, unclear. The resistance of many cancer to apoptosis has prompted the need to explore driving other cell death modalities, such as necroptosis or ferroptosis. In this case, whether CSCs are more sensitive than normal tissue-specific stems cells to the induction of these death modalities is an important issue. The search is for agents that kill CSCs and that spare tissue-specific stem cell populations. The resolve to this problem may well require the development of new synthetic compounds that target the specific attributes of CSCs when we have a clearer picture of their nature.

### Author contributions

The author confirms being the sole contributor of this work and has approved it for publication.

### Funding

This project received funding from the European Union's Seventh Framework Programme for research, technological development and demonstration under grant agreement No. 315902. GB was the lead within the Marie Curie Initial Training Network DECIDE (Decision-making within cells and differentiation entity therapies).

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### References

Adolfsson, J., Mansson, R., Buza-Vidas, N., Hultquist, A., Liuba, K., Jensen, C. T., et al. (2005). Identification of  $\mathrm{Fll}^{3+}$  lympho-myeloid stem cells lacking erythromegakaryocytic potential: A revised road map for adult blood lineage commitment. *Cell* 121, 295–306. doi:10.1016/j.cell.2005.02.013

Alizadeh, A. A., and Majeti, R. (2011). Surprise! HSC are aberrant in chronic lymphoblastic leukemia. *Cancer Cell* 20, 135–136. doi:10.1016/j.ccr.2011.08.001

Almodovar, K., Iams, W. T., Meador, C. B., Zhao, Z., York, S., Horn, S., et al. (2018). Longitudinal cell-free DNA analysis in patients with small cell lung cancer reveals dynamic insights into treatment efficacy and disease relapse. *J. Thorac. Oncol.* 13, 112–123. doi:10.1016/j.jtho.2017.09.1951

Angeli, J. P. F., Krysko, D. V., and Conrad, M. (2019). Ferroptosis at the crossroads of cancer acquired drug resistance and immune evasion. *Nat. Rev. Cancer* 19, 405–414. doi:10.1038/s41568-019-0149-1

Bertoni, F., and Zucca, E. (2006). Delving deeper into MALT lymphoma biology. J. Clin. Invest. 116, 22–26. doi:10.1172/JCI27476

Bonnet, D., and Dick, J. E. (1997). Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat. Med.* 7, 730–737. doi:10.1038/nm0797-730

Box, J. K., Paquet, N., Adams, M, N., Boucher, D., Bolderson, E., O'Byrne, K. J., et al. (2016). Nucleophosmin: From structure and function to disease development. *BMC Mol. Biol.* 17, 19. doi:10.1186/s12867-016-0073-9

Brown, G., and Ceredig, R. (2019). Modeling the hematopoietic landscape. Front. Cell Dev. Biol. 7, 104. doi:10.3389/fcell.2019.00104

Brown, G., Marchwicka, A., Cunningham, A., Toellner, K. M., and Marcinkowska, E. (2017). Antagonizing retinoic acid receptors increases myeloid cell production by cultured human hematopoietic stem cells. *Arch. Immunol. Ther. Exp.* (*Warsz.*) 65, 69–81. doi:10.1007/s00005-016-0411-0

Brown, G., and Petrie, K. (2021). The RARy oncogene: An achilles heel for some cancers. Int. J. Mol. Sci. 22, 3632. doi:10.3390/ijms22073632

Ceredig, R., Rolink, A. G., and Brown, G. (2009). Models of haematopoiesis: Seeing the wood for the trees. *Nat. Rev. Immunol.* 9, 293–300. doi:10.1038/nri2525

Challen, G. A., Boles, N. C., Chambers, S. M., and Goodell, M. A. (2010). Distinct hematopoietic stem cell subtypes are differentially regulated by TGF-beta1. *Cell Stem Cell* 6, 265–278. doi:10.1016/j.stem.2010.02.002

Chambers, T., and Rabbitts, T. H. (2015). LMO2 at 25 years: A paradigm of chromosomal translocation proteins. *Open Biol.* 5, 150062. doi:10.1098/rsob. 150062

Chen, K., Huang, Y.-H., and Chen, J.-L. (2013). Understanding and targeting cancer stem cells: Therapeutic implications and challenges. *Acta Pharmacol. Sin.* 34, 732–740. doi:10.1038/aps.2013.27

Chiu, H. J., Fischman, D. A., and Hammerling, U. (2008). Vitamin A depletion causes oxidative stress, mitochondrial dysfunction, and PARP1-dependent energy deprivation. *FASEB J.* 22, 3878–3887. doi:10.1096/fj.08-112375

Collins, A. T., Berry, P. A., Hyde, C., Stower, M. J., and Maitland, N. J. (2005). Prospective identification of tumorigenic prostate cancer stem cells. *Cancer Res.* 65, 10946–10951. doi:10.1158/0008-5472.CAN-05-2018

Conserva, M. R., Redavid, I., Anelli, L., Zagaria, A., Specchia, G., and Albano, F. (2019). RARG gene dysregulation in acute myeloid leukemia. *Front. Mol. Biosci.* 6, 114. doi:10.3389/fmolb.2019.00114

Corbin, A. S., Agarwal, A., Loriaux, M., Cortes, J., Deininger, M. W., and Druker, B. J. (2011). Human chronic myeloid leukemia stem cells are insensitive to imatinib despite inhibition of BCR-ABL activity. *J. Clin. Invest.* 121, 396–409. doi:10.1172/ JCI35721

Cox, C. V., and Blair, A. (2005). A primitive cell origin for B-cell precursor ALL. Stem Cell Rev. 1, 189–196. doi:10.1385/SCR:1:3:189

Dick, J. E. (2008). Stem cell concepts renew cancer research. Blood 112, 4793-4807. doi:10.1182/blood-2008-08-077941

Dixon, S. J., Lemberg, K. M., Lamprecht, M. R., Skouta, R., Zaitsov, E. M., Gleason, C. E., et al. (2012). Ferroptosis: An iron-dependent form of nonapoptotic cell death. *Cell* 149, 1060–1072. doi:10.1016/j.cell.2012.03.042

Downing, J. R., Wilson, R. K., Zhang, J., Mardis, E. R., Pui, C. H., Ding, L., et al. (2012). The pediatric cancer genome project. *Nat. Genet.* 44, 619–622. doi:10.1038/ng.2287

Drexler, H. G., MacLeod, R. A. F., and Uphoff, C. C. (1999). Leukemia cell lines: *In vitro* models for the study of philadelphia chromosome-positive leukemia. *Leuk. Res.* 23, 207–215. doi:10.1016/s0145-2126(98)00171-4

Edwards, R. H., Wasik, M. A., Finan, J., Rodriguez, R., Moore, J., Kamoun, M., et al. (1999). Evidence for early hematopoietic progenitor cell involvement in acute

promyelocytic leukemia. Am. J. Clin. Pathol. 112, 819–827. doi:10.1093/ajcp/112. 6.819

Elgendy, S. M., Alyammahi, S. K., Alhamad, D. W., Abdin, S. M., and Omar, H. A. (2020). Ferroptosis: An emerging approach for targeting cancer stem cells and drug resistance. *Crit. Rev. Oncol. Hematol.* 155, 103095. doi:10.1016/j.critrevonc.2020. 103095

Eramo, A., Lotti, F., Sette, G., Pilozzi, E., Biffoni, M., Di Virgilio, A., et al. (2008). Identification and expansion of the tumorigenic lung cancer stem cell population. *Cell Death Differ*. 15, 504–514. doi:10.1038/sj.cdd.4402283

Fialkow, P. J., Denman, A. M., Jacobson, R. J., and Lowenthal, M. N. (1978). Chronic myelocytic leukemia. Origin of some lymphocytes from leukemic stem cells. J. Clin. Invest. 62, 815–823. doi:10.1172/JCI109193

Ford, A. M., Bennett, C. A., Price, C. M., Bruin, M. C. A., Van Wering, E. R., and Greaves, M. (1998). Fetal origins of the TEL-AML1 fusion gene in identical twins with leukemia. *Proc. Natl. Acad. Sci. U. S. A.* 95, 4584–4588. doi:10.1073/pnas.95.8. 4584

Ford, A. M., Ridge, S. A., Cabrera, M. E., Mahmoud, H., Steel, C. M., Greaves, M., et al. (1993). *In utero* rearrangements in the trithorax-related oncogene in infant leukaemias. *Nature* 363, 358–360. doi:10.1038/363358a0

Fulda, S. (2013). The mechanism of necroptosis in normal and cancer cells. *Cancer Biol. Ther.* 14, 999-1004. doi:10.4161/cbt.26428

Fulda, S. (2009). Tumor resistance to apoptosis. Int. J. Cancer 124, 511-515. doi:10.1002/ijc.24064

Fuxa, M., and Skok, J. A. (2007). Transcriptional regulation in early B cell development. *Curr. Opin. Immunol.* 19, 129–136. doi:10.1016/j.coi.2007. 02.002

Garcia-Ramirez, L., Bhatia, S., Rodriguez-Hermandez, G., Gonzalez-Herrero, L., Walter, C., Gonzales de Tena-Davilo, S., et al. (2018). Lmo2 expression defines tumor cell identity during T-cell leukemogenesis. *EMBO J.* 37, e98783. doi:10. 15252/embj.201798783

Gekas, C., and Graf, T. (2013). CD41 expression marks myeloid-biased adult hematopoietic stem cells and increases with age. *Blood* 121, 4463–4472. doi:10.1182/ blood-2012-09-457929

Gong, Y., Fan, Z., Luo, G., Yang, C., Huang, Q., Fan, K., et al. (2019). The role of necroptosis in cancer biology and therapy. *Mol. Cancer* 18, 100. doi:10.1186/s12943-019-1029-8

Graham, S. M., Jorgensen, H. G., Allan, E., Pearson, C., Alcorn, M. J., Richmond, L., et al. (2002). Primitive, quiescent, Philadelphia-positive stem cells from patients with chronic myeloid leukemia are insensitive to STI571 *in vitro. Blood* 99, 319–325. doi:10.1182/blood.v99.1.319

Greaves, M. F. (1981). Analysis of the clinical and biological significance of lymphoid phenotypes in acute leukemia. *Cancer Res.* 41, 4752–4766.

Greaves, M. F., Maia, A. T., Wiemels, J. L., and Ford, A. M. (2003). Leukemia in twins: Lessons in natural history. *Blood* 102, 2321–2333. doi:10.1182/blood-2002-12-3817

Grover, A., Mancini, E., Moore, S., Mead, A. J., Atkinson, D., Rasmussen, K. D., et al. (2014). Erythropoietin guides multipotent hematopoietic progenitor cells toward an erythroid fate. *J. Exp. Med.* 211, 181–188. doi:10.1084/jem. 20131189

Hale, L. A., Tallafuss, A., Yan, Y-L., Dudley, L., Eisen, J. S., and Postlethwait, J. H. (2006). Characterization of the retinoic acid receptor genes raraa, rarab and rarg during zebrafish development. *Gene Expr. Patterns* 6, 546–555. doi:10.1016/j. modgep.2005.10.007

Heath, E. M., Chan, S. M., Minden, M. D., Murphy, T., Shlush, L. I., and Schimmer, A. D. (2017). Biological and clinical consequences of *NPM1* mutations in AML. *Leukemia* 31, 798–807. doi:10.1038/leu.2017.30

Hermann, P. C., Huber, S. L., Herrler, T., Aicher, A., Ellwart, J. W., Guba, M., et al. (2007). Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 1, 313–323. doi:10. 1016/j.stem.2007.06.002

Hirt, A., Schmid, A-M., Amman, R. A., and Leibundgut, K. (2011). In pediatric lymphoblastic leukemia of B cell origin, a small population of primitive blast cells is noncycling, suggesting them to be leukemia stem candidates. *Pediatr. Res.* 69, 194–199.

Holyoake, T., Jiang, X., Eaves, C., and Eaves, A. (1999). Isolation of a highly quiescent subpopulation of primitive leukemic cells in chronic myeloid leukemia. *Blood* 94, 2056–2064. doi:10.1182/blood.v94.6.2056.418k13\_2056\_2064

Huang, G. L., Luo, Q., Rui, G., Zhang, W., Zhang, Q. Y., Chen, Q. X., et al. (2013). Oncogenic activity of retinoic acid receptor  $\gamma$  is exhibited through activation of the Akt/NF- $\kappa B$  and Wnt/ $\beta$ -catenin pathways in cholangiocarcinoma. Mol. Cell. Biol. 33, 3416–3425. doi:10.1128/MCB.00384-13

Huang, G. L., Song, W., Zhou, P., Fu, Q. R., Lin, C. L., Chen, Q. X., et al. (2017). Oncogenic retinoic acid receptor  $\gamma$  knockdown reverses multi-drug resistance of human colorectal cancer via Wnt/ $\beta$ -catenin pathway. *Cell Cycle* 16, 685–692. doi:10. 1080/15384101.2017.1295180

Kadigamuwa, C., Choksi, S., Xu, Q., Cataisson, C., Greenbaum, S. S., Yuspa, S. H., et al. (2019). Role of retinoic acid receptor-gamma in DNA damage-induced necroptosis. *iScience* 17, 74–86. doi:10.1016/j.isci.2019.06.019

Kastner, P., and Chan, S. (2001). Function of RARalpha during the maturation of neutrophils. *Oncogene* 20, 7178–7185. doi:10.1038/sj.onc.1204757

Keedwell, R. G., Zhao, Y., Hammond, L. A., Wen, K., Qin, S., Atangan, L. I., et al. (2004). An antagonist of retinoic acid receptors more effectively inhibits growth of human prostate cancer cells than normal prostate epithelium. *Br. J. Cancer* 91, 580–588. doi:10.1038/sj.bjc.6602024

Kellaway, S. G., Keane, P., Edginton-White, B., Regha, K., Kennet, E., and Bonifer, C. (2021). Different mutant RUNC1 oncoproteins program alternative haematopoietic differentiation trajectories. *Life Sci. Alliance* 4, e20200084. doi:10.26508/lsa.202000864

Kikushige, Y., Iskikawa, F., Miyamoto, T., Shima, T., Urata, S., Yashimoto, T., et al. (2011). Self-renewing hematopoietic stem cell is the primary target in pathogenesis of human chronic lymphoblastic leukemia. *Cancer Cell* 20, 246–259. doi:10.1016/j.ccr.2011.06.029

Kondo, M., Wagers, A. J., Manz, M. G., Prohaska, S. S., Scheer, D. C., Belhack, G. F., et al. (2003). Biology of hematopoietic stem cells and progenitors: Implications for clinical application. *Annu. Rev. Immunol.* 21, 759–806. doi:10.1146/annurev. immunol.21.120601.141007

Kudryavtseva, A. V., Nyushko, K. M., Zaretsky, A. R., Shagin, D. A., Kaprin, A. D., Alekseev, B. Y., et al. (2016). Upregulation of Rarb, rarg, and rorc genes in clear cell renal cell carcinoma. *Biomed. Pharmacol. J.* 9, 967–975. doi:10.13005/bpj/1036

Land, H., Parada, L. F., and Weinberg, R. A. (1983). Tumorigenic conversion of primary embryo fibroblasts requires at least two cooperating oncogenes. *Nature* 304, 596–602. doi:10.1038/304596a0

Lathia, J. D., Mack, S. C., Mulkearns, E. E., Valentin, C. L. L., and Rich, J. N. (2015). Cancer stem cells in glioblastoma. *Genes Dev.* 29, 1203–1217. doi:10.1101/gad.261982.115

Lauth, M., Bergstrom, A., Shokkawa, T., and Toftgard, R. (2007). Inhibition of GLI-mediated transcription and tumor cell growth by small-molecule antagonists. *Proc. Natl. Acad. Sci. U. S. A.* 104, 8455–8460. doi:10.1073/pnas.0609699104

Lichtinger, M., Ingram, R., Hannah, R., Muller, D., Clarke, D., Assis, S. A., et al. (2012). RUNX1 reshapes the epigenetic landscape at the onset of haematopoiesis. *EMBO J.* 31, 4318–4333. doi:10.1038/emboj.2012.275

Liu, B., Yan, L., and Zhou, M. (2019). Target selection of CAR T cell therapy in accordance with the TME for solid tumors. *Am. J. Cancer Res.* 9, 228–241.

Long, M. D., Singh, P. K., Russell, J. R., Llimos, G., Rosario, S., Rivzvi, A., et al. (2019). The miR-96 and RAR $\gamma$  signaling axis governs androgen signaling and prostate cancer progression. *Oncogene* 38, 421–444. doi:10.1038/s41388-018-0450-6

Maia, A. T., Ford, A. M., Jalali, G. R., Harrison, C. J., Taylor, G. M., Eden, O. B., et al. (2001). Molecular tracking of leukemogenesis in a triplet pregnancy. *Blood* 98, 478–482. doi:10.1182/blood.v98.2.478

Malouf, C., and Ottersbach, K. (2018). The fetal liver lymphoid-primed multipotent progenitor provides the prerequisites for the initiation of t(4;11) MLL-AF4 infant leukemia. *Haematologica* 103, e571–e574. doi:10.3324/ haematol.2018.191718

Martin-Lorenzo, A., Auer, F., Chan, L. N., Garcia-Ramirez, I., Gonzalez-Herrero, I., Rodriguez-Hernandez, G., et al. (2018). Loss of Pax5 exploits sca1-BCR-ABL(p190) susceptibility to confer the metabolic shift essential for pB-ALL. *Cancer Res.* 78, 2669–2679. doi:10.1158/0008-5472.CAN-17-3262

Masciale, V., Banchelli, F., Grisendi, G., D'Amico, R., Majorana, U., Stefani, A., et al. (2022). The influence of cancer stem cells on the risk of relapse in adenocarcinoma and squamous cell carcinoma of the lung: A prospective cohort study. *Stem Cells Transl. Med.* 11, 239–247. doi:10.1093/stcltm/szab029

McCromak, M. P., Forster, A., Drynaan, L., Pannell, R., and Rabbitts, T. H. (2003). The LM02 T-cell oncogene is activated by chromosomal translocations or retroviral insertion but has no mandatory role in normal T-cell development. *Mol. Cell. Biol.* 23, 9003–9013.

Miyamoto, T., Nagafuji, K., Akashi, K., Harada, M., Kyo, T., and Akashi, T. (1996). Persistence of multipotent progenitors expressing AML1/ETO transcripts in long-term remission patients with t(8;21) acute myelogenous leukemia. *Blood* 87, 4789–4796. doi:10.1182/blood.v87.11.4789.bloodjournal87114789

Miyamoto, T., Weissman, I. L., and Akashi, K. (2000). AML1/ETO-expressing nonleukemic stem cells in acute myelogenous leukemia with 8;21 chromosomal translocation. *Proc. Natl. Acad. Sci. U. S. A.* 97, 7521–7526. doi:10.1073/pnas.97.13. 7521

Mooney, C. J., Cunningham, A., Tsapogas, P., Toellner, K. M., and Brown, G. (2017). Selective expression of flt3 within the mouse hematopoietic stem cell compartment. *Int. J. Mol. Sci.* 18, 1037. doi:10.3390/ijms18051037

Mossadegh-Keller, N., Sarrazin, S., Kandella, P. K., Espinosa, L., Stanley, E. R., Nutt, S. L., et al. (2013). M-csf instructs myeloid lineage fate in single haematopoietic stem cells. *Nature* 497, 239–243. doi:10.1038/nature12026

Munro, M. J., Wickremesekera, S. K., Peng, L., Tan, S. T., and Itinteang, T. (2018). Cancer stem cells in colorectal cancer: A review. *J. Clin. Pathol.* 71, 110–116. doi:10. 1136/jclinpath-2017-204739

Naik, S. H., Perie, L., Swart, E., Gerlach, C., van Rooij, N., de Boer, R. J., et al. (2013). Diverse and heritable lineage imprinting of early haematopoietic progenitors. *Nature* 496, 229–232. doi:10.1038/nature12013

Nestorowa, S., Hamey, F. K., Pijuan Sala, B., Diamanti, F., Shepherd, M., Laurenti, E., et al. (2016). A single-cell resolution map of mouse hematopoietic stem and progenitor cell differentiation. *Blood* 128, e20–31. doi:10.1182/blood-2016-05-716480

Pasquali, D., Thaller, C., and Eichele, G. (1996). Abnormal level of retinoic acid in prostate cancer tissues. *J. Clin. Endocrinol. Metab.* 81, 2186–2191. doi:10.1210/jcem. 81.6.8964849

Perez-Caro, M., Cobaleda, C., Gonzalez-Herrero, I., Vicente-Duenas, C., Bermejo-Rodriguez, C., Sanchez-Beato, M., et al. (2009). Cancer induction by restriction of oncogene expression to the stem cell compartment. *EMBO J.* 28, 8–20. doi:10.1038/emboj.2008.253

Petrie, K., Urban-Wojciuk, Z., Sbirkov, Y., Graham, A., Hamann, A., and Brown, G. (2020). Retinoic acid receptor gamma is a therapeutically targetable driver of growth and survival in prostate cancer. *Cancer Rep.* 3, e1284. doi:10.1002/cnr2.1284

Pombode de Oliveira, M. S., Awad el Seed, F. E., Matutes, E., Morilla, R., Luzzatto, L., Catovsky, D., et al. (1986). Lymphoblastic leukaemia in siamese twins: Evidence for identity. *Lancet* 7, 969–970. doi:10.1016/s0140-6736(86)90616-1

Pottier, C., Frenais, M., Gilon, M., Jerusalem, G., Longuespee, R., and Sounni, N. E. (2020). Tyrosine kinase inhibitors in cancer: Breakthrough and challenges of targeted therapy. *Cancers* 12, E731. doi:10.3390/cancers12030731

Prbavathy, D., Swarnalatha, Y., and Ramadoss, N. (2018). Lung cancer stem cells origin, characteristics and therapy. *Stem Cell Investig.* 5, 6. doi:10.21037/sci.2018. 02.01

Prince, M. E., Sivanandan, R., Kaczorowski, A., Wolf, G. T., Kaplan, M. J., and Dalerba, P. (2007). Identification of a subpopulation of cells with cancer stem cell properties in head and neck squamous cell carcinoma. *Proc. Natl. Acad. Sci. U. S. A.* 104, 973–978. doi:10.1073/pnas.0610117104

Purton, L. E., Dworkin, S., Olsen, G. H., Walkley, C. R., Fabb, S. A., Collins, S. J., et al. (2006). RARgamma is critical for maintaining a balance between hematopoietic stem cell self-renewal and differentiation. *J. Exp. Med.* 203, 1283–1293. doi:10.1084/jem.20052105

Quijano, C. A., Moore, D., 2nd, Arthur, D., Feusner, J., Winter, S. S., and Pallavicini, M. G. (1997). Cytogenetically aberrant cells are present in the CD34<sup>+</sup> CD33<sup>-</sup> 38<sup>-</sup> 19<sup>-</sup> marrow compartment in children with acute lymphoblastic leukemia. *Leukemia* 11, 1508–1515. doi:10.1038/sj.leu.2400754

Quintana, E., Shakleton, M., Sabel, M. S., Fuller, D. R., Johnson, T. M., and Morrison, S. J. (2008). Efficient tumour formation by single human melanoma cells. *Nature* 456, 593–598. doi:10.1038/nature07567

Radomska, H. S., Huettner, C. S., Zhang, P., Cheng, T., Scadden, D. T., and Tenen, D. G. (1998). CCAAT/enhancer binding protein alpha is a regulatory switch sufficient for induction of granulocytic development from bipotential myeloid progenitors. *Mol. Cell. Biol.* 18, 4301–4314. doi:10.1128/mcb.18.7.4301

Rau, A., Lieb, W. S., Seifert, O., Honer, J., Birnstock, D., Richter, F., et al. (2020). Inhibition of tumor cell growth and cancer stem cell expansion by a bispecific antibody targeting EGFR and HER3. *Mol. Cancer Ther.* 19, 1474–1485. doi:10.1158/ 1535-7163.MCT-19-1095

Rejeski, K., Dugue-Afonso, J., and Lubbert, M. (2021). AML1/ETO and its function as a regulator of gene transcription via epigenetic mechanisms. *Oncogene* 40, 5665–5676. doi:10.1038/s41388-021-01952-w

Riaz, I. B., Zahid, U., Kamal, M. U., Husnain, M., McBride, A., Hua, A., et al. (2017). Anti-CD19 and anti-CD20 CAR-modified T cells for B-cell malignancies: A systematic review and meta-analysis. *Immunotherapy* 9, 979–993. doi:10.2217/imt-2017-0062

Richkind, K. E., Loew, T., Meisner, L., Watson, D., and Wason, D. (1998). Identical cytogenetic clones and clonal evolution in pediatric monozygotic twins with acute myeloid leukemia: Presymptomatic disease detection by interphase fluorescence in situ hybridization and review of the literature. J. Pediatr. Hematol. Oncol. 20, 264–267. doi:10.1097/00043426-199805000-00015

Riddell, J., Gazit, R., Garrison, B. S., Guo, G., Saadatpour, A., Mandal, P. K., et al. (2014). Reprogramming committed murine blood cells to induced hematopoietic stem cells with defined factors. *Cell* 157, 549–564. doi:10.1016/j.cell.2014.04.006

Riggio, A., Varley, K. E., and Welm, A. L. (2021). The lingering mysteries of metastatic recurrence in breast cancer. *Br. J. Cancer* 124, 13–26. doi:10.1038/s41416-020-01161-4

Rodriguez-Hernandez, G., Casado-Garcia, A., Isidro-Hernandez, M., Picard, D., Raboso-Gallego, T., Aleman-Arteaga, S., et al. (2021). The second oncogenic hit determines the cell fate of ETV6- RUNX1 positive leukemia. *Front. Cell Dev. Biol.* 9, 704591. doi:10.3389/fcell.2021.704591

Rodriguez-Hernandez, G., Hauer, J., Martin-Lorenzo, A., Schafer, D., Bartenhagen, C., Garcia-Ramirez, I., et al. (2017). Infection exposure promotes ETV6-RUNX1 precursor B cell leukaemia via impaired H3K4 demethylases. *Cancer Res.* 77, 4265–4277.

Sanjuan-Pla, A., Macaulay, I. C., Jensen, C. T., Woll, P. S., Luis, T. C., Mead, A., et al. (2013). Platelet-biased stem cells reside at the apex of the haematopoietic stemcell hierarchy. *Nature* 502, 232–236. doi:10.1038/nature12495

Schafer, D., Olsen, M., Lahnemann, D., Stanulla, M., Slany, R., Schiegelow, A., et al. (2018). Five percent of healthy newborns have an *ETVG-RUNX1* fusion as revealed by DNA-based GIPFEL screening. *Blood* 131, 821–826. doi:10.1182/blood-2017-09-808402

She, J. J., Zhang, P. G., Wang, Z. M., Gan, W. M., and Che, X. M. (2008). Identification of side population cells from bladder cancer cells by dyecycle violet staining. *Cancer Biol. Ther.* 7, 1663–1668. doi:10.4161/cbt.7.10.6637

Sheykhasan, M., Manoochehri, H., and Dama, P. (2022). Use of CAR T-cell for acute lymphoblastic leukemia (ALL) treatment: A review study. *Cancer Gene Ther.* 29, 1080–1096. doi:10.1038/s41417-021-00418-1

Shimazu, T., Iida, R., Zhang, Q., Welner, R. S., Medina, K. L., Alberola-Lla, J., et al. (2012). CD86 is expressed on murine hematopoietic stem cells and denotes lymphopoietic potential. *Blood* 119, 4889–4897. doi:10.1182/blood-2011-10-388736

Sia, D., Villanueva, A., Friedman, S. L., and Clovet, J. M. (2017). Liver cancer cell of origin, molecular class, and effects on patient prognosis. *Gastroenterology* 152, 745–761. doi:10.1053/j.gastro.2016.11.048

Song, K., and Farzaneh, M. (2012). Signaling pathways governing breast cancer stem cells behavior. *Stem Cell Res. Ther.* 12, 245. doi:10.1186/s13287-021-02321-w

Spangrude, G. J., Heimfeld, S., and Weissman, I. L. (1988). Purification and characterization of mouse hematopoietic stem cells. *Science* 241, 58–62. doi:10. 1126/science.2898810

Swerdlow, S. H., Campo, E., Harris, N. L., Jaffe, E. S., Pileri, S. A., Stein, H., et al. (2008). WHO classification of tumours of haematopoietic and lymphoid tissues. 4th Edition. Lyon, France: IARC.

Taylor, W. R., Fedorka, S. R., Gad, I., Shah, R., Algahtani, H. D., Koranne, R., et al. (2019). Small-molecule ferroptotic agents with potential to selectively target cancer stem cells.. *Sci. Rep.* 9, 5926. doi:10.1038/s41598-019-42251-5

Vicente-Duenas, C., Gonzalez-Herrero, I., Sehgal, L., Garcia-Ramirez, I., Rodriguez-Hernandez, G., Pintado, B., et al. (2019). Dnmt1 links BCR-ABLp210 to epigenetic tumor stem cell priming in myeloid leukemia. *Leukemia* 33, 249–278. doi:10.1038/s41375-018-0192-z

Wai, H., A., Kawakami, K., Wada, H., Muller, F., Vernalis, A. B., Brown, G., et al. (2015). The development and growth of tissues derived from cranial neural crest and primitive mesoderm is dependent on the ligation status of retinoic acid receptor  $\gamma$ : Evidence that retinoic acid receptor  $\gamma$  functions to maintain stem/progenitor cells in the absence of retinoic acid. *Stem Cells Dev.* 24, 507–519. doi:10.1089/scd.2014. 0235

Wang, Y. (2017). Transcriptional regulatory network analysis for gastric cancer based on mRNA microarray. *Pathol. Oncol. Res.* 23, 785–791. doi:10.1007/s12253-016-0159-1

Whiteley, A. E., Price, T. T., Cantelli, G., and Sipkins, D. A. (2012). Leukaemia: A model metastatic disease. *Nat. Rev. Cancer* 21, 461–475. doi:10.1038/s41568-021-00355-z

Wiemels, J. L., Cazzaniga, G., Daniotti, M., Eden, O. B., Addison, G. M., Masera, G., et al. (1999). Pre-natal origin of acute lymphoblastic leukaemia in children. *Lancet* 354, 1499–1503. doi:10.1016/s0140-6736(99)09403-9

Wiemels, J. L., Ford, A. M., Van Wering, E. R., Postma, A., and Greaves, M. (1999). Protracted and variable latency of acute lymphoblastic leukemia after TEL-AML1 gene fusion *in utero. Blood* 94, 1057–1062. doi:10.1182/blood.v94.3.1057. 415k10\_1057\_1062

Winters, A. C., and Bernt, K. M. (2017). MLL-rearranged leukemias-an update on science and clinical approaches.. Front. Pediatr. 6, 4. doi:10.3389/fped.2017.00004

Wolman, I. D. (1962). Parallel responses to chemotherapy in identical twin infants with concordant leukemia.. *J. Pediatr.* 60, 91–95. doi:10.1016/s0022-3476(62)80014-6

Wou, S., Zhou, T., and Tian, T. A. (2022). A robust method for designing multistable systems. *Syst. Biol. Appl.* 8, 10.

Wrighton, K. H. (2011). A new platform for death. Nat. Rev. Mol. Cell Biol. 12, 547. doi:10.1038/nrm3174

Wu, C., Wei, Q., Utomo, V., Nadesan, P., Whetstone, H., Kandel, R., et al. (2007). Side population cells isolated from mesenchymal neoplasms have tumor initiating potential. *Cancer Res.* 67, 8216–8222. doi:10.1158/0008-5472.CAN-07-0999

Wu, S., Li, T., Liu, W., and Huang, Y. (2021). Ferroptosis and cancer: Complex relationship and potential application of exosomes. *Front. Cell Dev. Biol.* 9, 733751. doi:10.3389/fcell.2021.733751

Xu, Q., Jitkaew, S., Choksi, S., Kadigamuwa, C., Qu, J., Choe, M., et al. (2017). The cytoplasmic nuclear receptor RARy controls RIP1 initiated cell death when cIAP activity is inhibited.. *Nat. Commun.* 8, 425. doi:10.1038/s41467-017-00496-6

Yan, T. D., Wu, H., Zhang, H. P., Lu, N., Ye, P., Yu, F. H., et al. (2010). Oncogenic potential of retinoic acid receptor-gamma in hepatocellular carcinoma. *Cancer Res.* 70, 2285–2295. doi:10.1158/0008-5472.CAN-09-2968

Yang, L., Shi, P., Zhao, G., Xu, J., Peng, W., Zhang, J., et al. (2020). Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct. Target. Ther.* 5, 8. doi:10.1038/s41392-020-0110-5

Zhou, J., Chen, Q., Zou, Y., Chen, H., Qi, L., and Chen, Y. (2019). Stem cells and cellular origins of breast cancer: Updates in the rationale, controversies, and therapeutic implications. *Front. Oncol.* 9, 820. doi:10.3389/fonc.2019.00820