

The effects of ageing, BMI and physical activity on blood IL-15 levels

Prado, Guilherme Henrique Jesus do; Veiga Sardeli, Amanda; Lord, Janet; Cavaglieri, Cláudia Regina

DOI:

[10.1016/j.exger.2022.111933](https://doi.org/10.1016/j.exger.2022.111933)

License:

Creative Commons: Attribution-NonCommercial-NoDerivs (CC BY-NC-ND)

Document Version

Peer reviewed version

Citation for published version (Harvard):

Prado, GHJD, Veiga Sardeli, A, Lord, J & Cavaglieri, CR 2022, 'The effects of ageing, BMI and physical activity on blood IL-15 levels: a systematic review and meta-analyses', *Experimental gerontology*, vol. 168, 111933. <https://doi.org/10.1016/j.exger.2022.111933>

[Link to publication on Research at Birmingham portal](#)

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Experimental Gerontology

The effects of ageing, BMI and physical activity on blood IL-15 levels: A Systematic Review and Meta-analyses.

--Manuscript Draft--

Manuscript Number:	EXG-D-22-00457R1
Article Type:	Review Article
Section/Category:	Musculoskeletal System and Exercise
Keywords:	aging; Interleukin-15; Exercise; physical activity; BMI
Corresponding Author:	Amanda Veiga Sardeli State University of Campinas: Universidade Estadual de Campinas Campinas, BRAZIL
First Author:	Guilherme Prado
Order of Authors:	Guilherme Prado Amanda Veiga Sardeli Janet Mary Lord Cláudia Regina Cavaglieri
Abstract:	<p>Aim</p> <p>The purpose of the study was to test the effect of ageing and physical activity on IL-15 blood concentration by meta-analyses of the literature.</p> <p>Methods</p> <p>The search was performed in PubMed/MEDLINE, Web of Science, ProQuest, Embase and Cochrane databases. First meta-analysis compared blood IL-15 of healthy adults across three age groups (<35 years, 35-65 years, and >65 years); the second compared IL-15 levels between physically active and non-physically active individuals; the third tested the effect of exercise interventions on blood IL-15 levels on participants of any age, sex, and health condition.</p> <p>Results</p> <p>From 2582 studies retrieved, 67 were selected for the three meta-analyses (age effect: 59; physical activity cross-sectional effect: 5; exercise training effect: 14). Older adults had lower blood IL-15 than young and middle-aged adults (5.30 pg/ml [4.76; 5.83]; 7.11 pg/ml [6.33; 7.88]; 7.10 pg/ml [5.55; 8.65], respectively). However, the subgroup of overweight older adults had higher IL-15 than young and middle aged overweight adults; Habitual physical activity did not affect blood IL-15 (standardized mean difference [SMD] 0.61 [-0.65; 1.88], p=0.34); Exercise intervention reduced blood IL-15 in short-term interventions (<16 weeks) (SMD -0.14 [-0.27; -0.01], p=0.04), but not studies of more than 16 weeks of intervention (SMD 0.44 [-0.26; 1.15], p=0.22).</p> <p>Conclusion</p> <p>The present meta-analyses highlight the complex interaction of age, BMI and physical activity on blood IL-15 and emphasize the need to take these factors into account when considering the role of this myokine in health throughout life.</p>
Suggested Reviewers:	Fabio Lira fabio.lira@unesp.br Ana Maria Teixeira ateixeira@fcdef.uc.pt Quinn LeBris quinnL@u.washington.edu

	Nicole Gruta Nicole.la.gruta@monash.edu
	Charles Lutz ctlutz2@uky.edu
Opposed Reviewers:	
Response to Reviewers:	

Dear Editor and reviewers,

We thank you for spending your time in the evaluation of our manuscript. We have revised our manuscript with changes marked in yellow and our detailed response to each comment is described below.

We are looking forward to your response.

Yours sincerely

Reviewer #1: The present review manuscript is very well write and can be accepted after major review.

Major review

1) Many times the authors write "exercise", "physical activity", or "physical exercise". I understand that these term sound similar, but are differences between their. Please, explain about and adopt just one term.

Authors' reply: Thanks for the comment, we reviewed the use of the different terms, but adopt just one of them would not be possible since we really meant two different types: one meta-analysis was about physical activity effect of IL-15 and the other exercise training interventions. We opted to use chronic exercise interventions (and we deleted all the physical exercise terms), since this term suggests a repetition of more than one exercise session, and we used physically active individuals to reinforce we were talking about groups already trained in cross-sectional studies.

2) The BMI is highlight in title, but not in the aims. Why?

Authors' reply: Thank you for noticing, it should be more clearly included in the first aim. In the beginning of our plan of analysis BMI was just a confounding factor, but since we noticed a very important effect of this variable on IL-15, we opted to give higher importance to it across the manuscript. Now we added it to our first aim.

3) IL-15 exhibit impact to visceral and subcutaneous adipose tissue depots or just in visceral adipose tissue depot? Please, explain about heterogeneous adipose tissue response from IL-15.

Authors' reply: Previously, we did not give much attention to this heterogeneity, and we really appreciate the suggestion, because we found it could be another explanation to our surprising findings. We added the possible influence of these different associations in the 3rd paragraph of the discussion.

4) Explain about differences between the cardiorespiratory capacity and physical activity level, because not similar parameters. A subject might exhibit a higher cardiorespiratory capacity and low physical activity level, or inverse.

Authors' reply: We added a sentence in the discussion of the second meta-analysis explaining the limitation of the use of different ways to access physical activity that could be not a good representation of physical activity but rather differentiate only cardiorespiratory fitness.

Highlights

- IL-15 is an anti-inflammatory myokine that has important metabolic actions such as skeletal muscle anabolism and fat reduction
- Older adults have lower blood IL-15 than young and middle-aged adults
- Within overweight individuals, older adults have higher blood IL-15 than young and middle-aged adults
- There is no consensus about habitual physical activity modulating blood IL-15 levels
- Exercise reduces blood IL-15 in interventions of less than 16 weeks, but not in interventions longer than 16 weeks

The effects of ageing, BMI and physical activity on blood IL-15 levels: A Systematic Review and Meta-analyses.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Abstract

Aim: The purpose of the study was to test the effect of ageing and physical activity on IL-15 blood concentration by meta-analyses of the literature.

Methods: The search was performed in PubMed/MEDLINE, Web of Science, ProQuest, Embase and Cochrane databases. First meta-analysis compared blood IL-15 of healthy adults across three age groups (<35 years, 35-65 years, and >65 years); the second compared IL-15 levels between physically active and non-physically active individuals (cross-sectional studies); the third tested the effect of chronic exercise interventions on blood IL-15 levels on participants of any age, sex, and health condition.

Results: From 2582 studies retrieved, 67 were selected for the three meta-analyses (age effect: 59; physical activity cross-sectional effect: 5; exercise training effect: 14). Older adults had lower blood IL-15 than young and middle-aged adults (5.30 pg/ml [4.76; 5.83]; 7.11 pg/ml [6.33; 7.88]; 7.10 pg/ml [5.55; 8.65], respectively). However, the subgroup of overweight older adults had higher IL-15 than young and middle aged overweight adults; Habitual physical activity did not affect blood IL-15 (standardized mean difference [SMD] 0.61 [-0.65; 1.88], $p=0.34$); Chronic exercise reduced blood IL-15 in short-term interventions (<16 weeks) (SMD -0.14 [-0.27; -0.01], $p=0.04$), but not studies of more than 16 weeks of intervention (SMD 0.44 [-0.26; 1.15], $p=0.22$).

Conclusion: The present meta-analyses highlight the complex interaction of age, BMI and physical activity on blood IL-15 and emphasize the need to take these factors into account when considering the role of this myokine in health throughout life.

Keywords: Aging; Interleukin-15; Exercise; Physical activity; Exercise therapy; BMI.

Introduction

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

The cytokine IL-15 is mainly produced by monocytes and macrophages in response to pathogens and has immunoregulatory effects on a variety of immune cells including the activation of NK cells and T lymphocytes and induction of cytokine production by neutrophils and monocytes (1,2). It is also highly expressed in skeletal muscle (3–5) and thus has also been classified as a myokine with effects on many other tissues such as brain (6), bone (7), skeletal muscle (4,8) and adipose tissue (3). IL-15 has important metabolic actions, stimulating skeletal muscle anabolism and protecting against the negative effects of visceral adiposity (3,4,8,9). Specifically, IL-15 stimulates differentiated myocytes and muscle fibres to increase their contractile protein content and inhibit degradation of muscle proteins contributing to hypertrophy (8,9). IL-15 is the most abundant cytokine in skeletal muscle (10) and the levels of IL-15 in muscle are among the highest of any tissue (11), suggesting muscle activity might be an important regulator of its expression and biological function. IL-15 also prevents lipid deposition in pre-adipocytes and increases glucose uptake by muscle, potentially ameliorating the deleterious effects of white adipose tissue (3,12,13). These effects may, for example, help to prevent fat accumulation in the thymus with aging which leads to thymic atrophy and reduced production of naïve T cells with impact on immunity (14).

As such IL-15 has been proposed as a positive modulator of aging trajectory and age-related diseases such as sarcopenia and diabetes as well as obesity (12,15). Although some studies have shown blood concentrations of IL-15 to be significantly lower in older adults compared to young individuals (16,17), other studies do not report age differences (18,19). One confounding factor among these studies might be the participants differing health status, since some diseases and obesity appear to alter IL-15 in different ways (20–22). Thus, one aim of the present study was to confirm by meta-analysis of the current literature whether advancing age reduces blood IL-15 levels in healthy subjects, **also considering the influence of BMI on ageing effects.**

Chronic exercise is a potential anti-inflammatory therapy to delay or reduce the age-related increase in systemic inflammation, termed inflammageing (23,24). **Different of chronic exercise interventions that involved systematic exercise, physical activity “any bodily movement produced by the contraction of skeletal muscle that increases energy expenditure above a basal level” (25), and few studies have also tested associations between physical activity and blood IL-15 concentrations.** Higher IL-15 in physically active individuals has been reported in cross-sectional studies (26,27), though this is not a universal finding (22). Furthermore, interventional studies have shown more contradictory results, with exercise leading to increase (28), no change (29), and even a reduction (30) of blood IL-15. This lack of consensus may be due to the different health condition among subjects, or the variety of exercise protocols tested. In this way, the additional aims of this study were related to the potential of exercise to regulate blood IL-15 concentration. Specifically, we aimed to test the difference between blood IL-15 concentrations in physically active and non-physically active individuals, and to test blood IL-15 before and after chronic exercise interventions, by meta-analysis of the existing literature.

Methods

1 A systematic search was performed on PubMed, Web of Science,
2 ProQuest, Embase, and Cochrane databases in January 2021. First, the PubMed
3 syntax was exhaustively tested and rebuilt until approved by all reviewers as the
4 following format: ("Aged" [mh] OR "Aging" [mh] OR "Aged 80 and over" [tiab] OR
5 "Frail Elderly" [mh] OR "old" [tiab] OR "ancient" [tiab] OR "old-aged" [tiab] OR
6 "elder" [tiab] OR "aged 60 and over" [tiab] OR "age" OR "year") AND ("Interleukin-
7 15" [mh] OR "IL-15" [tiab] OR "IL15"[tiab] OR "Interleukin 15"[tiab]) AND
8 ("english"[Language] OR "Portuguese"[Language]) NOT ("review"[publication
9 type]). Then, other equivalent searches were planned for each of the other
10 databases (Table S1).

11 The selection of studies by their abstracts was made on Rayyan (31). The
12 studies retrieved after this selection were scrutinized in a spreadsheet and
13 clustered for each of the 3 main analyses. Only articles in English were included
14 and no restriction on publication date was imposed. Participants of both sexes
15 and over 18 years of age were considered.

16 Mean, standard deviation (SD), sample number (n) for each time point and
17 subgroup within studies were used to analyse the blood IL-15 concentration
18 picogram per millilitre (pg/ml). Data not presented as mean and standard
19 deviation (SD), were converted to mean and SD for analysis. The standard error
20 (SE) was converted to SD by the equation $SD = SE * (\sqrt{n})$, if SD was not provided
21 in the original study. When median and interquartile range (IQR) were reported,
22 median was accepted as mean and SD was estimated by equation $(SD = (IQR /$
23 $1.35))$ (32). When numerical data were not available, graphical data were
24 extracted from the image pixels converted in an online software
25 (<https://apps.automeris.io/wpd/>), in which the average of three recordings for
26 each data point was used for analysis.

27 The first meta-analysis included studies assessing blood IL-15
28 concentration in healthy individuals that necessarily reported the age of their
29 participants. The comparison between age groups considered the mean age
30 group of each study (<35, 35-65, ≥65). When the studies did not present an
31 average, we used the median; and in the absence of the median, we calculated
32 the mean between the minimum and maximum age of the participants included
33 in the study. Obese individuals were not excluded; however, the confounding
34 effect of obesity was isolated in further subgroup analysis. Thus, all studies were
35 divided into categories based on the mean BMI reported; being <25 (kg/m²)
36 normal weight, 25-30 (kg/m²) overweight and >30 (kg/m²) obese.

37 The second meta-analysis included studies that compared blood IL-15
38 concentration between physically active individuals and non-physically active
39 individuals, according to each primary study definition.

40 The third meta-analysis included studies assessing blood IL-15
41 concentration before and after a chronic exercise intervention. In this analysis
42 participants in different health conditions were included and compared in further
43 subgroup analysis. For this meta-analysis, characteristics of the chronic exercise

1 **intervention** protocols were also extracted. We categorized the studies according
2 to age (<64 years vs. ≥65 years), sex (men and women), exercise protocol
3 (aerobic, strength and combined strength plus aerobic), and length of intervention
4 (<16 weeks and ≥16 weeks). We also categorized the studies by health condition;
5 and when the study was not designed for a specific disease population, we
6 classified as non-specific diseases, since we could not ensure all of the
7 individuals were healthy.
8
9

10 **Quality of studies**

11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
Studies were not excluded based on their quality, but rather used for descriptive purpose. The Newcastle-Ottawa scale was used to assess the risk of bias regarding selection, comparability, and exposure between physically active and non-physically active groups in each study. The PEDro scale was used to assess the quality of interventional studies (33). 1: Eligibility criteria specified; 2: Random allocation; 3: Concealed allocation; 4: Groups similar at baseline (IL-15 concentration); 5: Subject blinding; 6: Instructor blinding; 7: Assessor blinding; 8: Less than 15% dropouts; 9: Intention-to-treat analysis; 10: Between-group statistical comparisons; 11: Point measures and variability data. Thus, the scores on the PEDro scale ranged from 0 (very low methodological quality) to 10 (high methodological quality). The quality of the studies was used only for qualitative purpose and was not an exclusion criterion.

31 **Statistical analysis**

32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
The three meta-analyses were performed using the Comprehensive Meta-Analysis (CMA) software, version 3.3.070. In the first meta-analysis we calculated the raw mean difference (RMD) of circulating IL-15 (mg/ml) between groups of different age groups (<35, 35-65 and >65). In the second, we calculated the standardized mean difference (SMD) of blood IL-15 between physically active and non-physically active individuals. In the third, SMD was calculated based on the difference between pre- and post-intervention of the training group (a) or the difference in variations (pre-post) between the training group and the control group (b) for studies containing a control group. In the third meta-analysis, we also chose to use SMD, since even using the same measurement units, the studies presented very different mean and SD magnitudes that would lead to bias. Subgroups of participants within each study were treated in the analysis as they were different studies.

53
54
55
56
57
58
59
60
61
62
63
64
65
When there was significant heterogeneity ($p \leq 0.05$), we applied randomized effects and when there was no significant heterogeneity ($p > 0.05$) we applied fixed effects. Publication biases were analysed using the Egger test.

57
58
59
60
61
62
63
64
65
The first meta-analysis was based on the studies mean age (<35, 35-65 and >65) and BMI was also included as confounding factor (normal weight vs. overweight vs. obese).

1 Since there was substantial inconsistency ($I^2 > 50\%$) in the third meta-
2 analysis, further subgroup analyses were run for age, health condition, sex,
3 exercise type and length of intervention. The Q test was used to identify
4 differences between subgroup categories. The p-value ≤ 0.05 was considered
5 significant for all analysis.
6

7 **Evidence quality**

8
9
10 The quality of evidence was assessed by GRADE approach, considering
11 the evaluation items of each study. For the meta-analysis of observational studies
12 (Comparison between IL-15 of physically active and non-physically active) starts
13 with 2 points and one or two points are added according to effect size, dose-
14 response gradient, and positive influence of confounding factors. Details of each
15 quality of evidence analysis will be described in the Results section, step by step.
16 This will lead to a quality of evidence ranging from very low (≤ 1) to high (4).
17
18

19
20 The quality of the evidence for the **chronic** exercise intervention effects on
21 IL-15 was also assessed by GRADE approach but considering the evaluation
22 items for interventional studies. In this type of analysis, we start with 4 points and
23 remove one or two points according to the severity of bias in each item. It will lead
24 to a quality of evidence that ranges from very low (≤ 1) to high.
25
26

27 **Results**

28
29
30 Figure 1 details the selection process of the studies for all meta-analyses.
31 In the Kuczynski et al. study (34) we assumed the data presented were the
32 interquartile range, although not described, since the distribution of IL-15 was
33 described as non-normal and the difference between the maximum and minimum
34 values until the mean was different; In the van der Zij et al. study (35) the control
35 group was not considered because the IL-15 values were below the quantification
36 level. In Chang et al. (16) two groups with participants under the age of eighteen
37 were excluded.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

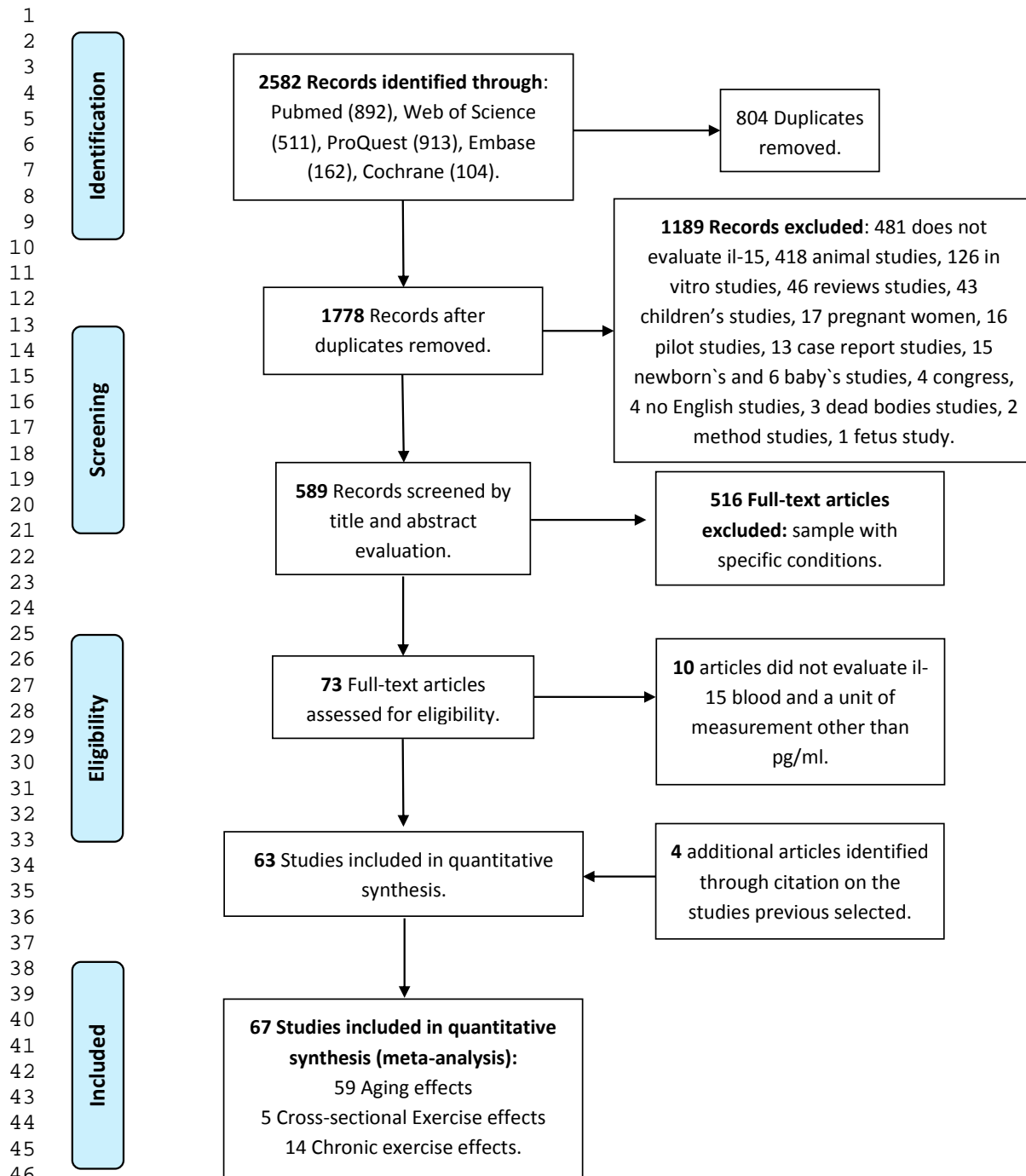


Figure 1. Flow diagram of study selection.

Older adults have lower blood IL-15 than middle aged and young adults, unless they are overweight

Table 1 shows the characteristics of the 59 studies selected to compare the concentration of IL-15 between individuals of different age groups. A hundred and two age subgroups were included in the analysis. Five studies were not included in this meta-analysis due to the different unit of measures used (36–40).

Table 2 shows that individuals >65 years have lower blood IL-15 concentrations compared to the other age groups (<35 years and 35-65). IL-15 was also affected by BMI differences, and this effect was not linearly associated with higher weight, since overweight individuals had higher blood IL-15 concentrations than normal weight or obese individuals (Table 2).

Therefore, isolating age effects within BMI subgroups, we found that lower IL-15 concentrations in older adults were not significantly different from other age categories within normal weight individuals (Table 2). However, within overweight individuals, there was a linear increase in IL-15 with age, showing older adults had significantly higher IL-15 than young adults (Table 2). Unfortunately, almost all the obese individuals were middle aged, and just one study included obese older adults, which limited the understanding of age effects within the obese population. It is noteworthy, that complementary analysis (not shown on Table 2) comparing age effects by categories based on range (without overlapping across age groups), reinforce normal weight older adults had significantly ($p = 0.011$) lower blood IL-15 (2.64 pg/ml [1.79; 3.49]) than young normal weight individuals (4.04 pg/ml [3.68; 4.40]). Within the overweight adults the analysis based on range also showed significantly higher IL-15 ($p < 0.001$) for older adults (11.53 pg/ml [4.48; 18.58]) (30,41–43) than young (3.14 pg/ml [2.77; 3.51]). It is noteworthy that among the analysis of categories based on age range groups, no study was included in the middle-aged adult group and just one study included obese participants exclusively within the older adults age range.

Table 1. Characteristics of the studies included in the healthy individual analysis.

First author, year (Subgroup)	n	Sex	Age (years)	Age category	Body mass Index (kg/m ²)	Body Mass Index category	IL-15 (pg/ml)
Ahn and Kim, 2020 (44) (High normal blood pressure)	12	Both	72.00 ± 3.81	>65	25.27 ± 2.53	overweight	20.93 ± 10.09
Ahn and Kim, 2020 (44) (Normal blood pressure)	18	Both	69.22 ± 4.14	>65	25.23 ± 3.87	overweight	22.39 ± 11.31
Al-Shukaili, 2013 (42)	30	Both	35.00 ± 7.00	<35	NR	NR	10.20 ± 10.30
B, 2020 (35)	65	Both	43.00 ± 11.00	35-65	38.00 ± 5.00	obese	10.30 ± 18.80
Bartlett and Duggal, 2020 (26) (Heathy)	25	Both	67.04 ± 5.97	>65	25.14 ± 3.90	overweight	34.92 ± 7.29
Bartlett and Duggal, 2020 (26) (Sedentary)	25	Both	63.36 ± 4.42	35-65	28.70 ± 4.56	overweight	7.23 ± 9.81
Bazgir, 2015 (27) (Athletes concentric exercise)	14	Men	24.10 ± 2.50	<35	23.80 ± 2.10	normal weight	1.79 ± 0.60
Bazgir, 2015 (27) (Athletes eccentric exercise)	14	Men	24.10 ± 2.50	<35	23.80 ± 2.10	normal weight	1.72 ± 0.40
Bazgir, 2015 (27) (Non-athletes)	14	Men	20.80 ± 2.30	<35	21.60 ± 2.60	normal weight	1.43 ± 0.17
Beavers, 2010 (30) (Training)	182	Both	76.40 ± 4.10	>65	30.70 ± 6.00	obese	1.77 ± 0.56
Beavers, 2010 (30) (Control)	186	Both	77.0 ± 4.40	>65	29.80 ± 5.50	overweight	1.76 ± 0.42
Biancotto, 2013 (43)	144	Both	41.50 ± 20.50	<35	NR	NR	76.08 ± 161.04

16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Bowman, 2018 (45)	102	Both	69.80 ± 7.70	>65	NR	NR	3.00 ± 0.60
Brunelli, 2015 (28) (Control)	17	Men	48.00 ± 1.72	35-65	31.01 ± 3.06	obese	0.78 ± 0.16
Brunelli, 2015 (28) (Training)	13	Men	49.29 ± 1.31	35-65	30.95 ± 0.40	obese	0.45 ± 0.07
Bugera, 2018 (46)	10	Men	25.78 ± 3.56	<35	25.93 ± 2.22	overweight	0.78 ± 0.69
Cassano, 2017 (47)	118	Both	42.10 ± 13.10	35-65	24.99 ± 4.10	normal weight	4.12 ± 8.44
Chang, 2016 (16) (20-39y)	40	Both	30.0 ± 0.80	<35	NR	NR	55.94 ± 5.85
Chang, 2016 (16) (40-59y)	48	Both	51.10 ± 0.80	35-65	NR	NR	56.21 ± 8.21
Chang, 2016 (16) (60-79y)	31	Both	68.3 ± 5.10	>65	NR	NR	37.37 ± 6.04
Choe, 2013 (48)	40	Both	55.40 ± 12.80	35-65	NR	NR	10.40 ± 0.90
Christiansen, 2013 (49) (Men)	16	Men	33.00 ± 11.00	<35	27.60 ± 6.00	overweight	7.00 ± 12.85
Christiansen, 2013 (49) (Women)	15	Women	41.10 ± 6.00	35-65	26.90 ± 5.00	overweight	6.40 ± 6.32
Christiansen, 2013 (49) (Normal weight)	15	Both	32.70 ± 12.00	<35	22.40 ± 2.00	normal weight	7.90 ± 8.39
Christiansen, 2013 (49) (Overweight/ Obese)	16	Both	41.30 ± 4.00	35-65	31.80 ± 3.00	obese	3.40 ± 8.16
Csencsits-Smith, 2016 (50)	15	Both	56.80 ± 9.20	35-65	27.30 ±	overweight	6.33 ± 9.68
Di Renzo, 2010 (51) (Normal weight)	20	Women	27.50 ± 7.50	<35	19.20 ± 6.48	normal weight	4.01 ± 5.00
Di Renzo, 2010 (51) (Obese)	20	Women	27.6 ± 7.40	<35	27.89 ± 20.34	overweight	6.94 ± 7.96
Di Renzo, 2010 (51) (Normal weight)	20	Women	27.70 ± 7.30	<35	22.64 ± 8.09	normal weight	6.64 ± 5.60
Duggal, 2018 (17) (Old)	65	Both	68.50 ± 11.50	>65	NR	NR	10.76 ± 16.09

16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Duggal, 2018 (17) (Young)	52	Both	28.00 ± 8.00	<35	NR	NR	21.58 ± 41.09
Duggal, 2018 (17) (Masters)	119	Both	67.00 ± 12.00	>65	NR	NR	29.50 ± 57.02
Gangemi, 2005 (19) (>95y)	30	Both	>95	>65	NR	NR	3.05 ± 1.41
Gangemi, 2005 (19) (30-59y)	21	Both	44.50 ± 14.50	35-65	NR	NR	1.73 ± 0.50
Gangemi, 2005 (19) (60-89y)	21	Both	74.50 ± 14.50	>65	NR	NR	1.94 ± 1.32
Gokkusu, 2010 (52)	162	Both	62.50 ± 12.20	35-65	20.30 ± 2.00	normal weight	1.12 ± 0.85
Gonza1ez-Reimers, 2011 (53)	13	Both	47.00 ± 9.51	35-65	NR	NR	1.69 ± 0.26
Gullick, 2013 (54)	30	Both	51.00 ± 20.00	35-65	NR	NR	28.10 ± 6.52
Hingorjo, 2018 (55) (<23 BMI)	75	Both	19.37 ± 0.63	<35	19.60 ± 1.95	normal weight	4.04 ± 1.60
Hingorjo, 2018 (55) (>23 BMI)	58	Both	19.37 ± 0.63	<35	27.70 ± 4.18	overweight	3.14 ± 1.44
Hu, 2016 (56) (Men)	45	Men	21.40 ± 1.50	<35	NR	NR	33.40 ± 11.10
Hu, 2016 (56) (Women)	11	Women	37.40 ± 1.50	35-65	NR	NR	10.60 ± 1.80
Jekarl, 2019 (57)	80	Both	70 ±	>65	NR	NR	10.35 ± 12.49
Johansson, 2017 (58)	18	Both	76 ±	>65	23.70 ± 1.85	normal weight	1.91 ± 0.43
Joy, 2016 (59) (Treatment)	11	Men	28.00 ± 5.00	<35	NR	NR	2.21 ± 0.49
Joy, 2016 (59) (Placebo)	14	Men	28.00 ± 5.00	<35	NR	NR	2.15 ± 0.48
Kakumu, 1997 (60)	10	Both	64 ±	35-65	NR	NR	6.70 ± 6.30
Knuiman, 2018 (61)	13	Men	21.20 ± 0.50	<35	22.00 ± 0.20	normal weight	3.21 ± 0.97
Kuczynski, 2005 (34)	22	Both	29.00 ± 4.00	<35	NR	NR	2.90 ± 3.33
Lambert, 2004 (62) (Placebo)	6	Men	64.00 ± 5.30	35-65	21.20 ± 2.90	normal weight	2.02 ± 0.95
Lambert, 2004 (62) (Treatment)	6	Men	66.60 ± 3.70	>65	24.30 ± 1.80	normal weight	1.84 ± 0.68

16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Lambert, 2004 (62) (Placebo)	5	Men	67.00 ± 6.10	>65	22.50 ± 2.80	normal weight	2.02 ± 0.29
Lambert, 2004 (62) (Treatment)	8	Men	66.90 ± 5.50	>65	22.90 ± 2.90	normal weight	2.26 ± 0.50
Lesiak, 2016 (63)	29	Men	51.80 ±	35-65	NR	NR	4.88 ± 0.60
Levinger, 2016 (38)	10	Both	67.40 ± 2.40	>65	28.20 ± 1.70	overweight	31.52 ± 5.00
Lis, 2015 (64) (Athlete with gluten)	13	Both	32.00 ± 7.00	<35	NR	NR	15.17 ± 11.94
Lis, 2015 (64) (Gluten-free athlete)	13	Both	32.00 ± 7.00	<35	NR	NR	12.65 ± 9.98
Luna, 2011 (65) (Training)	11	Men	40.40 ± 7.90	35-65	NR	NR	9.95 ± 79.48
Luna, 2011 (65) (placebo)	14	Men	40.40 ± 7.90	35-65	NR	NR	3.20 ± 14.80
Martinez-Hernandez, 2012 (66)	8	Both	47.10 ± 12.90	35-65	25.20 ± 3.60	overweight	1.75 ± 0.41
Micielska, 2019 (67) (Control)	13	Women	45.00 ± 13.00	35-65	NR	NR	3.29 ± 3.28
Micielska, 2019 (67) (Training)	20	Women	40.00 ± 11.00	35-65	NR	NR	3.98 ± 4.07
Michel Rentzos, 2007 (68)	15	Both	70.00 ± 8.00	>65	NR	NR	2.0 ± 0.22
M Rentzos, 2007 (69)	19	Both	65.8 ± 11.2	>65	NR	NR	3.0 ± 0.44
Minuzzi, 2019 (18) (Masters)	20	Both	53.10 ± 8.80	35-65	25.10 ± 4.60	overweight	24.32 ± 9.66
Minuzzi, 2019 (18) (Young)	9	Both	31.80 ± 3.00	<35	21.80 ± 2.00	normal weight	1.71 ± 2.16
Minuzzi, 2019 (18) (Old)	10	Both	54.20 ± 5.90	35-65	24.30 ± 3.20	normal weight	2.97 ± 3.98
Mustafa, 2015 (70)	23	Both	28.00 ±	<35	NR	NR	0.00 ± 9.40
Nathella, 2017 (71)	66	Both	41.00 ± 19.00	35-65	NR	NR	5.05 ± 6.74
Nishida, 2014 (72) (Men)	737	Men	57.30 ± 8.10	35-65	23.90 ± 2.90	normal weight	11.73 ± 1.42

23	Nishida, 2014 (72) (Women)	1838	Women	56.30 ± 8.10	35-65	22.60 ± 3.10	normal weight	11.49 ± 1.49
24	Nishida, 2015 (73) (Training)	31	Both	70.40 ± 5.80	>65	24.20 ± 3.70	normal weight	2.91 ± 0.50
26	Nishida, 2015 (73) (Control)	31	Both	69.70 ± 6.60	>65	22.50 ± 2.50	normal weight	3.36 ± 3.11
28	Notarnicola, 2015 (74)	19	Both	45.80 ± 1.80	35-65	NR	NR	0.41 ± 2.96
30	Oliver, 2016 (40)	10	Both	27.00 ± 4.00	<35	NR	NR	1.06 ± 0.23
32	Olszewski, 2001 (75)	20	Both	25.00 ± 3.00	<35	NR	NR	0.30 ± 0.01
33	Pérez-López, 2018 (22) (Lean physical activity)	25	Both	46.00 ± 4.10	35-65	23.90 ± 3.10	normal weight	1.85 ± 0.40
37	Pérez-López, 2018 (22) (Lean non-physically active)	28	Both	47.00 ± 3.90	35-65	24.20 ± 3.50	normal weight	2.70 ± 0.50
40	Pérez-López, 2018 (22) (Obese physical active)	64	Both	44.40 ± 8.80	35-65	37.40 ± 4.70	obese	2.50 ± 0.60
43	Pérez-López, 2018 (22) (Obese non-physically active)	79	Both	44.30 ± 8.10	35-65	38.0 ± 6.00	obese	3.30 ± 0.70
46	Phenekos, 2004 (76)	25	Both	52.30 ± 12.80	35-65	NR	normal weight	2.03 ± 0.18
48	Prestes, 2009 (29)	35	Women	63.18 ± 4.8	35-65	NR	normal weight	31.38 ± 70.16
49	Ragino, 2020 (77)	50	Both	35.00 ± 6.88	<35	22.90 ± 3.40	normal weight	2.10 ± 2.37
51	Rinnov, 2014 (78) (Training)	10	Men	30.50 ± 5.50	<35	25.10 ± 2.10	normal weight	0.89 ± 0.66
53	Rinnov, 2014 (78) (Control)	5	Men	25.20 ± 3.30	<35	23.20 ± 2.40	normal weight	1.12 ± 0.31
55	Sánchez-Jiménez (79)	46	Both	46.30 ± 15.10	35-65	26.40 ± 4.40	overweight	4.00 ± 2.50
57	Shammam, 2015 (80)	70	Both	33.00 ± 7.83	<35	NR	NR	34.92 ± 9.03
58	Shibatomi, 2001 (81)	33	Both	44.70 ± 12.70	35-65	NR	NR	2.74 ± 1.02

15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Siewko, 2019 (20)	60	Both	38.50 ± 31.11	35-65	22.70 ± 13.55	normal weight	0.20 ± 2.51
Tagoma, 2019 (82)	42	Women	29.20 ± 10.70	<35	NR	NR	72.10 ± 63.55
Urzi, 2019 (55) (Control)	9	Women	88.90 ± 5.30	>65	29.10 ± 5.10	overweight	8.37 ± 3.26
Urzi, 2019 (55) (Training)	11	Women	84.40 ± 7.70	>65	28.00 ± 5.50	overweight	6.25 ± 3.14
Uzawa, 2014 (83)	20	Both	48.5 ± 23.50	35-65	NR	NR	4.42 ± 1.55
Xia, 2011 (84)	45	Both	46.60 ± 13.40	35-65	NR	NR	264.31 ± 161.73
Yalcin, 2018 (41)	80	Both	75.00 ±	>65	29.05 ± 39.00	overweight	5.10 ± 35.81
Yang, 2017 (85)	22	Both	31.10 ± 7.10	<35	NR	NR	81.76 ± 50.00
Zhu, 2016 (86)	25	Both	32.20 ± 9.04	<35	NR	NR	19.50 ± 124.95

Note: Data are presented as mean ± SD. IL-15: mean standard deviation of IL-15 in pg/ml; y: year; SD: standard deviation; NR: Not reported.

Table 2. BMI and age subgroup analysis.

Subgroup	K	N	References	Mean	LL	UL	I ²	P-value (between)
Age								
<35y	33	962	(16–18,27,34,40,42,43,46,49,51,56,59,61,64,70,75,77,78,80,82,85–87)	7.11	6.33	7.88	99.58*	<0.001
35–65y	40	3.870	(16,18–20,22,26,28,29,35,47–50,52–54,56,60,62,63,65–67,71,73,76,79,81,83,84,88)	7.10	5.55	8.65	99.97*	
>65y	23	1.120	(16,17,19,26,30,41,44,45,55,57,58,62,68,73,89,90)	5.30	4.76	5.83	99.08*	
BMI								
Normal Weight	27	3.342	(18,20,22,27,47,49,51,52,58,61,62,72,73,77,78,87)	3.27	1.26	5.28	99.96*	<0.001
Overweight	17	574	(18,26,30,41,44,46,49–51,55,66,79,87)	8.28	7.04	9.52	98.27*	
Obese	7	436	(22,28,30,35,49)	2.07	1.22	2.92	99.75*	
Age within normal weight								
Young	12	259	(18,27,49,51,61,77,78,87)	2.32	1.78	2.85	96.10*	0.573
Middle-aged	9	2984	(18,20,22,47,52,62,72)	4.25	0.67	7.84	99.98*	
Older adults	6	99	(58,62,73)	2.31	1.85	2.77	92.97*	
Age within overweight								
Young	4	104	(30,44,46,55)	3.28	1.22	5.35	96.07*	0.005
Middle-aged	6	129	(26,49,50,66,79)	7.62	4.76	10.48	96.76*	
Older adults	7	341	(26,41,44,55)	14.24	5.87	22.60	99.12*	
Age within obese								
Young	-	-	-	-	-	-	-	0.427
Middle-aged	6	182	(22,28,35,49)	2.18	1.17	3.19	99.73*	
Older adults	1	254	(30)	1.77	1.69	1.85	0.00	

Note: y: years; K: number of studies; N: number of individuals; LL: lower limit; UL: upper limit; *: Significant heterogeneity (p<0.05).

No difference in blood IL-15 between physically active and non-active adults

The characteristics of the five studies (6 subgroups) comparing physically active and non-active individuals are detailed in Table 3. The level of physical activity was classified in different ways across the studies. The studies considered physically active those who undertook regular physical activity for at least six months, or had higher number of steps per day, or higher VO₂max, or had a proven specific intensity or volume of exercise in each bout or in each week.

There was no difference between blood IL-15 of physically active individuals and non-physically active individuals (Figure 2). In general, the studies showed a low risk of bias, with their quality ranging from 5 to 6 in the New Castle Ottawa scale (Table S2). We evaluated the quality of evidence as low (GRADE score=2), due to the low magnitude of difference between groups, and no clear dose-response effect or positive influence of confounding factors. Within each age category (middle aged and older adults) there was also no difference between groups (data not shown).

Table 3. Characteristics of the studies is a cross-sectional analysis.

First author, year (Subgroup)	n (CO)	Sex	Trained age (CO)	Training time	Physical activity level (CO)	Physically active BMI (kg/m ²)	Control BMI (kg/m ²)	Physically active IL-15 (pg/ml)	Control IL15 (pg/ml)
Bartlett, 2020 (26)	25 (25)	Both	67.04 ± 5.97 (63.36 ± 4.42)	NR	Steps day 10.500 - 15.000 (2.000 - 4.000)	25.14 ± 3.90	28.70 ± 4.56	34.92 ± 7.29	7.23 ± 9.81
Bazgir, 2015 (27)	14 (14)	Men	24.10 ± 2.50 (20.80 ± 2.30)	NR	PA > 6 months (No regular exercise)	23.80 ± 2.10	21.60 ± 2.60	1.79 ± 0.60	1.43 ± 0.17
Duggal, 2018 (17)	125 (75)	Both	67.00 ± 12.00 (68.50 ± 11.50)	NR	Cycle 60-100 km in 5.5 - 6.5 hours (Adults who did not practice regular physical activity)	NR	NR	29.50 ± 57.02	10.76 ± 16.09
Minuzzi, 2019 (18)	20 (10)	Both	53.10 ± 8.80 (54.20 ± 5.90)	24 years	VO ₂ max 40.33 ± 11.15 (29.29 ± 4.14)	25.10 ± 4.60	24.30 ± 3.20	24.32 ± 9.66	2.97 ± 3.98
Pérez-López, 2018 (22) (lean)	25 (28)	Both	46.00 ± 4.10 (47.00 ± 3.90)	NR	≥180 min/week (<180 min/week)	23.90 ± 3.10	24.20 ± 3.50	1.85 ± 0.40	2.70 ± 0.50
Pérez-López, 2018 (22) (obese)	64 (79)	Both	44.40 ± 8.80 (44.30 ± 8.10)	NR	≥180 min/week (<180 min/week)	37.40 ± 4.70	38.00 ± 6.00	2.50 ± 0.60	3.30 ± 0.70

Note: CO: control; NR: not reported.

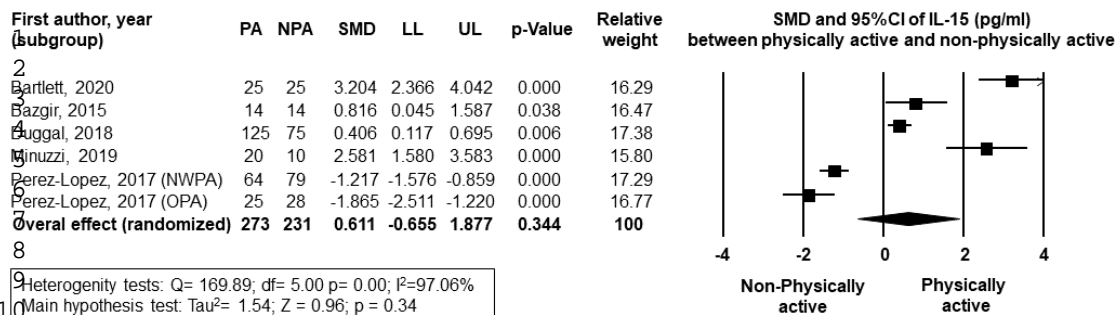


Figure 2. Forest plot of meta-analysis 2: difference between long-term trained and untrained individuals in the blood concentration of IL-15. PA: physically active; NPA: non-physically active; LL:

lower limit; UL: upper limit; NWPA: normal weight physically active; OPA: obese physically active.

Effect of chronic exercise intervention on blood IL-15

Fourteen studies (18 subgroups) were included in the third meta-analysis (Table 4). Most studies included older adults, some included middle-aged participants, and only two included young participants. Most participants were female or both sexes, with only three studies including only males. Some studies have included postmenopausal women or individuals with comorbidities such as prehypertension, obesity, cancer, type 2 diabetes mellitus, mild cognitive impairment and chronic obstructive pulmonary disease. As for the chronic exercise protocols, they varied between aerobic, strength and combined, with medium to high intensities. Most studies included in this analysis presented low quality due to lack of concealed allocation of groups, blinding of participants, instructors and evaluators, high drop-out rates without intention-to-treat analysis, varying their PEDro score from 1 to 5 points (Table S3).

Figure 3 shows that chronic exercise intervention did not change the blood IL-15 concentration. The GRADE approach suggested low quality of evidence (GRADE score = 2), based on (1) high inconsistency ($I^2=77.65$); (2) poor quality of original studies. We did not remove any point of indirect evidence, since all tested the effects of the intervention, with most of them including a control group for comparisons, there was no publishing bias (Egger test p -value > 0.05); and no imprecision (sample size training group: 572 and sample size control group:363).

Table 5 shows the subgroup analysis of the intervention studies. Analysis of individual characteristics shows that individuals aged ≥ 65 years reduced the blood concentration of IL-15 with training, while younger individuals did not. The health status between healthy and unhealthy individuals and the difference between men and women are not determinants for the increase in the blood concentration of IL-15 with training (Table 5). Analysis of the characteristics of the chronic exercise interventions showed that interventions with <16 weeks duration reduced IL-15, whereas intervention ≥ 16 weeks did not. The weekly frequency and type of exercise protocol did not affect IL-15 concentration (Table 5).

Table 4. Characteristics of the intervention analysis studies.

First author, year (Subgroup)	Age	Health condition	Sex	Chronic exercise group			Control group			Protocol	Weekly frequency	Length of Intervention	Length of Inte. category
				IL-15 pre	IL-15 post	N	IL-15 pre	IL - 15 post	N				
Am and Kim, 2020 (44)	72.00 ± 3.81	PH	Women	20.93 ± 10.09	17.34 ± 6.82	12	22.39 ± 11.31	17.59 ± 8.75	18	CT	3	24 Weeks	>16
Banitalebi, 2019 (91) (CT)	54.14 ± 5.43	DM2	Women	3.81 ± 0.23	3.60 ± 0.25	14	3.86 ± 0.25	3.76 ± 0.29	14	CT	3	10 Weeks	<16
Banitalebi, 2019 (91) (Sprint)	55.36 ± 5.94	DM2	Women	3.94 ± 0.23	3.71 ± 0.38	14	3.86 ± 0.25	3.76 ± 0.29	14	AT	3	10 Weeks	<16
Beavers, 2010 (30)	76.40 ± 4.10	Healthy	Both	1.77 ± 0.56	1.72 ± 0.42	182	1.76 ± 0.42	1.80 ± 0.54	186	CT	2-3	24 Weeks	≥16
Brunelli, 2015 (28)	49.29 ± 1.31	OBS	Men	0.45 ± 0.07	1.17 ± 0.10	13	0.78 ± 0.16	0.55 ± 0.08	17	CT	3	24 Weeks	≥16
Gomez, 2011 (92)	50.00 ± 5.00	CA	Women	41.10 ± 25.10	32.30 ± 13.60	8	35.60 ± 10.60	55.40 ± 25.20	8	CT	3	8 Weeks	<16
Lambert, 2004 (62) (PRT)	67.00 ± 6.10	Healthy	Men	2.02 ± 0.29	2.79 ± 0.58	5	2.02 ± 0.95	2.97 ± 0.71	6	RT	3	12 Weeks	<16
Lambert, 2004 (62) (TRT)	66.90 ± 5.50	Healthy	Men	2.26 ± 0.50	2.51 ± 0.76	8	1.84 ± 0.68	2.59 ± 0.83	6	RT	3	12 Weeks	<16

16															
17															
18															
19															20
20															
21	Macielska,	40.00	Healthy	Women	3.98 ±	3.50 ± 5.36	20	3.29 ±	3.21 ±	13	AT	3	5 Weeks	<16	
22	2019 (67)	±			4.07			3.28	1.83						
23		11.00													
24															
25	Nishida,	70.40	Healthy	Women	NR	0.12 ± 0.62*	31	NR	-0.41 ±	NR	CT	NR	12 Weeks	<16	
26	2015 (73)	± 5.80							3.02*						
27															
28	Prestes,	63.18	PM	Women	31.38 ±	54.90 ±	35	NR	NR	NR	RT	2	16 Weeks	≥16	
29	2009 (29)	± 4.80			70.16	198.83									
30															
31	Riechman,	21.00	Healthy	Both	1.67 ±	1.64 ± 0.55	124	NR	NR	NR	RT	3	10 Weeks	<16	
32	2004 (93)	± 2.4			0.53										
33															
34	Rimov,	30.50	Healthy	Both	0.89 ±	1.14 ± 0.53	10	1.12 ±	1.09 ±	5	AT	5	12 Weeks	<16	
35	2014 (78)	± 5.50			0.66			0.31	0.29						
36															
37	Tsai, 2019	66.00	MCI	Both	0.70 ±	0.62 ± 1.56	19	0.81 ±	1.74 ±	18	AT	3	16 Weeks	≥16	
38	(94)	± 7.68			1.48			1.52	1.57						
39															
40	Tsai, 2019	65.44	MCI	Both	0.30 ±	1.03 ± 1.48	18	0.81 ±	1.74 ±	18	RT	3	16 Weeks	≥16	
41	(94)	± 6.76			1.48			1.52	1.57						
42															
43															
44	Silva, 2018	64.88	COPD	Both	89.11 ±	65.80 ±	16	NR	NR	NR	RT	3	12 Weeks	<16	
45	(95) (MG)	±11.17			100.30	42.23									
46															
47	Silva, 2018	69.35	COPD	Both	57.73 ±	42.75 ±	32	NR	NR	NR	RT	3	12 Weeks	<16	
48	(95) (EG)	± 8.97			33.12	32.91									
49															
50	Urzi, 2019	84.40	Healthy	Women	6.25 ±	7.75 ± 3.34	11	8.37 ±	8.63 ±	9	RT	3	12 Weeks	<16	
51	(55)	± 7.70			3.14			3.26	3.41						
52															
53															
54															
55															
56															
57															
58															
59															
60															
61															
62															
63															
64															
65															

Note: N: individuals number; TA: trained age; PRT: placebo resistance training; CT: combined training; TRT: testosterone resistance training; IL-15 pre: interleukin 15 pre **chronic exercise intervention**; IL-15 post: interleukin 15 post **chronic exercise intervention**; T: trained; CO: control; MG: machine group; EG: elastic group; PH: pre hypertensive; OBS: obese; CA: cancer; DM2: diabetes mellitus 2; MCI: mild Cognitive Impairment; COPD: chronic obstructive pulmonary disease; PM: post menopause; RT: resistance training; AT: aerobic training; CT: combined training; *: dates in delta of change; NR: not reported.

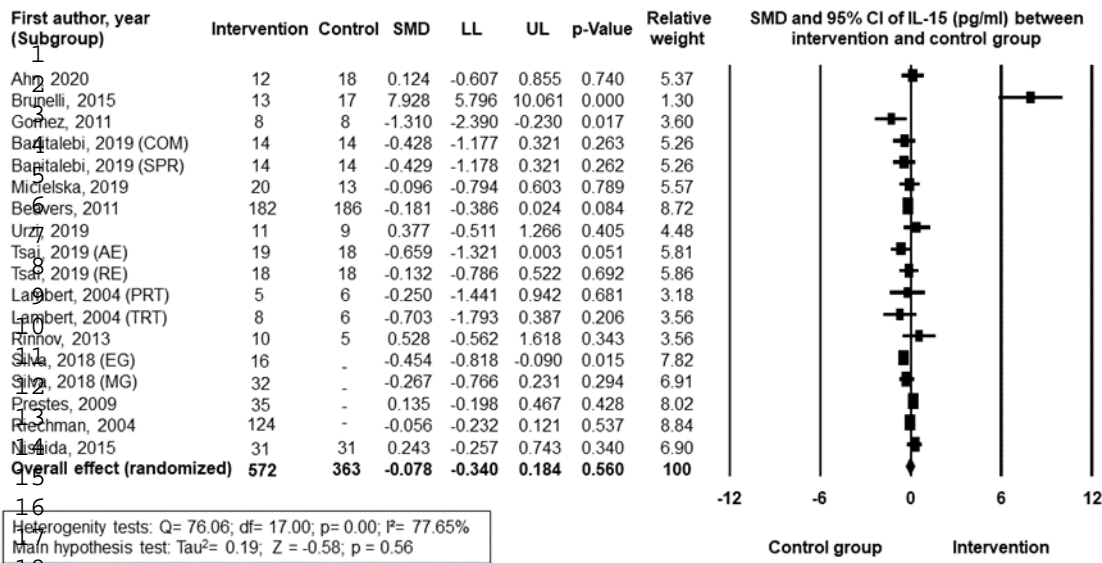


Figure 3. Forest plot of the effect of chronic exercise intervention on the IL-15 concentration. LL: lower limit; UL: upper limit; COM: combined exercise; SPR: sprint exercise; AE: aerobic exercise; RE: resistance exercise; PRT: placebo resistance training; TRT: resistance training treatment; EG: elastic group; MG: machine group.

Table 5. Subgroup analyses of the intervention with chronic exercise intervention in the blood concentration of IL-15.

Subgroups	K	N	T	N	CO	References	SMD	LL	UL	P-value (within)	I ²	P-value (between)
Age												
<64y	9	254	71			(28,29,66,67,78,91,93,95)	0.13	-0.39	0.66	0.62	87.50%*	0.26
≥65y	9	318	292			(30,44,55,62,73,95,96)	-0.19	-0.34	-0.04	0.01	20.01%	
Health condition												
Non-specific diseases	8	391	256			(30,55,62,67,73,78,93)	-0.08	-0.20	0.04	0.21	0.00%	0.78
Specific diseases	10	181	107			(28,29,44,66,91,95,96)	0.00	-0.54	0.54	1.00	86.96%*	
Sex												
Men	3	26	29			(28,62)	2.21	-1.89	6.31	0.29	96.21%*	0.38
Women	8	145	107			(29,44,55,67,73,91,97)	0.00	-0.21	0.21	0.98	33.80%	
Type of exercise												
Aerobic	4	63	50			(67,78,91,96)	-0.29	-0.67	0.09	0.13	20.69%	0.31
Combined	6	260	274			(28,30,44,66,73,91)	0.50	-0.40	1.41	0.27	92.04%*	
Resistance	8	249	39			(29,55,62,93,95,96)	-0.10	-0.23	0.03	0.14	18.33%	
Length of intervention												
<16 wk	12	293	106			(55,62,66,67,73,78,91,93,95)	-0.14	-0.27	-0.01	0.04	29.97%	0.09
≥16 wk	6	279	257			(28–30,44,96)	0.44	-0.26	1.15	0.22	91.66%*	

Note: T: exercise trained; CO: control; K: number of studies; N: number of individuals; LL: lower limit; UL: upper limit; y: years; wk: weeks; *: Significant heterogeneity (p<0.05).

Discussion

The main findings of the present study were that: (1) older adults had lower circulating levels of IL-15 than young and middle-aged adults; however, the overweight subgroup of older adults have higher IL-15 than young and middle-aged overweight adults; (2) cross-sectional studies did not show significant effects of habitual physical activity on IL-15; and (3) chronic exercise intervention reduces IL-15 in subgroups of studies with less than 16 weeks of interventions and in older adults above 65 years.

Although we explored age effects on blood IL-15, we did this by an indirect subgroup analysis instead of a direct within studies comparison meta-analysis. Overall, there was significantly lower IL-15 in older adults than young and middle-aged adults in a very heterogeneous meta-analysis. This agrees with previous studies in animals (12,98). We assumed this is a negative effect of ageing, as in general IL-15 function is reported to be beneficial. For example, ultra-long-lived individuals (>95 y) have significantly higher IL-15 than young or older adults (19). Since inflammaging is highly associated with health span and life span (99), higher IL-15 in long lived individuals might be a signal of protection to reach advanced ages. It also makes biological sense as IL-15 has many effects that could be considered anti-ageing; thus, IL-15 contributes to the maintenance of naive T-cell populations by promoting survival and expansion of naive T cells, without leading to telomere shortening (17,100). Among other IL-15 roles, it has been shown to increase insulin sensitivity in animals (101), improve glucose uptake by skeletal muscle cells in vitro (2) and improve mitochondrial oxidative functions in humans (102) as well as reducing lipid synthesis in adipocytes, increasing lipid metabolism, and reducing white adipose tissue in rats (101).

The age effects observed in the overall analysis were largely comprised of studies including normal weight individuals and subgroup analysis showed that overweight older adults had higher IL-15 than overweight young and middle-aged. We speculate that the cumulative effect of being overweight over many years could lead to impaired sensitivity to IL-15, which in turn would trigger an exacerbated production to compensate, similar to what is known for insulin and leptin resistance (103–105). In fact, the effect of IL-15 on reducing white adipose tissue and improving lipid metabolism in healthy wild obese rats was abolished in leptin deficient obese mice (106). Another explanation could be a more pathological type of increase in fat mass of the younger overweight individuals, increasing mainly visceral fat (107) that is negatively associated to IL-15 concentration (3), compared to a more physiological and well distributed type of increase in fat across different tissues with ageing (108), that won't be strongly correlated to IL-15 decrease (3). Nevertheless, the effect of obesity and ageing on IL-15 is still to be clarified, considering here obese individuals had lower IL-15 than normal weight and overweight, while overweight had higher IL-15 than normal weight individuals.

1 We were not able to confirm any effects of physical activity on IL-15 in the
2 meta-analysis of these very heterogeneous cross-sectional studies. However, we
3 noted that physically active older adults had higher IL-15 than non-physically
4 active older adults (17,18,26); while the studies with younger individuals that are
5 expected to have naturally higher IL-15 without physical activity (22) showed the
6 physically active individuals had lower IL-15 than their counterparts. Indeed,
7 Duggal et al. (17) did not find significant differences between IL-15 of healthy
8 older adults compared to the young adults and it was the highly active master
9 cyclists in this study that displayed higher IL-15 (17). The confounding factors
10 such as age and BMI causing heterogeneity between studies need to be further
11 explored in future meta-analysis, with more studies and diversified population. It
12 is noteworthy, that the cardiorespiratory fitness, assessed by VO₂max in one
13 study (18) does not directly represent physical activity levels, although VO₂max
14 is improved by some types of physical activities, especially the vigorous ones
15 (109). Likely, the higher VO₂max observed in Minuzzi et al. (18) is dependent of
16 very higher physical activity levels in their population since they included master
17 athletes and thus it may justify, the higher IL-15 SMD in this study.

18 There was also contrasting effects in the meta-analysis of chronic exercise
19 intervention studies. In the studies with individuals above 65 years in this meta-
20 analysis, the IL-15 was reduced after training in contrast to the effects observed
21 in cross-sectional studies. Also, reduction of IL-15 was seen in interventions with
22 less than 16 weeks. These findings are hard to reconcile with the high expression
23 of IL-15 in skeletal muscle and its classification as a myokine would suggest it
24 would increase in exercising muscle, but chronic exercise intervention does not
25 always lead to increase in IL-15 (4,5,110) and when it occurs it seems to be
26 pulsatile and short lasting (110).

27 To ensure the robustness of our results, we ran a sensitivity analysis to
28 test chronic exercise intervention effects only in controlled studies, and it led to
29 the same results of the analysis with all studies (SMD 0.13 [0.38; 0.64], p=0.61).
30 The other characteristics of samples or of the interventions seemed not to
31 influence IL-15 levels.

32 The effect of acute increases in IL-15 on long-term IL-15 baseline
33 concentration remains to be confirmed (29). While some chronic aerobic exercise
34 interventions do not affect IL-15 concentration (111), some chronic resistance
35 exercise interventions have shown significant up-regulation of IL-15 expression
36 (93,112). The difference between these aerobic and resistance protocols could
37 be due to their type of muscle fibre recruited, since in general, one would expect
38 acute resistance exercise to recruit more type II fibre than aerobic and IL-15
39 mRNA level is enhanced in skeletal muscles dominated by type II fibres with
40 acute resistance exercise (4).

41 Perspective

42 The integration of previous studies with meta-analyses considerably
43 increased our perspective on the effects of ageing and physical activity on IL-15
44 regulation and challenged previous notions that this anti-inflammatory cytokine
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1 would be higher in those with better health. The first new finding was that IL-15
2 is not consistently reduced in older adults; and, considering only overweight
3 Individuals, IL-15 is higher in the older adults. In this way, future studies should
4 investigate the potential of increased IL-15 as a compensatory mechanism,
5 intended to increase muscle mass, reduce fat mass, and improve immune
6 response, when stimulated by specific conditions such as obesity. The second
7 unexpected finding was the reduction of IL-15 with short training interventions,
8 that could be associated with a lower need for this compensatory mechanism.
9 This finding also reinforced the transient nature of the IL-15 response since longer
10 duration interventions did not change IL-15. This also highlights the need for more
11 studies exploring the exact role of IL-15 following exercise training.
12
13
14
15
16
17

18 Limitations

19
20 This work has some limitations. To identify age effects on blood IL-15 we
21 did it by subgroup analysis based on studies that did not compare defined age
22 groups. However, the age effects were clearly similar in the two different
23 approaches we used, considering exclusive age groups without overlap and the
24 huge total number of studies with some overlap.
25

26 The non-linear effects of obesity on IL-15 could be caused by different
27 percentage of muscle mass between individuals, since muscle is a major
28 producer of IL-15; however, most studies did not report the volume of muscle
29 mass, precluding further analysis.
30

31 Additionally, although we clarify part of exercise effects on IL-15, the
32 quality of evidence in both cross-sectional and interventional meta-analysis were
33 low and very low, respectively. Here, we comprehensively identified potential
34 confounding factors of IL-15 changes with exercise, such as length of
35 intervention, age and BMI, and future controlled studies will be necessary to
36 confirm these findings.
37
38
39

40 Conclusion

41
42 We confirmed that older adults (> 65 years) have lower circulating IL-15
43 concentrations than young and middle-aged individuals; however, it seems to be
44 an effect that occurs only in normal weight individuals. On the other hand,
45 overweight older adults had higher IL-15 than young adults within overweight
46 individuals. Both physical activity and chronic exercise effects on IL-15 were not
47 confirmed by the meta-analyses, and their high heterogeneity and low quality
48 suggest more studies will be needed to clarify the impact on IL-15. Among other
49 potential confounding factors, the time of intervention and age of participants
50 deserves further investigation, since we identified shorter exercise interventions
51 and interventions in older adults reduced IL-15. Given the importance of IL-15 for
52 muscle, immune system and adipose tissue health, understanding the effect of
53 lifestyle factors, such as exercise and general body weight control, will be
54 fundamental to improve interventions for older adult's health.
55
56
57
58

59 References

60
61
62
63
64
65

1. Ye J. Beneficial metabolic activities of inflammatory cytokine interleukin 15 in obesity and type 2 diabetes. Vol. 9, *Frontiers of Medicine*. Higher Education Press; 2015. p. 139–45.
2. Patidar M, Yadav N, Dalai SK. Interleukin 15: A key cytokine for immunotherapy. *Cytokine and Growth Factor Reviews* [Internet]. 2016;31:49–59. Available from: <http://dx.doi.org/10.1016/j.cytogfr.2016.06.001>
3. Nielsen AR, Hojman P, Erikstrup C, Fischer CP, Plomgaard P, Mounier R, et al. Association between interleukin-15 and obesity: Interleukin-15 as a potential regulator of fat mass. *Journal of Clinical Endocrinology and Metabolism*. 2008;93(11):4486–93.
4. Nielsen AR, Mounier R, Plomgaard P, Mortensen OH, Penkowa M, Speerschneider T, et al. Expression of interleukin-15 in human skeletal muscle - Effect of exercise and muscle fibre type composition. *Journal of Physiology*. 2007;584(1):305–12.
5. Hemingway E, Charles Scribner's Sons. *For whom the bell tolls*. 471 p.
6. Shi SX, Li YJ, Shi K, Wood K, Ducruet AF, Liu Q. IL (interleukin)-15 bridges astrocyte-microglia crosstalk and exacerbates brain injury following intracerebral hemorrhage. *Stroke*. 2020;967–74.
7. Li G, Zhang L, Wang D, AlQudsy L, Jiang JX, Xu H, et al. Muscle-bone crosstalk and potential therapies for sarco-osteoporosis. *JOURNAL OF CELLULAR BIOCHEMISTRY*. 2019;120(9):14262–73.
8. Quinn LBS, Strait-Bodey L, Anderson BG, Argilés JM, Havel PJ. Interleukin-15 stimulates adiponectin secretion by 3T3-L1 adipocytes: Evidence for a skeletal muscle-to-fat signaling pathway. *Cell Biology International*. 2005;29(6):449–57.
9. Inoue S, Unsinger J, Davis CG, Muenzer JT, Ferguson TA, Chang K, et al. IL-15 Prevents Apoptosis, Reverses Innate and Adaptive Immune Dysfunction, and Improves Survival in Sepsis. *The Journal of Immunology*. 2010;184(3):1401–9.
10. Nieman DC, Davis JM, Henson DA, Walberg-Rankin J, Shute M, Dumke CL, et al. Carbohydrate ingestion influences skeletal muscle cytokine mRNA and plasma cytokine levels after a 3-h run. *Journal of Applied Physiology*. 2003;94(5):1917–25.
11. Grabstein KH, Eisenman J, Shanebeck K, Rauch C, Srinivasan S, Fung V, et al. Cloning of a T cell growth factor that interacts with the β chain of the interleukin-2 receptor. *Science* (1979). 1994;264(5161):965–8.
12. Quinn LS, Anderson BG, Strait-Bodey L, Wolden-Hanson T. Serum and muscle interleukin-15 levels decrease in aging mice: correlation with declines in soluble interleukin-15 receptor alpha expression. *Exp Gerontol*. 2010 Feb;45(2):106–12.
13. So B, Kim HJ, Kim J, Song W. Exercise-induced myokines in health and metabolic diseases. *Integrative Medicine Research*. 2014;3(4):172–9.

14. Billard MJ, Gruver AL, Sempowski GD. Acute endotoxin-induced thymic atrophy is characterized by intrathymic inflammatory and wound healing responses. *PLoS ONE*. 2011;6(3).
15. Ye J. Beneficial metabolic activities of inflammatory cytokine interleukin 15 in obesity and type 2 diabetes. *Frontiers of Medicine*. 2015;9(2):139–45.
16. Chang WS, Kim EJ, Lim YM, Yoon D, Son JY, Park JW, et al. Age-related changes in immunological factors and their relevance in allergic disease development during childhood. *Allergy, Asthma and Immunology Research [Internet]*. 2016;8(4):338–45. Available from: <https://www.embase.com/search/results?subaction=viewrecord&id=L611153409&from=export>
17. Duggal NA, Pollock RD, Lazarus NR, Harridge S, Lord JM. Major features of immunosenescence, including reduced thymic output, are ameliorated by high levels of physical activity in adulthood. *Aging Cell*. 2018;17(2).
18. Minuzzi LG, Chupel MU, Rama L, Rosado F, Munoz VR, Gaspar RC, et al. Lifelong exercise practice and immunosenescence: Master athletes cytokine response to acute exercise. *CYTOKINE*. 2019 Mar;115:1–7.
19. Gangemi S, Basile G, Monti D, Merendino RA, di Pasquale G, Bisignano U, et al. Age-related modifications in circulating IL-15 levels in humans. *Mediators Inflamm*. 2005 Aug;2005(4):245–7.
20. Siewko K, Maciulewski R, Zielinska-Maciulewska A, Poplawska-Kita A, Szumowski P, Wawrusiewicz-Kurylonek N, et al. Interleukin-6 and Interleukin-15 as Possible Biomarkers of the Risk of Autoimmune Diabetes Development. Fang Y, editor. *BioMed Research International [Internet]*. 2019;2019:7. Available from: <https://search.proquest.com/scholarly-journals/interleukin-6-15-as-possible-biomarkers-risk/docview/2312467371/se-2?accountid=8113>
21. Kacani L, Stoiber H. Role of IL-15 in HIV-1-associated hypergammaglobulinaemia. 1997;14–8.
22. Pérez-López A, Valadés D, Vázquez Martínez C, de Cos Blanco AI, Bujan J, García-Honduvilla N. Serum IL-15 and IL-15R α levels are decreased in lean and obese physically active humans. *Scandinavian Journal of Medicine and Science in Sports*. 2018;28(3):1113–20.
23. Sardeli AV, Tomeleri CM, Cyrino ES, Fernhall B, Cavaglieri CR, Chacon-Mikahil MPT. Effect of resistance training on inflammatory markers of older adults: A meta-analysis. *Experimental Gerontology*. 2018;111(July):188–96.
24. Duggal NA, Niemi G, Harridge SDR, Simpson RJ, Lord JM. Can physical activity ameliorate immunosenescence and thereby reduce age-related multi-morbidity? *Nature Reviews Immunology [Internet]*. 2019;19(9):563–72. Available from: <http://dx.doi.org/10.1038/s41577-019-0177-9>
25. Centers for Disease Control and P. Physical Activity. 2002.

- 1 26. Bartlett DB, Duggal NA. Moderate physical activity associated with a higher
2 naïve/memory T-cell ratio in healthy old individuals: potential role of IL15.
3 Age Ageing. 2020 Apr;49(3):368–73.
- 4 27. Bazgir B, Salesi M, Koushki M, Amirghofran Z. Effects of Eccentric and
5 Concentric Emphasized Resistance Exercise on IL-15 Serum Levels and
6 Its Relation to Inflammatory Markers in Athletes and Non-Athletes. Asian
7 Journal of Sports Medicine [Internet]. 2015 Sep;6(3). Available from:
8 [https://search.proquest.com/scholarly-journals/effects-eccentric-](https://search.proquest.com/scholarly-journals/effects-eccentric-concentric-emphasized/docview/1777491290/se-2?accountid=8113)
9 [concentric-emphasized/docview/1777491290/se-2?accountid=8113](https://search.proquest.com/scholarly-journals/effects-eccentric-concentric-emphasized/docview/1777491290/se-2?accountid=8113)
- 10 28. Brunelli DT, Chacon-Mikahil MPT, Gáspari AF, Lopes WA, Bonganha V,
11 Bonfante ILP, et al. Combined Training Reduces Subclinical Inflammation
12 in Obese Middle-Age Men. Medicine & Science in Sports & Exercise
13 [Internet]. 2015 Oct;47(10):2207–15. Available from:
14 <https://insights.ovid.com/crossref?an=00005768-201510000-00024>
- 15 29. Prestes J, Shiguemoto G, Botero JP, Frollini A, Dias R, Leite R, et al.
16 Effects of resistance training on resistin, leptin, cytokines, and muscle force
17 in elderly post-menopausal women. J Sports Sci. 2009 Dec;27(14):1607–
18 15.
- 19 30. Beavers KM, Hsu FC, Isom S, Kritchevsky SB, Church T, Goodpaster B, et
20 al. Long-term physical activity and inflammatory biomarkers in older adults.
21 Med Sci Sports Exerc. 2010 Dec;42(12):2189–96.
- 22 31. Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan-a web and
23 mobile app for systematic reviews. Systematic Reviews [Internet].
24 2016;5(1):1–10. Available from: [http://dx.doi.org/10.1186/s13643-016-](http://dx.doi.org/10.1186/s13643-016-0384-4)
25 [0384-4](http://dx.doi.org/10.1186/s13643-016-0384-4)
- 26 32. Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from
27 the median, range, and the size of a sample. BMC Medical Research
28 Methodology. 2005;5:1–10.
- 29 33. Maher CG, Sherrington C, Herbert RD, Moseley AM, Elkins M. Reliability
30 of the PEDro scale for rating quality of randomized controlled trials.
31 Physical Therapy. 2003;83(8):713–21.
- 32 34. Kuczynski S, Winiarska H, Abramczyk M, Szczawinska K, Wierusz-
33 Wysocka B, Dworacka M, et al. IL-15 is elevated in serum patients with
34 type 1 diabetes mellitus. DIABETES RESEARCH AND CLINICAL
35 PRACTICE. 2005 Sep;69(3):231–6.
- 36 35. B van der ZIJ, S van der VE, Wester VL, Nagtzaam NMA, C van REF,
37 Leenen PJM, et al. Obesity-associated T-cell and macrophage activation
38 improve partly after a lifestyle intervention. International Journal of Obesity
39 [Internet]. 2020 Sep;44(9):1838–50. Available from:
40 [https://search.proquest.com/scholarly-journals/obesity-associated-t-cell-](https://search.proquest.com/scholarly-journals/obesity-associated-t-cell-macrophage-activation/docview/2436697716/se-2?accountid=8113)
41 [macrophage-activation/docview/2436697716/se-2?accountid=8113](https://search.proquest.com/scholarly-journals/obesity-associated-t-cell-macrophage-activation/docview/2436697716/se-2?accountid=8113)
- 42 36. Capó X, Martorell M, Llompарт I, Sureda A, Tur JA, Pons A.
43 Docosahexanoic acid diet supplementation attenuates the peripheral
44 mononuclear cell inflammatory response to exercise following LPS
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- activation. Cytokine [Internet]. 2014;69(2 CC-Complementary Medicine):155-164. Available from: <https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01041594/full>
37. Han D, Zhang Y, Chen J, Hua G, Li J, Deng X, et al. Transcriptome analyses of differential gene expression in the bursa of Fabricius between Silky Fowl and White Leghorn. *Sci Rep*. 2017 Apr;7:45959.
 38. Levinger P, Caldow MK, Bartlett JR, Peake JM, Smith C, Cameron-Smith D, et al. The level of FoxO1 and IL-15 in skeletal muscle, serum and synovial fluid in people with knee osteoarthritis: a case control study. *Osteoporosis international: a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA*. 2016 Jun;27(6):2137–43.
 39. Woo SH, Bhattacharya S, Derby G, Taylor I, Myers BD, Lafayette RA. A serum cytokine network in immunoglobulin a nephropathy. *Nephrology Reviews* [Internet]. 2012;4(1):36–41. Available from: <https://www.embase.com/search/results?subaction=viewrecord&id=L622647441&from=export>
 40. Oliver JM, Jenke SC, Mata JD, Kreutzer A, Jones MT. Acute Effect of Cluster and Traditional Set Configurations on Myokines Associated with Hypertrophy. *Int J Sports Med* [Internet]. 2016;37(13):1019-1024. Available from: <https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01342497/full>
 41. Yalcin A, Silay K, Balik AR, Avcioğlu G, Aydin AS. The relationship between plasma interleukin-15 levels and sarcopenia in outpatient older people. *Aging Clin Exp Res*. 2018 Jul;30(7):783–90.
 42. Al-Shukaili A, AL-Ghafri S, Al-Marhoobi S, Al-Abri S, Al-Lawati J, Al-Maskari M. Analysis of Inflammatory Mediators in Type 2 Diabetes Patients. Migdalis I, editor. *International Journal of Endocrinology* [Internet]. 2013;2013. Available from: <https://search.proquest.com/scholarly-journals/analysis-inflammatory-mediators-type-2-diabetes/docview/2407662987/se-2?accountid=8113>
 43. Biancotto A, Wank A, Perl S, Cook W, Olnes MJ, Dagur PK, et al. Baseline levels and temporal stability of 27 multiplexed serum cytokine concentrations in healthy subjects. *PLoS One*. 2013;8(12):e76091.
 44. Ahn N, Kim K. Can Active Aerobic Exercise Reduce the Risk of Cardiovascular Disease in Prehypertensive Elderly Women by Improving HDL Cholesterol and Inflammatory Markers? *Int J Environ Res Public Health*. 2020 Aug;17(16).
 45. Bowman GL, Dayon L, Kirkland R, Wojcik J, Peyratout G, Severin IC, et al. Blood-brain barrier breakdown, neuroinflammation, and cognitive decline in older adults. *Alzheimers Dement*. 2018 Dec;14(12):1640–50.
 46. Bugera EM, Duhamel TA, Peeler JD, Cornish SM. The systemic myokine response of decorin, interleukin-6 (IL-6) and interleukin-15 (IL-15) to an

- acute bout of blood flow restricted exercise. *Eur J Appl Physiol* [Internet]. 2018;118(12):2679-2686. Available from: <https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01707943/full>
- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
47. Cassano P, Bui E, Rogers AH, Walton ZE, Ross R, Zeng M, et al. Inflammatory cytokines in major depressive disorder: A case-control study. *Aust N Z J Psychiatry*. 2017 Jan;51(1):23–31.
48. Choe J yoon, Lee H, Kim SG, Kim MJ, Park S hoon, Kim S kyu. The distinct expressions of interleukin-15 and interleukin-15 receptor [alpha] in Behçet's disease. *Rheumatology International* [Internet]. 2013 Aug;33(8):2109–15. Available from: <https://search.proquest.com/docview/1411044696?accountid=8113>
49. Christiansen T, Bruun JM, Paulsen SK, Ølholm J, Overgaard K, Pedersen SB, et al. Acute exercise increases circulating inflammatory markers in overweight and obese compared with lean subjects. *European Journal of Applied Physiology* [Internet]. 2013 Jun;113(6):1635–42. Available from: <https://search.proquest.com/docview/1412462909?accountid=8113>
50. Csencsits-Smith K, Suescun J, Li K, Luo S, Bick DL, Schiess M. Serum Lymphocyte-Associated Cytokine Concentrations Change More Rapidly over Time in Multiple System Atrophy Compared to Parkinson Disease. *Neuroimmunomodulation*. 2016;23(5–6):301–8.
51. di Renzo L, Galvano F, Orlandi C, Bianchi A, di Giacomo C, la Fauci L, et al. Oxidative stress in normal-weight obese syndrome. *Obesity (Silver Spring)*. 2010 Nov;18(11):2125–30.
52. Gokkusu C, Aydin M, Ozkok E, Tulubas F, Elitok A, Pamukcu B, et al. Influences of genetic variants in interleukin-15 gene and serum interleukin-15 levels on coronary heart disease. *Cytokine*. 2010 Jan;49(1):58–63.
53. Gonza1ez-Reimers E, Fernandez-Rodriguez CM, Santolaria-Fernandez F, de la Vega-Prieto MJMJ, Martin-Gonzalez C, Gornez-Rodriguez MA, et al. Interleukin-15 and Other Myokines in Chronic Alcoholics. *Alcohol and Alcoholism* [Internet]. 2011 Sep;46(5):529–33. Available from: <https://search.proquest.com/scholarly-journals/interleukin-15-other-myokines-chronic-alcoholics/docview/1023093813/se-2?accountid=8113>
54. Gullick NJ, Abozaid HS, Jayaraj DM, Evans HG, Scott DL, Choy EH, et al. Enhanced and persistent levels of interleukin (IL)-17⁺ CD4⁺ T cells and serum IL-17 in patients with early inflammatory arthritis. *Clin Exp Immunol*. 2013 Nov;174(2):292–301.
55. Urzi F, Marusic U, Ličen S, Buzan E. Effects of Elastic Resistance Training on Functional Performance and Myokines in Older Women-A Randomized Controlled Trial. *J Am Med Dir Assoc*. 2019 Jul;20(7):830-834.e2.
56. Hu S, Chen WC, Hwang GS, Chen ST, Kuo SB, Chen Y, et al. Changes in plasma steroids and cytokines levels in betel chewing patients in Taiwan. *STEROIDS*. 2016 Jul;111(SI):134–8.

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
57. Jekarl DW, Kim JY, Ha JH, Lee S, Yoo J, Kim M, et al. Diagnosis and Prognosis of Sepsis Based on Use of Cytokines, Chemokines, and Growth Factors. *Dis Markers*. 2019;2019:1089107.
 58. Johansson P, Almquist EG, Wallin A, Johansson JO, Andreasson U, Blennow K, et al. Reduced cerebrospinal fluid concentration of interleukin-12/23 subunit p40 in patients with cognitive impairment. *PLoS One*. 2017;12(5):e0176760.
 59. Joy JM, Vogel RM, Moon JR, Falcone PH, Mosman MM, Pietrzkowski Z, et al. Ancient peat and apple extracts supplementation may improve strength and power adaptations in resistance trained men. *BMC Complement Altern Med [Internet]*. 2016;16 CC-:224. Available from: <https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01263562/full>
 60. Kakumu S, Okumura A, Ishikawa T, Yano M, Enomoto A, Nishimura H, et al. Serum levels of IL-10, IL-15 and soluble tumour necrosis factor-alpha (TNF-alpha) receptors in type C chronic liver disease. *Clin Exp Immunol*. 1997 Sep;109(3):458–63.
 61. Knuiman P, Hopman MTE, Hangelbroek R, Mensink M. Plasma cytokine responses to resistance exercise with different nutrient availability on a concurrent exercise day in trained healthy males. *Physiological Reports [Internet]*. 2018 Jun;6(11). Available from: <https://search.proquest.com/scholarly-journals/plasma-cytokine-responses-resistance-exercise/docview/2054462754/se-2?accountid=8113>
 62. Lambert CP, Flynn MG, Sullivan DH, Evans WJ. Effects of megestrol acetate on circulating interleukin-15 and interleukin-18 concentrations in healthy elderly men. *J Gerontol A Biol Sci Med Sci*. 2004 Aug;59(8):855–8.
 63. Lesiak A, Bednarski I, Pałczyńska M, Kumiszczka E, Kraska-Gacka M, Woźniacka A, et al. Are interleukin-15 and -22 a new pathogenic factor in pustular palmoplantar psoriasis? *Postepy Dermatologii i Alergologii [Internet]*. 2016;33(5):336–9. Available from: <https://search.proquest.com/scholarly-journals/are-interleukin-15-22-new-pathogenic-factor/docview/1838510614/se-2?accountid=8113>
 64. Lis D, Stellingwerff T, Kitic CM, Ahuja KDK, Fell J. No Effects of a Short-Term Gluten-free Diet on Performance in Nonceliac Athletes. *Med Sci Sports Exerc*. 2015 Dec;47(12):2563–70.
 65. Luna LA, Bachi AL, Novaes e BRR, Eid RG, Suguri VM, Oliveira PW, et al. Immune responses induced by *Pelargonium sidoides* extract in serum and nasal mucosa of athletes after exhaustive exercise: modulation of secretory IgA, IL-6 and IL-15. *Phytomedicine [Internet]*. 2011;18(4 CC-Complementary Medicine):303-308. Available from: <https://www.cochranelibrary.com/central/doi/10.1002/central/CN-00972890/full>

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66. Martínez-Hernández PL, Hernanz-Macías Á, Gómez-Candela C, Grande-Aragón C, Feliu-Batlle J, Castro-Carpeño J, et al. Serum interleukin-15 levels in cancer patients with cachexia. *Oncol Rep*. 2012 Oct;28(4):1443–52.
67. Micielska K, Gmiat A, Zychowska M, Kozłowska M, Walentukiewicz A, Lysak-Radomska A, et al. The beneficial effects of 15 units of high-intensity circuit training in women is modified by age, baseline insulin resistance and physical capacity. *Diabetes Res Clin Pract*. 2019 Jun;152:156–65.
68. Rentzos M, Paraskevas GP, Kapaki E, Nikolaou C, Zoga M, Tsoutsou A, et al. Circulating interleukin-15 in dementia disorders. *J Neuropsychiatry Clin Neurosci*. 2007;19(3):318–25.
69. Rentzos M, Nikolaou C, Andreadou E, Paraskevas GP, Rombos A, Zoga M, et al. Circulating interleukin-15 and RANTES chemokine in Parkinson's disease. *Acta Neurol Scand*. 2007 Dec;116(6):374–9.
70. Mustafa T, Brokstad KA, Mfinanga SG, Wiker HG. Multiplex Analysis of Pro- or Anti-Inflammatory Serum Cytokines and Chemokines in relation to Gender and Age among Tanzanian Tuberculous Lymphadenitis Patients. *Tuberc Res Treat*. 2015;2015:561490.
71. Nathella PK, Banurekha V v, Nair D, Babu S. Diminished plasma levels of common γ -chain cytokines in pulmonary tuberculosis and reversal following treatment. *PLoS One* [Internet]. 2017 Apr;12(4). Available from: <https://search.proquest.com/scholarly-journals/diminished-plasma-levels-common-gamma-chain-cytokines/docview/1989977365/se-2?accountid=8113>
72. Nishida Y, Higaki Y, Taguchi N, Hara M, Nakamura K, Nanri H, et al. Objectively measured physical activity and inflammatory cytokine levels in middle-aged Japanese people. *Prev Med (Baltim)*. 2014 Jul;64:81–7.
73. Nishida Y, Tanaka K, Hara M, Hirao N, Tanaka H, Tobina T, et al. Effects of home-based bench step exercise on inflammatory cytokines and lipid profiles in elderly Japanese females: A randomized controlled trial. *Arch Gerontol Geriatr*. 2015;61(3):443–51.
74. Notarnicola A, Lapadula G, Natuzzi D, Lundberg IE, Iannone F. Correlation between serum levels of IL-15 and IL-17 in patients with idiopathic inflammatory myopathies. *Scand J Rheumatol*. 2015 May;44(3):224–8.
75. Olszewski WL, Pazdur J, Kubasiewicz E, Zaleska M, Cooke CJ, Miller NE. Lymph draining from foot joints in rheumatoid arthritis provides insight into local cytokine and chemokine production and transport to lymph nodes. *Arthritis Rheum*. 2001 Mar;44(3):541–9.
76. Phenekos C, Vryonidou A, Gritzapis AD, Baxevanis CN, Goula M, Papamichail M. Th1 and Th2 serum cytokine profiles characterize patients with Hashimoto's thyroiditis (Th1) and Graves' disease (Th2). *Neuroimmunomodulation*. 2004;11(4):209–13.
77. Ragino YI, Oblaukhova VI, Polonskaya Y v, Kuzminykh NA, Shcherbakova L v, Kashtanova E v. The blood cytokine profile of young people with early ischemic heart disease comorbid with abdominal obesity. *Journal of*

- Personalized Medicine [Internet]. 2020;10(3):1–12. Available from: <https://www.embase.com/search/results?subaction=viewrecord&id=L2004911734&from=export>
- 1
2
3
4 78. Rinnov A, Yfanti C, Nielsen S, Åkerström TCA, Peijs L, Zankari A, et al. Endurance training enhances skeletal muscle interleukin-15 in human male subjects. *Endocrine*. 2014;45(2):271–8.
 - 5
6
7
8 79. Sánchez-Jiménez R, Cerón E, Bernal-Alcántara D, Castillejos-López M, Gonzalez-Trujano E, Negrete-García MC, et al. Association between IL-15 and insulin plasmatic concentrations in patients with pulmonary tuberculosis and type 2 diabetes. *Tuberculosis (Edinb)*. 2018 Jul;111:114–20.
 - 9
10
11
12
13
14
15 80. Shammam KH, Al-Tu'ma FJ, El-Yassin HD. Myokines in alcoholic myopathy. *JOURNAL OF CONTEMPORARY MEDICAL SCIENCES*. 2015;1(3):20–3.
 - 16
17
18
19 81. Shibatomi K, Ida H, Yamasaki S, Nakashima T, Origuchi T, Kawakami A, et al. A novel role for interleukin-18 in human natural killer cell death: high serum levels and low natural killer cell numbers in patients with systemic autoimmune diseases. *Arthritis Rheum*. 2001 Apr;44(4):884–92.
 - 20
21
22
23
24
25 82. Tagoma A, Haller-Kikkatalo K, Roos K, Oras A, Kirss A, Ilonen J, et al. Interleukin-7, T helper 1, and regulatory T-cell activity-related cytokines are increased during the second trimester of healthy pregnancy compared to non-pregnant women. *Am J Reprod Immunol*. 2019 Dec;82(6):e13188–e13188.
 - 26
27
28
29
30
31 83. Uzawa A, Kawaguchi N, Himuro K, Kanai T, Kuwabara S. Serum cytokine and chemokine profiles in patients with myasthenia gravis. *Clin Exp Immunol*. 2014 May;176(2):232–7.
 - 32
33
34
35 84. Xia L, Shen H, Xiao W, Lu J. Increased serum TWEAK levels in Psoriatic arthritis: relationship with disease activity and matrix metalloproteinase-3 serum levels. *Cytokine*. 2011 Mar;53(3):289–91.
 - 36
37
38
39
40 85. Yang J, Azat M, Peng P, Chang Y, Gao R, Li W, et al. Serum levels of TNF-alpha, IL-1 beta, IL-9, and IL-15 in acute respiratory distress syndrome. *INTERNATIONAL JOURNAL OF CLINICAL AND EXPERIMENTAL PATHOLOGY*. 2017;10(1):781–8.
 - 41
42
43
44
45 86. Zhu H, Mi W, Luo H, Chen T, Liu S, Raman I, et al. Whole-genome transcription and DNA methylation analysis of peripheral blood mononuclear cells identified aberrant gene regulation pathways in systemic lupus erythematosus. *Arthritis Research & Therapy [Internet]*. 2016;18. Available from: <https://search.proquest.com/scholarly-journals/whole-genome-transcription-dna-methylation/docview/1807961899/se-2?accountid=8113>
 - 46
47
48
49
50
51
52
53
54
55 87. Hingorjo MR, Zehra S, Saleem S, Qureshi MA. Serum Interleukin-15 and its relationship with adiposity Indices before and after short-term endurance exercise. *PAKISTAN JOURNAL OF MEDICAL SCIENCES*. 2018;34(5):1125–31.
 - 56
57
58
59
60
61
62
63
64
65

- 1 88. Notarnicola A, Lapadula G, Natuzzi D, Iannone F. Possible interplay
2 between interleukin-15 and interleukin-17 into the pathogenesis of
3 idiopathic inflammatory myopathies. *Reumatismo*. 2014 Nov;66(3):215–
4 23.
- 5 89. Quinn KM, Hussain T, Kraus F, Formosa LE, Lam WK, Dagley MJ, et al.
6 Metabolic characteristics of CD8(+) T cell subsets in young and aged
7 individuals are not predictive of functionality. *Nat Commun*. 2020
8 Jun;11(1):2857.
- 9 90. Rentzos M, Zoga M, Paraskevas GP, Kapaki E, Rombos A, Nikolaou C, et
10 al. IL-15 is elevated in cerebrospinal fluid of patients with Alzheimer's
11 disease and frontotemporal dementia. *J Geriatr Psychiatry Neurol*. 2006
12 Jun;19(2):114–7.
- 13 91. Banitalebi E, Kazemi A, Faramarzi M, Nasiri S, Haghighi MM. Effects of
14 sprint interval or combined aerobic and resistance training on myokines in
15 overweight women with type 2 diabetes: a randomized controlled trial. *Life*
16 *Sci* [Internet]. 2019;217:101-109. Available from:
17 [https://www.cochranelibrary.com/central/doi/10.1002/central/CN-](https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01702848/full)
18 [01702848/full](https://www.cochranelibrary.com/central/doi/10.1002/central/CN-01702848/full)
- 19 92. Gomez AM, Martinez C, Fiuza-Luces C, Herrero F, Perez M, Madero L, et
20 al. Exercise training and cytokines in breast cancer survivors. *Int J Sports*
21 *Med*. 2011 Jun;32(6):461–7.
- 22 93. Riechman SE, Balasekaran G, Roth SM, Ferrell RE. Association of
23 interleukin-15 protein and interleukin-15 receptor genetic variation with
24 resistance exercise training responses. *Journal of Applied Physiology*.
25 2004;97(6):2214–9.
- 26 94. Tsai CL, Pai MC, Ukropec J, Ukropcová B. Distinctive Effects of Aerobic
27 and Resistance Exercise Modes on Neurocognitive and Biochemical
28 Changes in Individuals with Mild Cognitive Impairment. *Curr Alzheimer*
29 *Res*. 2019;16(4):316–32.
- 30 95. Silva BS de A, Lira FSFS, Rossi FEFE, Ramos D, Uzeloto JS, Freire APCF,
31 et al. Inflammatory and Metabolic Responses to Different Resistance
32 Training on Chronic Obstructive Pulmonary Disease: A Randomized
33 Control Trial. *FRONTIERS IN PHYSIOLOGY*. 2018 Mar;9:262.
- 34 96. Tsai CL, Pai MC, Ukropec J, Ukropcova B. Distinctive Effects of Aerobic
35 and Resistance Exercise Modes on Neuro-cognitive and Biochemical
36 Changes in Individuals with Mild Cognitive Impairment. *Current Alzheimer*
37 *Research*. 2019;16(4):316–32.
- 38 97. Gómez AM, Martínez C, Fiuza-Luces C, Herrero F, Pérez M, Madero L, et
39 al. Exercise training and cytokines in breast cancer survivors. *Int J Sports*
40 *Med* [Internet]. 2011;32(6):461-467. Available from:
41 [https://www.cochranelibrary.com/central/doi/10.1002/central/CN-](https://www.cochranelibrary.com/central/doi/10.1002/central/CN-00801376/full)
42 [00801376/full](https://www.cochranelibrary.com/central/doi/10.1002/central/CN-00801376/full)
- 43 98. Marzetti E, Carter CS, Wohlgemuth SE, Lees HA, Giovannini S, Anderson
44 B, et al. Changes in IL-15 expression and death-receptor apoptotic
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

- 1 signaling in rat gastrocnemius muscle with aging and life-long calorie
2 restriction. *Mechanisms of Ageing and Development*. 2009;130(4):272–80.
- 3 99. Cevenini E, Monti D, Franceschi C. Inflamm-aging. *Current Opinion in*
4 *Clinical Nutrition and Metabolic Care*. 2013;16(1):14–20.
- 5 100. Wallace DL, Bérard M, Soares M v.d., Oldham J, Cook JE, Akbar AN, et
6 al. Prolonged exposure of naïve CD8+ T cells to interleukin-7 or interleukin-
7 15 stimulates proliferation without differentiation or loss of telomere length.
8 *Immunology*. 2006;119(2):243–53.
- 9 101. Sun H, Liu D. IL-15/sIL-15R[alpha] gene transfer suppresses Lewis lung
10 cancer growth in the lungs, liver and kidneys. *Cancer Gene Therapy*
11 [Internet]. 2016 Feb;23(2–3):54–60. Available from:
12 [https://search.proquest.com/scholarly-journals/il-15-sil-15r-alpha-gene-](https://search.proquest.com/scholarly-journals/il-15-sil-15r-alpha-gene-transfer-suppresses/docview/1778707375/se-2?accountid=8113)
13 [transfer-suppresses/docview/1778707375/se-2?accountid=8113](https://search.proquest.com/scholarly-journals/il-15-sil-15r-alpha-gene-transfer-suppresses/docview/1778707375/se-2?accountid=8113)
14
- 15 102. Nadeau L, Patten DA, Caron A, Garneau L, Pinault-Masson E, Foretz M,
16 et al. IL-15 improves skeletal muscle oxidative metabolism and glucose
17 uptake in association with increased respiratory chain supercomplex
18 formation and AMPK pathway activation. *Biochimica et Biophysica Acta -*
19 *General Subjects* [Internet]. 2019;1863(2):395–407. Available from:
20 <https://doi.org/10.1016/j.bbagen.2018.10.021>
21
- 22 103. Zhao S, Kusminski CM, Elmquist JK, Scherer PE. Leptin: Less is more.
23 *Diabetes*. 2020;69(5):823–9.
- 24 104. Galic S, Oakhill JS, Steinberg GR. Adipose tissue as an endocrine organ.
25 *Molecular and Cellular Endocrinology*. 2010;316(2):129–39.
- 26 105. Chen W, Balland E, Cowley MA. Hypothalamic Insulin Resistance in
27 Obesity: Effects on Glucose Homeostasis. *Neuroendocrinology*.
28 2017;104(4):364–81.
- 29 106. Alvarez B, Carbó N, López-Soriano J, Drivdahl RH, Busquets S, López-
30 Soriano FJ, et al. Effects of interleukin-15 (IL-15) on adipose tissue mass
31 in rodent obesity models: Evidence for direct IL-15 action on adipose tissue.
32 *Biochimica et Biophysica Acta - General Subjects*. 2002;1570(1):33–7.
- 33 107. Goossens GH. The Metabolic Phenotype in Obesity: Fat Mass, Body Fat
34 Distribution, and Adipose Tissue Function. *Obesity Facts*. 2017 Jul
35 1;10(3):207–15.
- 36 108. Kuk JL, Saunders TJ, Davidson LE, Ross R. Age-related changes in total
37 and regional fat distribution. Vol. 8, *Ageing Research Reviews*. 2009. p.
38 339–48.
- 39 109. Aadahl M, Kjaer M, Kristensen JH, Mollerup B, Jørgensen T. Self-reported
40 physical activity compared with maximal oxygen uptake in adults [Internet].
41 Vol. 14, *European Journal of Cardiovascular Prevention and Rehabilitation*.
42 2007. Available from:
43 <https://academic.oup.com/eurjpc/article/14/3/422/5932945>
44
- 45 110. Tamura Y, Watanabe K, Kantani T, Hayashi J, Ishida N, Kaneki M.
46 Upregulation of circulating IL-15 by treadmill running in healthy individuals:
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

is IL-15 an endocrine mediator of the beneficial effects of endurance exercise? *Endocr J.* 2011;58(3):211–5.

111. Ostrowski K, Hermann C, Bangash A, Schjerling P, Nielsen JN, Pedersen BK. A trauma-like elevation of plasma cytokines in humans in response to treadmill running. *Journal of Physiology.* 1998;513(3):889–94.
112. Nadeau L, Aguer C. Interleukin-15 as a myokine: Mechanistic insight into its effect on skeletal muscle metabolism. *Applied Physiology, Nutrition and Metabolism.* 2019;44(3):229–38.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

The effects of ageing, BMI and physical activity on blood IL-15 levels: A Systematic Review and Meta-analyses.

Guilherme Henrique Jesus do Prado¹, Amanda Veiga Sardeli*^{1,2,3,4}, Janet Mary Lord^{3,4}, Cláudia Regina Cavaglieri^{1,2}

1 Laboratory of Exercise Physiology, School of Physical Education, University of Campinas, Campinas-SP, Brazil.

2 Gerontology program, School of Medical Sciences, University of Campinas, Campinas-SP, Brazil.

3 MRC-Versus Arthritis Centre for Musculoskeletal Ageing Research, Institute of Inflammation and Ageing, University of Birmingham, Birmingham, UK.

4 NIHR Birmingham Biomedical Research Centre, University Hospital Birmingham and University of Birmingham, Birmingham, UK.

* Corresponding Author: Amanda Veiga Sardeli, Institute of Inflammation and Ageing, Queen Elizabeth Hospital, Mindelsohn Way, Birmingham, West Midlands, B15 2WB, UK, e-mail: amandavsardeli@gmail.com.