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Rees, Nia Paddison; Shaheen, Walaa; Quince, Christopher; Tselepis, Chris; Horniblow, Richard D; Sharma, Naveen; Beggs, Andrew D; Iqbal, Tariq H; Quraishi, Mohammed Nabil

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Systematic review of donor and recipient predictive biomarkers of response to faecal microbiota transplantation in patients with ulcerative colitis



Nia Paddison Rees, a,b Walaa Shaheen, a,b Christopher Quince, a,b Chris Tselepis, a,d Richard D Horniblow, a,d Naveen Sharma, a,b,e Andrew D Beggs, a,b,e Tariq H Iqbal, a,e,f,* and Mohammed Nabil Quraishi a,b,e,***

Summary

Background Faecal microbiota transplantation (FMT) has previously been explored as a treatment for ulcerative colitis (UC) however, biomarkers that predict and / or are associated with clinical response are poorly defined. The aim of this systematic review was to identify donor and recipient clinical, microbial and metabolomic predictive biomarkers of response to FMT in UC.

Methods A systematic search of the relevant literature of studies exploring FMT in UC was conducted. Data on microbial diversity, taxonomic changes, metabolic changes, donor and recipient microbiota relationship and baseline predictors was examined.

Findings 2852 studies were screened, and 25 papers were included in this systematic review. Following FMT, alpha diversity was seen to increase in responders along with increases in the abundance of *Clostridiales* clusters (order) and *Bacteroides* genus. Metabolomic analysis revealed short chain fatty acid (SCFA) production as a marker of FMT success. Donors or FMT batches with higher microbial alpha diversity and a greater abundance of taxa belonging to certain Bacteroides and *Clostridia* clusters were associated with clinical response to FMT. Baseline clinical predictors of response in patients with UC included younger age, less severe disease and possibly shorter disease duration. Baseline recipient microbial predictors at response consisted of higher faecal species richness, greater abundance of *Candida* and donor microbial profile similarity.

Interpretation Distinct changes in gut microbiota profiles post-FMT indicate that certain baseline characteristics along with specific microbial and metabolomic alterations may predispose patients towards a successful therapeutic outcome. Opportunities towards a biomarker led precision medicine approach with FMT should be explored in future clinical studies.

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Key words: Ulcerative colitis; Faecal microbiota transplantation; Biomarkers

E-mail address: nabil.quraishi@nhs.net (M.N. Quraishi). PROSPERO registration ID: 331957

Introduction

Ulcerative colitis (UC) is a subtype of inflammatory bowel disease (IBD) that is characterised by chronic inflammation of the colonic mucosa with patients typically presenting with bloody diarrhoea. Whilst the precise aetiology of UC remains unclear, it is considered to be triggered by dysregulated and sustained immune responses to gut microbiota in genetically susceptible

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^aUniversity of Birmingham Microbiome Treatment Centre, Birmingham, UK

^bInstitute of Cancer and Genomic Sciences, University of Birmingham, UK

^cEarlham Institute, Norwich, UK

^dInstitute of Clinical Sciences, School of Biomedical Sciences, University of Birmingham, UK

^eUniversity Hospitals Birmingham NHS Foundation Trust, Birmingham, UK

finstitute of Microbiology and Infection, University of Birmingham, UK

^{**}Address for correspondence: Dr MN Quraishi; Department of Gastroenterology, Birmingham Heartlands Hospital, University Hospitals Birmingham NHS Foundation Trust, Birmingham, UK. Phone number: 01214243295

^{*} Joint senior authors

Research in context

Evidence before this study

To date, eight double-blind randomised placebo-controlled trials on the use of FMT to treat UC have been published, 6 of which have reported positive findings. Whilst these studies highlight the capability of FMT to ameliorate UC, very little is known about the underpinning mechanisms. The lack of well-defined biomarkers and treatment targets makes it pragmatically challenging to determine the frequency and interval at which treatment with FMT should be administered.

Added value of this study

Through a systematic review of the current evidence base, we describe clinical, microbial and metabolomic biomarkers that are predictive of response at baseline (pre-FMT), and are associated with response following FMT treatment in patients with active UC.

Implications of all the available evidence

The findings of this systematic review highlight the possibility of enhancing a sustained response to FMT through biomarker-based selection and optimisation of donors and patients before and during the treatment with FMT. Utilising precision medicine in this field deserves further exploration as it has the potential to facilitate an individualised, biomarker driven 'treat to microbiome/metabolome' target approach with FMT in patients with UC.

individuals.^{2,3} Patients with UC possess an altered gut microbiota composition, known as dysbiosis, characterised by reduced microbiota diversity, decrease in the phylum Bacteroidetes and Firmicutes along with a corresponding increase in Proteobacteria.^{4,5} This has led to a focus on the modulation of gut bacteria as a treatment method for UC, primarily through faecal microbiota transplantation (FMT). FMT is the procedure of transferring processed faecal matter of a healthy individual into another individual with a microbiota mediated disease.⁶

To date, eight double-blind randomised placebo-controlled trials (RCTs) on the use of FMT to treat UC have been published, 6 of which have reported positive findings.⁷⁻¹⁴ Whilst these studies highlight the capability of FMT to ameliorate UC, very little is known about the underpinning mechanisms. The heterogeneity in study designs both with regards to FMT preparation and administration protocols as well as patient selection makes it challenging to draw solid conclusions for its adoption into clinical practice. Furthermore, it remains unclear if specific donor or recipient characteristics may predict response to FMT or denote successful response following FMT.^{15,16} This lack of well-defined biomarkers

and treatment targets makes it pragmatically challenging to determine the frequency and interval at which treatment with FMT should be administered.

Currently, there are no published systematic reviews that explore predictive biomarkers of FMT in patients with UC. This systematic review aims to answer whether clinical, microbial and metabolomic predictive biomarkers exist and if so, which of these are predictive of response at baseline (pre-FMT), and are associated with response following FMT treatment in patients with active UC.

Methods

Search strategy and study selection

The systematic review was conducted in accordance with preferred reporting items for systematic reviews and meta-analyses (PRISMA) criteria. The databases MEDLINE, EMBASE, CINAHL and Cochrane Library, were searched for suitable articles from commencement to January 2022 using search terms outlined (Supplement Table 1). In addition, references included in earlier review articles were searched to identify any additional studies. Results from the searches were imported into a bibliography manager (EndNote X9) and duplicate studies were removed.

Randomised control trials (RCTs) and non-randomised studies were included with exclusion of case reports and conference abstracts. Double blind RCTs were further split based on comparators (placebo and non-placebo controlled studies). Studies consisting of patients of all ages with active UC examining any of the following: clinical, microbial (diversity and taxonomic changes) and metabolomic biomarkers at baseline and post FMT treatment predictive of induction and maintenance of clinical remission in patients with active UC were included. Studies were excluded if they had under 10 patients in the FMT treatment arm or only included patients with concurrent infections. No restriction on language or the comparator type for comparative study designs was implemented. Abstracts of the papers identified by the initial search were evaluated by the lead and senior authors for appropriateness to the study question. All relevant papers were obtained and analysed in detail. Articles were independently assessed by two reviewers using pre-defined eligibility criteria and any disagreements were resolved by consensus.

Data extraction

Data was extracted independently by the two reviewers onto a Microsoft Excel spreadsheet (Microsoft, Washington, USA) from the eligible studies. Data relating to donor and patient demographics, treatment groups/comparator(s) and outcome measures were collected. Exploratory data on changes in alpha and beta diversity,

| Reference | No of subjects | Control/ Comparator | Treatment | Median / Mean Age(years) | Gender (% Male) | Average disease severity indices at baseline | Treatment Duration | Relevant study characteristics |
|---|-------------------|---|--|--|------------------------------------|---|---|---|
| Paramsothy et al * (2019) ³³ | 81 | Placebo group (n = 40) | Treatment protocol Initial colonoscopic infusion followed by intensive FMT infusion enemas (n = 41) FMT preparation Pooled from multiple donors | FMT arm - 35.6 (27.8- 48.9) Placebo arm - 35.4 (27.7-45.6) | FMT arm — 54% Placebo arm — 63% | FMT arm — 8 (average Total Mayo score) Placebo arm — 8 (average Total Mayo score) | FMT treatment 5 day- s/week for 8 weeks | Patients in the placebo group were eligible to receive open-label FMT after the double-blind study period 314 faecal samples collected from the patients at screen ing, every 4 weeks during treatment, and 8 weeks after the blinded or open- label FMT therapy |
| Moayyedi et al (2015) ⁸ | 75 | Placebo group (n = 37) | Treatment protocol Exam- ined by flexible sigmoidos- copy followed by FMT infusion via enema (n = 38) | FMT arm — 42.2 (±15.0) Placebo arm — 35.8 (±12.1) | FMT arm — 47% Placebo arm — 70% | FMT arm $-$ 8.24 (±2.61) Total Mayo Clinic score Placebo arm $-$ 7.86 (±2.28) Total Mayo Clinic score | FMT treatment 1 day/- week for 6 weeks | Patients provided stool sam- ples when the study begar and during each week of FMT for microbiome analysis |
| Costello et al (2019) ⁹ | 73 | Autologous FMT control group (n = 35) | FMT preparation Single donor per patient Treatment protocol Anaero- bically prepared pooled donor FMT via colonoscopy followed by 2 enemas over 7 days (n = 38) FMT preparation Pooled from multiple donors | Donor FMT arm — 38.5 ²⁸⁻⁵² Autologous FMT arm — 35 ²⁵⁻⁴⁶ | dFMT— 53% aFMT — 57% | dFMT arm − 7.2 (±1.7) Mean Total Mayo score aFMT − 7.4 (±1.9) Mean Total Mayo score | FMT treatment per week with patients moni- tored at 8 weeks and 12 months post-FMT | Open-label therapy was offered to autologous FMT participants at 8 weeks and they were followed up for 12 months Recipient stool samples were collected at baseline (week 0) and weeks 4, 8, and 52 fmicrobiome, metabolome, and faecal calprotectin |
| Rossen <i>et al</i> (2015) †,10 | 48 | Autologous FMT control group (n = 25) | Treatment protocol Pre-treat- ment with bowel lavage fol- lowed by 2 duodenal infusions of a suspension of donor faeces via nasoduo- denal tube (n = 23) | Donor FMT arm — 40 ³³⁻⁵⁶ Autologous FMT arm — 41 ³⁰⁻⁴⁸ | dFMT arm — 47.8% aFMT arm — 44% | dFMT arm — 10 ⁵⁻¹¹ Median SCCAI score aFMT arm — 8 ⁴⁻¹¹ Median SCCAI score | FMT treatment at the start of the study (week 0) and 3 weeks later (week 3) | assessment Faecal samples were collectee at baseline before bowel lavage and 6 and 12 weeks after FMT |
| Crothers <i>et al</i> (2021) ¹³ | 12 | Placebo group (n = 6) | FMT preparation Single donor per patient Treatment protocol FMT induction by colonoscopy, followed by oral administra- tion of frozen encapsulated cFMT (n = 6) FMT preparation Single donor for induction Multiple (2 pre-defined) donors during maintanence | FMT arm — 41 (±15) Placebo arm — 52 ±15) | FMT arm - 67% Placebo arm - 50% | FMT arm $-6.3~(\pm 2.0)$ Mean Total Mayo score Placebo arm $-6.7~(\pm 1.2)$ Mean Total Mayo score | Daily cFMT treatment for 12 weeks | Subjects were followed for 36 weeks and longitudinal clir ical assessments Subjects in both arms of the study were pre-treated wit antibiotics for 7 days prior to FMT (or placebo) procedure Subject stool samples were obtained weekly throughout the study period, begin ning prior to antibiotic pretreatment, and ending at 18-weeks follow-up |

| Reference | No of subjects | Control/ Comparator | Treatment | Median / Mean Age(years) | Gender (% Male) | Average disease severity indices at baseline | Treatment Duration | Relevant study characteristics |
|---|-------------------|---------------------------|--|--|------------------------------------|--|--|--|
| Pai <i>et al</i> (2021) ¹² | 25 | Placebo group (n = 12) | Treatment protocol FMT administered by rectal enema (n=13) FMT preparation Multiple donors per patient (not pooled) | Overall 10.5 ⁴⁻¹⁷ Individual arms not specified | Not specified | Not specified | Total 12 enemas (given biweekly) | Seven patients randomized to the placebo arm crossed over to the open-label arm after 30 weeks of placebo treatment |
| Haifer <i>et al</i> (2021) ¹¹ | 35 | Placebo group (n=20) | Treatment protocol Six FMT capsules four times a day for 1 week, then six cap- sules twice daily for 1 week, followed by six capsules daily for the remaining 6 weeks. Each capsule con- tains 0.35g lyophilised stool. (n=15) | FMT arm - 37.1 (31.8–46.8) Placebo arm - 36.7 (25.1–42.0) | FMT arm — 60% Placebo arm — 45% | FMT arm - 5 ⁵⁻⁹ median total Mayo score Placebo arm - 7 ⁵⁻⁸ median total Mayo core | 8 weeks of capsules dur- ing induction, followed by 2 capsules daily for remaining 58 weeks for maintenance. | Antibiotic pre-treatment in both groups. 10 patients randomised to FMT arm with clinical response entered mainte- nance phase of the study - 4 assigned to FMT and 6 assigned to FMT withdrawal |
| | | | FMT preparation Two donors, unclear if pooled | | | | | |

Table 1: Randomised control studies of FMT in ulcerative colitis

- * Further post hoc microbiome and mycobiome analysis reported separately³¹FMT-faecal microbiota transplantation, cFMT-capsulised faecal microbiota transplantation, dFMT-donor FMT, aFMT-autologous FMT, SCCAI-simple clinical colitis activity index.

microbial taxa, metabolome and donor-patient microbiota similarities following FMT were collected. No unclear or missing data was noticed which would have required approaching the study authors for clarification. Risk of bias of the included RCTs was assessed with the Cochrane Collaboration's risk of bias tool and non-randomised/cohort studies was with the Newcastle-Ottawa quality assessment scale (NOS). 17,18 If there were any discrepancies a third reviewer was consulted.

Role of funders

No specific funding has been received for this systematic review. This is independent work conducted by the authors of the review.

Results

Study characteristics

The search strategy generated 2852 citations from which 25 articles investigating the use of FMT in UC patients satisfied the study selection criteria for further assessment (Figure 1). Of these, 7 were placebo controlled double blind RCTs⁷⁻¹³ (Table 1; total of 8 RCTs but one did not report predictive associations and failed to meet inclusion criteria for this systematic review), 2 were non-placebo controlled blinded randomised studies 19,20 and 14 were non-randomised or observational studies²¹⁻³⁰ (Table 2). In addition, 2 studies performed post-hoc microbiota analysis from their placebo controlled double blind RCTs.31,32 All the RCTs received a low bias ranking overall (Supplementary Table 2). None of the non-randomised / cohort studies scored at the highest end of the NOS scale, with a mean score of 5 (range 4 to 6) out of 9 (Supplementary Table 3).

Changes in microbial diversity

Five RCTs reported on changes in alpha diversity following FMT as presented in Table 3.7.9-II,13 Three observed a significant increase in alpha diversity relative to baseline following FMT in all patients regardless of clinical response.7.9,11 The FOCUS study observed this change being more pronounced in patients who entered clinical remission compared to those who did not. 7.33 In contrast the LOTUS study and the RCT by Costello and colleagues observed that the increase in alpha diversity following FMT was no longer significant when stratified by clinical response. 9,11 In comparison, the TURN trial observed a significant increase in alpha diversity in both donor FMT and autologous FMT responders but not in non-responders. 10,31 Amongst the non-randomised studies, only one study consisting demonstrated a significant increase in alpha diversity at post-FMT compared with pre-FMT, with this effect disappearing at 6 months.³⁴ Non-significant trends reported including

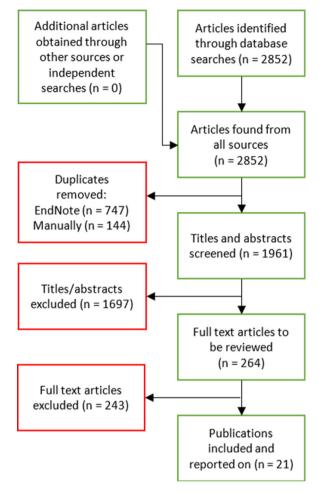


Figure 1. PRISMA flow diagram summarising the screening process for the systematic review

in increase in alpha diversity following FMT were observed in three non-randomised studies^{22,24,27} and one study showed a decrease in diversity with each sequential FMT treatment.²¹

Five RCTs reported on changes in beta diversity following FMT.^{7,8,10-12,31,33} Four observed a significant change in beta diversity following FMT in comparison to the placebo/inactive arm and relative to pre-FMT baseline. 7,8,10,11,31,33 Both the FOCUS trial and the RCT by Moayyedi and colleagues demonstrated a significant difference in the gut microbial composition following FMT. Furthermore, they demonstrated the gut microbial profiles following FMT were more similar to donors regardless of clinical response with Moayyedi demonstrating that this similarity was only seen between FMT treated recipient and their respective donor. Similarly, the TURN trial demonstrated that the microbiota composition of responders in the donor FMT group shifted from overlap with non-responders at baseline to healthy donors following FMT.31 These microbial compositional

| Reference | No. of Subjects | Control/Comparator | Treatment | Median / Mean Age (years) | Gender (% Male) | Average disease severity indices at baseline | Treatment duration | Relevant study characteristics |
|--|--------------------|---|--|--|---|---|---|--|
| Brezina et al (2021) ^{1,20} | 45 | 5-ASA treatment group (n = 22) | Treatment protocol Multi-session FMT enemas (n = 23) FMT preparation | FMT am — 39 ²⁵⁻⁶³ 5-ASA am — 39.5 ²⁷⁷⁰ | FMT arm — 52% 5-ASA arm — 50% | FMT arm – 6 ⁴⁻¹⁰ Total Mayo score 5-ASA arm – 6 ⁴⁻¹⁰ Total Mayo score | FMT treatment 5 times in the first week and then once weekly for 5 weeks | Faecal samples were collected at baseline and each study visit at weeks 2, 4, 6, and 12 in the FMI group and at the 1 year follows. |
| Shabat <i>et al</i> (2021) 1-19 | 22 | Single donor FMT by colonoscopy without dietary conditioning of donors (group 1 (n = 17)), UC Exclusion Diet (UCED) alone for patients (group 3 (n = 15)) | Treatment proto col Treatment proto col FMT by colonoscopy with dietary pre-conditioning of the donor for 14 days and a UCED for the patients (group 2 (n = 19)) FMT preparation Single donor per patient | Group 1 – 43.1 (±14.7) Group 2 – 43.5 (±10.5) Group 3 – 33.3 (±9.8) | Group 1 – 70.6% Group 2 – 73.7% Group 3 – 73.3% | Group $1-7^{6/9}$ Median SSCAI score Group $2-8^{6/10}$ Median SCCAI score Group $3-6^{5/6}$ Median SCCAI score | FMT by colonoscopy on day 1 and rectal enemas on days 2 and 14 | tollow-up in all patients. Three arm study exploring role of donor and recipient dietary conditioning in optimisation of response to FMT |
| Tian et al (2019) ²¹ | 20 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol Pre-treatment with bowel lavage followed by FMT via gastros- copy FMT preparation Single donor per patient | 62.50 (±77.14) | 9858 | 5.00 (±2.75) Mayo score | FMT treatment 5 times, once every 3 weeks | analysis performed on the bac- terial RNA from stool of healthy donors and patients with UC before treatment and after the first and second treatment (groups do, d1, and |
| Li <i>et al</i> (2020) ²² | 202 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol Single FMT via gastroscopy FMT preparation Single donor per patient | 36 (29-49.25) | 58.2% | 7 ⁵⁻⁹ Median Partial Mayo score | Patients received 1 infusion (2 nd and 3 rd courses were given to patients who relansed) | 42 UC stool samples were included: 22 samples at baseline and 20 at 5 days after the first course of FMT |
| Kump <i>et al</i> (2018) ²³ | 27 | Antibiotic pre-treatment only group (n = 10) | Treatment proto col Multi-session FMT via colonos- copy / copy / (n = 17) FMT preparation Single donor per patient | FMT am — 44 (±18) Antibiotic arm — 36 (±13) | FMT arm - 82% Antibiotic arm – 30% | FMT arm — 8.9 (±1.6) Mean Total Mayo score Antibiotic am — 8.1 (±3.1) Mean Total Mayo score | FMT treatment 5 times, at 14-day intervals (1 st treatment given via leocolonoscopy, the 4 subsequent sessions at days 14, 28, 42 and 56 were via flexible cirmoidoscopy | Faecal samples for microbiota analyses were collected at each study visit |
| Jacob <i>et al</i> (2017) ²⁴ | 20 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol Single FMT delivery via colonos- copy FMT preparation Pooled from multiple donors | 38.4 (±12.6) | %09 | 8.1 (±2.4) Mean Total Mayo score | Patients received 1 infusion | 165 rRNA gene sequencing was performed on recipient faecal DNA samples pre- and 2 and 4 weeks post- transplant |
| Fang <i>et al</i> (2021) ²⁵ | 20 | FMT routine therapy control group (n = 10) | Treatment protocol Monotherapy with a single fresh FMI via colonoscopy (in = 10) FMT preparation Single donor per patient | FMT am – 51.5 (±12.7) Control arm – 44.6 (±14.9) | Unclear | FMT arm — 9.5 (±2.5) Total Mayo score Control arm — 8.6 (±2.9) Total Mayo score | Patients received 1 infusion | Fresh faecal samples from the donors and pre-FMT and post-FMT samples from patients were collected |
| Table 2 (Continued) | (pai | | | | | | | |

| Reference | No. of Subjects | Control/Comparator | Treatment | Median / Mean Age (years) | Gender (% Male) | Average disease severity indices at baseline | Treatment duration | Relevant study characteristics |
|---|--------------------|--|---|---|--|--|---|--|
| Cui <i>et al</i> (2015) ²⁶ | 15 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol Step-up FMT treatment via endo- scopic infusion tube FMT preparation Unclear if multiple / pooled donos or single donor per patient | 31,711-48 | 73.3% | 86.7% Severe disease (53) 13.3% Moderate disease (52) | Initial FMT given followed by a 2 nd FMT after after 1 week, followed by 1 short course of steroid therapy and monitoring for 3 months after the and exact after the and exact after the after the and exact | Faecal samples from patients and donors pre-FMT, 1 week post-FMT and 4 months post-FMT were collected and stored for microbiota analysis by 165 rRNA sequencing |
| Chen <i>et al</i> (2020) ²⁷ | 47 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol Single-donor FMT via colonic transendoscopic enteral tubing ing FMT preparation Single donor programs | 44.4 (±15.5) | 57% | 5.9 (±2.0) Total Mayo score | Z"-FMI Total of 3 FMT treat- ments given every other day for 1 week | Molecular microbiological analyses were performed using faecal samples obtained from patients 1 day prior to FMT, and 4 and 12 weeks after FMT |
| Sood et al (2020) ²⁸ | 140 | No active comparator | Finger double per patent. Treatment protocol Multi-session FMT via colonos- copy FMT preparation Circle double are accepted. | 35 (±11) | 62.36% | 8.07 (±2.00) Mean Mayo Clinic score | FMT treatment at weeks 0, 2, 6, 10, 14, 18 and 22 | Single-centre prospectively analysis patients with active UC treated with FMT. Predictive clinical biomarkers of |
| Okahara <i>et al</i> (2020) ²⁹ | 92 | Antibiotic treatment alone (n = 37) | Treatment protocol Antibiotic per treatment fol- lowed by FMT (n = 55) FMT preparation Single donor per patient | Mono-AFM arm − 42.5 (±14.7) A-FMT arm − 40.1 (±13.3) | Mono-AFM arm - 48.7% A-FMT arm - 69.1% | Mono-AFM arm — 1.8 (±0.8) Mayo Endo- scopic score A-FMT arm — 1.9 (±0.7) Mayo Endo- | Patients received antibiotic pre-treatment for 2 weeks prior to fresh FMT by colonoscopy | Clinical response was observed at 4 weeks post-treatment and maintenance response observed at 24 weeks post-treatment |
| Zhao et <i>al</i> (2021) ³⁰ | 911 | No active comparator | Treatment protocol Variable infusions / treatment sets of FMT delivered via vari- ous routes — upper Gi, lower Gl or capsule. FMT preparation Unclear | Not reported | Not reported | scopic score Not reported | Undear | Retrospective review of UC patients treated with FMT. Explored early recurrence defined as an increase in Mayo score by ≥ 2 within one week of FMT. |
| Goyal et al (2018) ³⁴ | 72 | Before vs after FMT treatment (+ comparison against the healthy donor profile) | Treatment protocol FMI delivery via faceal suspension into the distal duodenum or proximal jejnum, followed by a flush of normal saline and then delivery of faceal suspension into the terminal ileum and right colon FMI preparation Single donor per patient | 12 ⁹⁻²¹ | 57.1% | Mayo Endoscopic score 1 – 33.3% Mayo Endoscopic score 2 – 41.7% Mayo Endoscopic score 3 – 16.7% | Patients received 1 infusion | response explored response explored Patients treated with antibiotics (metronidazole/ancomycin) for 5 days starting 7 days before FMT Patients also took ome prazole (or equivalent) for 7 days starting 5 days prior to FMT All participants received 24mg of toperamide 2 hours prior to FMT Clinical response and adverse events were assessed at 1 week, 1 month, and 6 months after FMT |
| Table 2 (Continued) | (þa | | | | | | | |

| Reference | No. of Subjects | Control/Comparator | Treatment | Median / Mean Age (years) | Gender (% Male) | Average disease severity indices at baseline | Treatment duration | Relevant study characteristics |
|---|--------------------|---|--|------------------------------|-----------------|--|---------------------------------|---|
| Uygun <i>et al</i> (2017) ³⁸ | 30 | Before vs after FMT treatment | Treatment protocol Pre-treatment with bowel lavage followed by FMT via endo- scopic infusion catheter FMT preparation Single donor per patient | 34.6 (±10.3) | 46.7% | Severe disease – 66.7% Moderate disease – 33.3% | Patients received 1 infusion | Fresh stool samples from the donors were collected Clinical remission and response rates were calculated for par- ticipants at week 12 post-FMT |
| Nishida <i>et al</i> (2016) ³⁶ | 41 | Before vs after FMT treat- ment (+ comparison against the healthy donor profile) | Treatment protocol Single FMT infusion via colonos- copy FMT preparation Single donor per patient | 39.6 (±16.9) | 68.3% | 5.6 (±2.4) Full Mayo score | Patients received 1 infusion | Primary end point — clinical response at 8 weeks |
| Gogokhia <i>et al</i> (2019) ³⁹ | 20 | Before vs after FMT treat- ment (+ comparison against the healthy donor profile) | Treatment protocol Two-donor FMT via colonoscopy to the terminal ileum FMT preparation Unclear if multiple / pooled donors or single donor per patient | | | | Patients received 1 infusion | Faecal samples collected pre- FMT, week 2 and week 4 post- FMT |

Table 2: Non-placebo controlled blinded randomised trials and non-randomised studies of FMT in ulcerative colitis*

- * consisting of \geq 10 patients in FMT treated arms

 † non-placebo controlled blinded randomised trialsFMT-faecal microbiota transplantation, cFMT-capsulised faecal microbiota transplantation, dFMT-donor FMT, aFMT-autologous FMT, SCCAI-simple clinical colitis activity index.

| Reference | Bioinformatic methodology | lpha+eta Diversity (after FMT) | Taxonomic Changes (after FMT) | FMT) |
|---|---|---|---|--|
| | | Responders Non-responders | Responders | Non-responders |
| Paramsothy et al (2019) ³³ | 165 FRNA analysis MOTHUR pipeline Shotgun metagenomics Filtering - DeconSeq, FastQC Analysis - SolexaQA, MetaPhlAn2, HUMAnN2. | In all patients α -diversity \uparrow (Phylogen ett., richness and Shannon's diversity, $P < 0.0001$). β -diversity (multivariate dispersion) changed ($P = 0.0001$) following FMT, however these were more pronounced in patients entering remission | ↑ faecal + mucosal species richness ↑ Eubactenium hallii (Firmicutes), Roseburia inuli- nivorans (Firmicutes Lachnospiraceae), Egger- thella species and Runinococcus bromii (Firmicutes Runinococcus) ↑ Firmicutes (Scillibacter and Clostridum XVIII) | ↓ faecal + mucosal species richness ↑ Fusobacterium (Fusobacterium gonidiaformans) (most consistent association), Sutreella (Sutreella vadsworthensis), Haemophilus, Escherichia, Megamonas, Clostridium XIVa, Prevotella (Prevotella copri) Dialister, Velionella and Bilophila |
| Moayyedi <i>et al</i> (2015) ⁸ | 165 rRNA analysis Analysis — Phyloseq R package and QIIME. | α -cliversity not reported Significant change in β -cliversity (Bray-Curtis dissimilarity) following FMT with no association with clinical response. ($\rho=0.02$) | † Lachnospiraceae family and <i>Ruminococcus</i> in donor B (associated with successful FMT) | † Escherichia and Streptococcus in donor A |
| Costello <i>et al</i> (2019)° | 165 rRNA analysis Unspecified in-house and open source software. Differential abundance analysis Ine4, mice, and gimmTMB R packages | ↑ α-diversity (OTU analysis) in all patients following FMT with no association with clinical response. β-diversity not reported | ↑ Methanobrevibacter smithii, Peptococcus niger (Firmicutes), Faecalicoccus pleomorphus (Firmicutes), Osenalia sp. (Actinobacteria), Acidaminococcus intestini (Firmicutes), Senegalimussilia anaevolia (Actinobacteria), Prevatella copri (Bacteroidetes), Clostridium methylpentosum (Firmicutes), Alistipes indistincus (Bacteroidetes), Slackia isoflavoniconvertens (Actinobacteria) and Odoribacter splanchnicus strain (Bacteroidetes) ↓ Anaerostipes caccae, Gordonibacter pamelaeae and Clostridium adenense Abundance change in Anaerofilum pentosovorans (Firmicutes), Bacteroides coprophilius (Bacteridetes), Castridium nethylpentosum (Firmicutes), cutes), Acidaminococcus intestini (Firmicutes), | Abundance change in Fusicatenibacter saccharivorans (Firmicutes) and Paraprevotella sylaniphila (Bacteroidetes) |
| Rossen et al (2015) ¹⁰ Grothers et al | 165 rRNA analysis USEARCH algorithms and unspecified independent classification techni- ques. Differential abundance analy- sis using Canoco5 165 rRNA analysis | † α-diversity (Shannon's No change in diversity index, P = 0.06 (FMT-D), P = 0.01 (FMT-A)) β-diversity shift (redundancy) No change in α-diversity (Shannon Index) | Sempaginimossilia anaerobia (Actinobacteria) † Clostridium IV, XIVa and XVIII (Firmicutes)(FMT-D | None presented |
| (2021) ¹³ Pai et al (2021) ¹² Haifer et al (2021) ¹¹ | QIIME2 pipeline 165 rRNA analysis Custom Perl scripts, Phyloseq R package and QIIME. 165 rRNA analysis MOTHUR pipeline | α -diversity not reported β -diversity (unspecified measure) changed in FMT arm (not significant) — no association with clinical response α diversity (richness) \uparrow in all patients with changes seen in β -diversity (ANOSIM) following FMT. However, no change in | Alistipes spp. and Excherichia spp. associated with achieving composite clinical outcome Increase in Bacteroides OTU19 (100% similarity to Bacteroides ovatus and Bacteroides | None presented Increase in Bacteroides OTU14 (100% similarity to <i>Bacteroides caccae</i> increase |
| Tian <i>et al</i> (2019) ²¹ | 165 FRNA analysis Full pipeline not described. Differential analysis using LEFSe. | α or β in relation to response or non-response. α or β in relation index and α -diversity (Shannon index and α -diversity (ANOVA), unchanged following FMT in all patients with no association with clinical response. | sylanisoivens) † Bacteroidetes, Proteus, Prevotella, Phascolarcto-bacterium and Lactobacillus (11), Clostridiaceae (42) ‡ Firmicutes, Streptococcus | † Bacteroidetes, Proteus ↓ Firmicutes, Streptococcus |
| Li et al (2020) ²² | 165 FRNA analysis Combination of MOTHUR, UPARSE and R | α -diversity \uparrow (Shannon index and Chao Lindex) β -diversity (MDS) shift (trend) (both analogous to the donors). No separate data in responders | ↑ Holdemania Anaeostipes, Bifidobacterium, Clostridium IV and Odoribacter (analogous to donors) Eubacte- rium and Ruminnoccccus (close to donors) Differences in relative abundance of Eggerthelia, Lactobacillus and Ruminocccus positively cor- related to efficacy (P < 0.05) | Notable difference in Eubacterium and Ruminococcus abundance compared with donors ($P<0.001$) |
| Table 3 (Continued) | (þa | | | |

| Reference | Bioinformatic methodology | α + β Dive | rsity (after FMT) | Taxonomic Changes (after F | MT) |
|--|--|--|---|--|---|
| | | Responders | Non-responders | Responders | Non-responders |
| Leonardi <i>et al</i> (2020) ³² | ITS1 analysis BLAST with ITS1 database filowed by QIIME v1.6 Bacterial analysis as per Paramsothy et al. (2019) ³³ | \uparrow bacterial α -diversity (\uparrow <i>Candida</i> pre-FMT had $\uparrow \alpha$ No change to mycobiota divo No association with clinical | • | Reduction in abundance of <i>Candida</i> positively associated with clinical response | No change in relative abundance of <i>Candida</i> |
| Kump et al (2018) ²³ | 16S rRNA analysis Combination of UCHIME, MOTHUR | No change in α -diversity (ric Significant change in β -dive | | ↑ Akkermansia muciniphila ↓ Dialister | No increase in A. muciniphila |
| Jacob <i>et al</i> (2017) ²⁴ | and QIIME v1.8 165 rRNA analysis USEARCH and UPARSE algorithms / pipelines | distance) α -diversity \uparrow (OTUs P = 0.004) Difference in β -diversity (Bra < 0.034). No association | y-Curtis dissimilarity) post-FMT (P | No taxonomic data presented | |
| Fang et al (2021) ²⁵ | 165 rRNA analysis Full pipeline not described. Differential analysis using LEfSe. | No difference in α -diversity | • | ↑ Bacteroidetes and <i>Prevotella and</i> ↓ Proteobacteria and <i>Escherichia</i> post FMT. Association with clinical response data not presented. | |
| Cui <i>et al</i> (2015) ²⁶ | 165 rRNA analysis Not described. | (n=4). | ormed on a subset of patients | No taxonomic data presented | |
| Chen et al (2020) ²⁷ | 16S rRNA analysis UPARSE and QIIME v1.7 | $\uparrow \alpha$ -diversity (Shannon inde no association with clinic | x) week 4 but then ↓ at week 12 — al response | \uparrow <i>F. Prausnitzii</i> (P < 0.05) — no association with clinical | ll response |
| Brezina <i>et al</i> (2021) ²⁰ | 16S rRNA analysis QIIME2 pipeline. Differential analysis using LEfSe. | α-diversity ↑ (Shannon entre | ppy index) | † Bacteroidales, Prevotellaceae, Veilllonellaceae and Desulfobacteria | ↑ Staphylococcaceae, Lactobacillaceae and Bifidobacteriaceae |
| Fuentes <i>et al</i> (2017) ³¹ | 165 rRNA USEARCH algorithms and unspecified independent classification techniques. Differential abundance analysis using Canoco5 | Analysis of TURN patients | | ↑ Clostridium XIVa (Anaerostipes caccae, Copro- coccus eutactus or Eubacterium rectale (similar levels to healthy donors)) ↓ Enterococcus, Proteobacteria Positive association to Clostridium IV (F. prausnit- zii) and XIVa (Eubacterium hallii, Roseburia intestinalis and Butyrivlbria crossotus) | ↓ Clostridium XIVa (Anaerostipes caccae, Coprococcus eutactus or Eubacterium rectale) ↑ Enterococcus, Proteobacteria and R. gnavus (P = 0.014) Positive association with Bacteroidetes groups (B. vulgatus and B. fragilis) |
| Goyal <i>et al</i> (2018) ³⁴ | 165 rRNA analysis QIIME pipeline. Differential analysis using LEfSe. | † α-diversity (OTU) Change in β-diversity (weighted UniFrac) - both seen 1-month post-FMT. No statistically significant difference in α-diversity seen at 6 months post- FMT | No significant increase in α -diversity (OTU) at 1- and 6-months post-FMT No change in β -diversity (weighted UniFrac) 1- month post-FMT | ↑ Lachnospiraceae and ↓ Enterobacteriaceae at 1 week 1 month and 6 months post-FMT | ζ |
| Nishida <i>et al</i> (2016) ³⁶ | 165 rRNA analysis Full pipeline not described. Phyloseq R package for diversity analysis | No difference in α - and β -di index) at week 8 | versity (Bray-Curtis dissimilarity | No taxonomic data presented | |
| Gogokhia et al (2020) ³⁹ | Virome analysis Filtering using BBMAP following by analysis usin VirMAP pipeline | Not reported | | No change in relative abundance of Caudovirales bacteriophages 4 weeks post FMT | Increase in relative abundance of Caudovirales bacteriophages 4 weeks post FMT |

Table 3: Studies characterising changes in microbial diversity and profiles following FMT for UC
FMT-faecal microbiota transplantation, OUT-Operational taxonomic units, QIIME-Quantitative Insights Into Microbial Ecology, LEfSe-Linear discriminant analysis Effect Size, MDS-Multidimensional scaling

shifts were not however observed in the patients treated with autologous FMT.

Five non-randomised studies measured changes to beta diversity in UC patients receiving FMT. ^{21-24,35} Of these three studies demonstrated a change in beta diversity following FMT relative to baseline community profiles. ^{23,24} The study by Jacob *et al* and Goyal *et al* demonstrated that this shift in the beta diversity resulted in a greater similarity with the donor faecal microbiota. ^{24,34} A similar donor-recipient similarly trend in beta diversity was observed by Li *et al* however no clear difference between responders and non-responders following FMT was seen. ²²

Taxonomic changes

Six of the seven eligible placebo controlled RCTs reported on microbial taxonomic changes following FMT through analysis of stool 16S rRNA profiles as presented in Table 3.⁷⁻¹² In addition, the FOCUS trial performed stool metagenomic analysis and 16S rRNA on colonic mucosal biopsies collected at baseline and at the end of the FMT treatment period (week 8).³³

Changes associated with response to FMT

A significant increase in taxa belong to the Clostridia class (specifically XVIII) in responders to FMT were observed in four RCTs. 8,9,31,33 Notably within this class and increase in taxa belonging to the families Oscillospiraceae (Ruminococcus bromii, Anaerofilum pentosovorans, Clostridium methylpentosum), Lachnospiraceae (Roseburia inulinivorans, Eubacterium hallii) and Clostridiaceae was observed in responders. Increases in taxa belonging to the Clostridia class were also reported in several of the non-randomised FMT studies. Faecalibacterium prausnitzii was reported to significantly increase in responders 4 weeks post-FMT relative to baseline.²⁷ A significantly lower relative abundance of Ruminococcus and Eubacterium compared to healthy donors was reported in non-responders to FMT in a study by Li and colleagues with a non-significant increase in Ruminococcus in responders.22

Four studies reported a significant increase in taxa belonging to phylum Bacterioidetes following FMT in responders. ^{9,11,12,31} Specifically, these included *Bacteroides coprophilus, Bacteroides OTU19* (100% similarity to *Bacteroides ovatus* and *Bacteroides xylanisolvens*) and *Alistipes spp.*

In addition to Clostridia and Bacteroidetes, a significant increase was reported in Eggerthella (Actinobacteria), Senegalimassilia anaerobia (Actinobacteria), Acidaminococcus intestine (Negativicutes) and Escherichia (Proteobacteria). Within the non-placebo controlled or non-randomised studies Brezina and colleagues demonstrated a significant increase in Bacteroidales, Prevotellaceae, Veillonellaceae and Desulfobacteria in responders.²⁰

A significant increase in taxa belonging to the order *Bacteroidales* and *Verrucomicrobiales* and class *Coriobacteriia* was noted in responders compared to non-responders in another study.²² The non-randomised paediatric FMT study by Goyal *et al* demonstrated a significant decrease in *Enterobacteriaceae* and an increase in *Lachnospiraceae* following FMT, however this difference was not significant when sub-grouped by response.³⁴

Analysis of the gut mycobiome, as part of a post-hoc analysis of the FOCUS trial noted that decreased *Candida* abundance post-FMT was indicative of clinical response.³² The LOTUS study in contrast did not report any changes in alpha or beta diversity metrics of the mycobiome upon disease flare.¹¹

Changes associated with lack of response to FMT

Changes in microbial taxa associated with lack of response to FMT were reported by four RCTs. 8,9,11,33 These included a significant increase in species belonging to phylum Fusobacteria (Fusobacterium gonidiaformans), phylum Proteobacteria (Bilophila, Haemophilus, Escherichia, Sutterella wadsworthensis) and family Prevotellaceae (Paraprevotella xylaniphila, Prevotella copri). In addition, a significant increase in Dialister, Veillonella, Megamonas, Fusicatenibacter saccharivorans, Clostridium XIVa and Bacteroides OTU14 (100% similarity to Bacteroides caccae) was observed in non-responders. Responders in the LOTUS trial who developed a disease flare on FMT withdrawal had an enrichment of Streptococcus OTU45 (100% similarity to Streptococcus parasanguinis and other phylogenetically related species) along with depletion of Blautia OTU35 (100% similarity to Blautia faecis). II No clear alpha diversity change was however noted. Within the non-placebo controlled or non-randomised studies Brezina and colleagues demonstrated that Staphylococcaceae, Lactobacillaceae and Bifidobacteriaceae were significantly higher in non-responders.²⁰

Metabolomic analysis

Two RCTs^{9,33} analysed changes in microbial metabolites following FMT treatment. The FOCUS trial identified 97 metabolites that were different between baseline and following FMT treatment regardless of clinical response.33 Of these metabolites, N-acetylmuramate, xanthine, 2-deoxyinosine, ribothymidine and X- 17009 (unnamed biochemical) were significantly increase post-FMT but were not altered by placebo. The trial reported significant differences in global metabolomic profiles following FMT in clinical responders in comparison to baseline, after placebo and after FMT in clinical non-responders. Specifically, 228 metabolites differentiated between positive and negative outcomes following FMT of which 33 of these were different in patients achieving clinical response. Metabolites such belonging to benzoate degradation, glycerophospholipid

metabolism, secondary bile acid biosynthesis, ppGpp biosynthesis and biosynthesis of ansamycins pathways were associated with positive outcomes following FMT. In contrast metabolites associated with heme and lysine metabolic pathways were associated with a negative outcome after FMT. Faecal metabolome analysis in the Costello study that was specifically targeted to short chain fatty acid levels reported no significant differences from baseline in stool concentrations of butyrate, acetate, propionate, iso-butyrate, valerate, iso-valerate and caproate following FMT regardless of clinical response or treatment arm (donor versus autologous).⁹

Whilst TURN trial did not report changes in faecal metabolic profiles they performed functional predictive analysis using PICRUSt and qPCR.31 Microbiota of nonresponders in this study had a significantly lower butyrate production capacity, reflected by the butyrate-acetoacetate CoA transferase and ButCoA gene copies, compared with donors and responders. ButCoA levels were increased by 6.7-fold in responders, especially those who remained in remission at ≥1-year FU. A nonrandomised study that used similar predictive functional analysis gut microbiota reported on significant differences in pathways of pyruvate metabolism, sulfur metabolism, pantothenate and CoA biosynthesis, glyoxylate and dicarboxylate metabolism, synthesis and degradation of ketone bodies and other transporters were between donor, pre- and post-FMT groups.²⁵

Donor characteristics association with clinical response

Two RCTs that explored donor recipient association demonstrated that microbial profiles of recipients were significantly more similar to their respective donors following FMT compared to controls as presented in Table 4.8,31 Notably the study by Moayyedi and colleagues noted that one particular donor, 'Donor B', was associated with greater success rate) in their respective recipients with a non-significant trend for faecal microbiota from responders having greater similarly to donor B than non-responders. 8

Four RCTS reported on the association of clinical response with taxonomic characteristics in donor stool with inconsistent findings. 8,11,31,33 Abundance of specific taxa belonging to Bacteroidetes phylum within donor stool and correlation with a favourable clinical response have been observed in both the FOCUS and LOTUS clinical trials. As the FOCUS trial used pooled FMT, specific donor-recipient relationships could not be explored. Effective donor batches leading to >50% remission in patients contained a higher abundance of Bacteroides OTU187, specifically Bacteroides fragilis and Bacteroides finegoldii, whilst ineffective batches were associated with Clostridium XIVA. There was also a nonsignificant trend towards an association between ineffective batches and the taxa Bacteroides uniformis, Bacteroides coprocola, Sutterella Wadsworthenesis

Streptococcus OTU56. The LOTUS study manufactured oral lyophilised FMT capsules from two separate donors. They demonstrated that the donor with a significantly higher bacterial diversity (greater species evenness) with significant differences in relative abundances of Bacteroidetes taxa was associated with a favourable clinical response. Higher taxonomic classification was however not provided in the study. An open label nonrandomised study demonstrated that clinical response was significantly greater donors with a higher abundance of faecal *Bifidobacterium*, *Lactobacillales* and *Clostridium* clusters IV and XI.³⁶ No significant difference in donor-recipient gut microbial similarity was observed between responders and non-responders.

Moayyedi and collegues noted that Donor B had enrichment of *Lachnospiraceae* and the genera *Ruminococcus*. In contrast, the TURN study observed a greater abundance of *Ruminococcus gnavus* in donors of patients who relapsed compared with donors of patients who achieved sustained remission. However, post-hoc analysis of the TURN study with at least one year follow up of patients in this trial observed that donor faecal samples consisting of *E. coli* and *Aeromonas* were positively associated with patients who relapsed.³¹

Donor (and recipient) faecal microbiome optimisation prior to stool collection and FMT administration was explored in the CRAFT UC study. 19 A specific diet named UC exclusion diet (UCED) was administered as part of this study and comprised mandatory foods such as certain fruits and vegetables, prescribed amounts of chicken and eggs and certain foods that were restricted with the aim of decreasing exposure to sulphated amino acids, total protein, heme, saturated fat and food additives. Donor and recipient dietary conditioning UCED was attempted with patients randomised to either Group I - standard low intensity FMT followed by standard diet, Group 2 - low intensity FMT from donors pre-conditioned with UCED and post FMT recipient conditioning with UCED or Group 3 - UCED alone. Numerically higher, but not statistically significant clinical remission rates and mucosal healing in Group 3 (UCED alone) compared to the FMT arms (Groups 1 and 2). The authors showed that the UCED diet preconditioning of donors reduced the alpha diversity of donor stool microbiota rather than an anticipated increase. Recipient microbiome data or donor-recipient response was not presented as part of the study.

Baseline predictors of response

Clinical predictors. Baseline clinical predictors were reported in three RCTs and two non-randomised studies. Using demographic information obtained from baseline questionnaires, Moayyedi *et al* reported a trend towards patients receiving immunosuppressant therapy

| Reference | Donor Relationship (after FMT) | | |
|---|---|---|--|
| | Responders | Non-responders | |
| Paramsothy et al (2019) ³³ | ↑ homogeneity in taxonomic profiles to a level seen in donors Donor batches with ↑ Bacteroides OTU187 (Bacteroides fragilis and Bacteroides finegoldii) | Donor batches with ↑ Clostridium XIVA and association with Bacteroides unifo mis, Bacteroides coprocola and Strepto coccus OTU56 | |
| Moayyedi <i>et al</i> (2015) ⁸ | ↑ microbiota similarity to donor B (enrichment of <i>Lachnospiraceae</i> and <i>Ruminococcus</i>) | \downarrow microbiota similarity to donor B | |
| Rossen <i>et al</i> (2015) ¹⁰ | • Microbiota composition overlap with healthy donors (FMT-D) characterised by | ↓ similarity index to corresponding donors | |
| | ↑ Clostridium clusters IV, XIVa, XVIII and ↓ Bacteroidetes • Microbiota composition shift away from non-responders (FMT-A (different direction to FMT-D responders)) characterised by ↑ Bacilli, Proteobacteria and Bacteroidetes ↑ similarity index to corresponding donors | ↓ similarity to donors which they received faeces from (P = 0.02) | |
| Haifer <i>et al</i> (2021) ¹¹ | ↑ similarity to donors which they received faeces from (P = 0.02) Donor 1 (favourable donor) had a significantly higher bacterial diversity driven by higher species evenness with compositional differences largely | Not reported | |
| Jacob <i>et al</i> (2017) ²⁴ | related to differences in relative abundances of Bacteroidetes taxa † Similarity with donor FMT samples Donors achieving clinical remission clustered together | Not reported | |
| Chen <i>et al</i> (2020) ²⁷ | Abundance of <i>F. prausnitzii</i> ↑ towards levels similar to those of donors | Not reported | |
| Li et al (2020) ²² | \downarrow Dissimilarity between patients and donors ($\alpha+\beta$ diversities analogous to donors) | \downarrow Dissimilarity between patients and donors ($\alpha + \beta$ diversities analogous to donors) | |
| Fuentes <i>et al</i> (2017) ³¹ | \uparrow Similarity to donors (FMT-D) (P = 0.02) Trend of \uparrow similarity to donors (patients with sustained remission) (P = 0.1) | \downarrow Similarity to donors (FMT-D) (P = 0.02) Trend of | |
| | No significant differences in similarity values of FMT-A patients | ↓ similarity to donors (relapsers) (P = 0. Donor batches associated with Proteo- bacteria (E. coli and Aeromonas) and | |
| | | ↑ abundance of <i>Ruminococcus gnavus</i> No significant differences in similarity v ues of FMT-A patients | |
| Kump <i>et al</i> (2018) ²³ | All recipients' microbiotas, regardless of response, shifted towards the respective donor microbiota | All recipients' microbiotas, regardless of response, shifted towards the respec- tive donor microbiota | |
| Shabat <i>et al</i> (2021) ¹⁹ | UCED preconditioning of donors led to reduction of alpha diversity of donor stool with numerically higher remission rates compared with FMT alone (or UCED and FMT). | | |
| Okahara et <i>al</i> (2020) ²⁹ | ↑ Cumulative non-relapse rate in sibling FMT than parent-child FMT Donor Bacteroidetes species (<i>Bacteroides uniformis</i> and <i>Parabacteroides distasonis</i> and <i>Bacteroides dorei</i>) persisted in patients with no UC recurrence after 24 months | ↓ Cumulative non-relapse rate in ≥11- year difference group that 0-10-year difference group | |
| | ↑ Similarity of 10 Bacteroidetes species to donor levels | | |

Table 4: Data summary table of the relationship between patients' and donors' microbiota post-FMT

FMT-faecal microbiota transplantation, FMT-D-donor faecal microbiota transplantation, FMT-A-autologous faecal microbiota transplantation, UCED-Ulcerative colitis exclusion diet

at baseline acquiring a greater benefit from FMT.⁸ Additionally, the authors found that patients were statistically significantly far more likely to respond to FMT if they had received a recent diagnosis of UC (defined as ≤I year). In contrast the FOCUS trial observed an

inverse relationship between endoscopic severity and the primary outcome however this was no longer seen when controlled for other factors.⁷ Correlation with clinical response was also noted with age but directionality was not reported. No relation was however observed between the primary outcome and anatomical disease extent, smoking status, disease duration, any concomitant immunosuppressive (steroids, biologics, immunomodulatory) use. Similarly, the RCT by Costello *et al* did not observe any interactions between age at diagnosis/randomisation, disease duration/distribution, gender, non-steroid based medication use or macronutrient intake with a change in total Mayo score following donor FMT. Use of oral steroids at randomisation was however associated with a greater reduction in total Mayo score.

Amongst the non-randomised studies, in a singlecentre prospectively study of open label FMT statistically significant association between moderate disease severity (Mayo score 6-9) and remission in UC patients, along with endoscopic Mayo score 2.28 In addition, the authors noted that severe disease (Mayo score >10) and endoscopic Mayo score 3 were both significantly correlated with FMT failure. A previous study by the same authors reported that patients treated earlier on in the disease course or those with mild disease had higher rates of clinical remission.³⁷ They noticed that in biologic-experienced patients, endoscopic Mayo score 2 was a predictor of response whereas in biologic-naÿve patients younger age, moderate disease severity, shorter disease duration and endoscopic Mayo score 2 were all significantly predictive of a positive outcome. They described young age as a baseline factor which determined participants' response with patients under 40 years demonstrating greater rates of remission. In a univariate analysis performed in an uncontrolled study that consisted II disease recurrences in II6 UC patients with active disease reported significant associations with a baseline high Mayo score, recent use of steroids to induce remission, low serum albumin, and peripheral blood lymphocyte deficiency were associated with a higher recurrence rate following FMT.30 These are however recognised factors associated with unfavourable disease outcomes irrespective of treatment. No association with disease extent was observed and disease duration was not explored. Two other non-randomised studies did not demonstrate any differences in clinical characteristics between responders and nonresponders.36,38

Microbial predictors. Analysis of potential baseline microbial predictors of response in the FOCUS trial found that patients who achieved the primary outcome tended to have higher faecal species richness at baseline compared with patients not achieving the primary outcome.³³ They also observed a similar non-significant trend in the mucosal microbiome in which a higher baseline species richness as well as an increased abundance of specific species of Bacteroides (*B. fragilis* and *B. finegoldii*) with was associated with a positive therapeutic outcome(33). Gut mycobiome analysis of the

FOCUS trial observed that a greater abundance of *Candida* pre-FMT was associated with a clinical response (and increased bacterial diversity post-FMT).³² An open label study of 20 patients with active UC observed that FMT responders had a lower relative abundance of *Caudovirales* bacteriophages at baseline compared to non-responders. The relative abundance of *Caudovirales* in non-responders appeared to increase after FMT while no change was observed in responders.³⁹

Patients receiving autologous FMT in the TURN trial had a greater likelihood of response to treatment if they possessed baseline microbiota profiles more similar to donor samples or to patients in sustained remission following donor FMT. Differences in baseline microbiota profiles between responders and non-responders was however not found to be a predictor of response for patients receiving donor FMT.³¹ Higher levels of Bacteroidetes, particularly *B. vulgatus*, and *Prevotella* in non-responders at baseline were associated with relapse at the I year follow up. A non-randomised study of FMT in paediatric patients with UC observed that the abundance of Fusobacterium was significantly greater at baseline in non-responders compared to responders.³⁴

Metabolomic predictors. Potential baseline metabolomic predictors of response was only reported as part of the FOCUS study.³³ Fifteen metabolites were identified-N-methylphenylalanine, N-acetylarginine, caproate, lignoceroyl ethanolamide, biotin were associated with an increased positive clinical outcome whilst the metabolites 5-aminovalerate, oleoyl-arachidonoyl-glycerol, linoleoyl-arachidonoyl-glycerol, sphingomyelin, sphingomyelin, gulonate and heme were identified as being associated with increased negative outcome.

Discussion

This systematic review outlines potential donor and recipient clinical and microbial biomarkers that predict and denote clinical response to FMT in patients with UC. Examination of 7 double blind placebo controlled RCTs and 12 non-randomised studies in FMT in UC identified specific consistent findings in gut microbial profiles that correlate with a favourable clinical response along with clinical and microbial profiles that have the potential of predicting response to FMT (summarised in Figure 2).

Following FMT, the overall trends of biomarkers discovered in responding patients' microbiota communities were (a) an increase in bacterial diversity (alpha and beta), (b) increases in Firmicutes and Bacteroidetes along with key taxa belonging to these phyla, and (c) recipient microbial profiles with increased similarity to donor profiles. Responders of FMT had microbial profiles more similar to that of their donors, possibly

FMT Donors Patients with UC Responders Increase in faecal microbial alpha diversity. abundance of Clostridiales clusters IV / XIVa and Bacterioides, increase in faecal short chain fatty acids concentration Non responders Optimum donor features Baseline predictors of response No change in faecal microbial alpha Greater faecal microbial alpha Younger age, short disease diversity, increase in Proteobacteria, diversity, higher abundance of duration, mild to moderate Terrabacteria and Fusobacteria Lachnospiraceae. Ruminococcus. disease, greater faecal microbial

Summary of key predictors and biomarkers of response to FMT in UC

Figure 2. Summary of key predictors and biomarkers of response to FMT in UC. FMT, faecal microbiota transplantation, UC, ulcerative colitis.

richness

suggesting that the donor microbiota composition profile may be used as a potential microbial treatment target for individualisation of FMT treatment regimens. At a taxonomic level, studies have consistently demonstrated an increase in abundance of Clostridium clusters IV and XIVa (members of the Firmicutes phylum), post-FMT this is associated with a favourable clinical response. These include the Lachnospiraceae and Ruminococcaceae families and are likely to induce this response through immune regulation of colonic inflammatory pathways. 40,41 SCFAs are the product of bacterial fermentation of polysaccharide, oligosaccharide and particular amino acids which are non-digestable by the host.⁴² Producers of SCFAs specifically Clostridium clusters have a crucial role in maintaining intestinal function.43 SCFAs have been shown to induce the differentiation of naÿve CD4 T cells into immunosuppressive, anti-inflammatory IL-10-producing regulatory T cells. 41,44 Consistently SCFA synthesis, and the presence of components contributing to this synthesis, appears to be a metabolomic biomarker of response post-FMT.^{25,31,33,45} For instance, observed gene copy levels of ButCoA were increased in those patients who received successful FMT therapy whilst the microbial capacity for butyrate production of the microbiota decreased in patients lacking a response to FMT.31 The FOCUS study also identified increased levels of heme and lipopolysaccharide biosynthesis at both baseline and post-FMT as potential biomarkers associated with a negative outcome.33 Not only do various bacterial pathogens produce heme, but it is also a vital source of iron required for their survival with murine studies suggesting its role in colonic inflammation.46

Bacteroides fragilis and lower

abundance of Proteobacteria

Certain baseline recipient characteristics were found to be important factors in determining a favourable outcome. FMT recipients with younger age, less severe and less extensive disease and potentially shorter duration of UC (< I year) have been shown to associated with a greater likelihood of response. These predictive baseline factors are not too different to that of biological/small

molecule therapies in UC. Whilst other biomarkers were identified in this systematic review such as the prior or concurrent immunosuppressant use in predicting response, further research is needed to corroborate these findings. There is some evidence to suggest that patients with higher faecal microbial richness at baseline, greater abundance of *Candida*, lower abundance of *Candovirales* and a microbial composition closer to donors at baseline are more likely to have a favourable response. It is plausible that the dysbiosis seen in patients with a recent UC diagnosis as well as a relatively lower degree of microbial aberrancy is more easily manipulated with FMT resulting in a greater likelihood of successful and sustained donor microbiome engraftment and clinical response.

Only the FOCUS study reported on baseline metabolomic predictors of response, with the findings of the RCT provide significant insight into the bacterial metabolites which give a higher likelihood of achieving a positive FMT outcome. One of the most notable metabolites was biotin (vitamin B7), with diet and synthesis by commensal microbiota in the gut being its primary source in humans. Biotin results in the downregulation of the NF- κ B gene thereby restricting release of various pro-inflammatory cytokines in the gut epithelium.

Greater microbial richness in donor stool was associated with an increased rate of clinical response in patients with active UC. 11,23,47 Engraftment of donor-derived microbiota ameliorates UC symptoms through either replenishing bacterial species whose abundance is decreased prior to treatment or, providing bacteria which create an unfavourable environment for disease-associated bacteria so as to repress their growth. 50 Having a high bacterial species richness, therefore, may increase the chances that certain bacterial strains engraft in the gut of the recipient and become permanent members of their microbiota community. 51 Along with increased bacterial richness, specific taxa were identified in donor stool associated with remission,

whilst others were found in those associated with treatment failure. Donor stool which included high abundances of Bacteroides OTU187 in addition to the families Lachnospiraceae and Ruminococcaceae were more likely to induce a response in recipients, whereas the presence of Clostridium XIVA was seen in ineffective batches.^{8,11,33} The TURN study in contrast observed a greater abundance of Ruminococcus gnavus in donors of patients who relapse. However, it is important to note that the microbial profiles of donors were similar to the baseline profiles of the UC patients in this study. Preselecting donors based on a richer microbial diversity and greater abundances of SCFA producing bacteria or pooling FMT from donors to control for variability in donor microbial diversity. Pooling FMT is, however, no longer practical as it presents major challenges with 'look back' exercises and root cause analysis in cases of FMT related adverse events. One option would be to pre-condition donors with a diet that is associated with increasing microbial diversity. The CRAFT UC study attempted this with preconditioning donors with a designer diet (UCED) that consisted of dietary exclusion of specific components such as saturated fat and food additives that are thought to contribute to an immune mediated inflammatory response. 19 Paradoxically the UCED diet resulted in a reduction in donor microbial richness and may have potentially contributed to the unfavourable outcomes seen with donor pre-conditioned FMT. Nevertheless, optimum microbiome-based donor selection as well as pre-conditioning with a diet that is associated with increasing gut microbial diversity are likely to play an important role enhancing response with FMT.52,53

The findings of this systematic review highlight the possibility of enhancing a sustained response to FMT through biomarker-based selection and optimisation of donors and patients before and during the treatment with FMT. Utilising precision medicine, would facilitate an individualised, biomarker driven 'treat to microbiome/metabolome' target approach with FMT in UC early in the disease. After the pre-defined clinical target is reached, the need for further FMT is tracked based on loss of this specified microbiome target. Studies are now needed to help define these targets with leading candidates that include alpha diversity, specific faecal SCFA producing strains such as Clostridiales and faecal butyrate levels. There are a few limitations of this systematic review. The heterogeneity of the study designs that include mode and frequency of FMT administration, the use of a single or pooled donor approaches, variable placebo and active comparators and differences in microbial analytical strategies may make interpretation in the context of a systematic review challenging. However, the reproducibility and consistency of several of the findings reported in this review, in addition to biological plausibility, does bring a level of confidence. We excluded studies with less than ten (FMT treated) participants for quality control. None of these excluded

studies had detailed exploratory mechanistic data that would have significantly influenced the findings in the review.

To conclude, there is evidence of existing predictive biomarkers for the treatment of UC with FMT, the most well-defined of these being microbial indicators. Despite the exponential growth in research into FMT over recent years, the mechanistic understanding on the basis of this treatment is poor. It also remains unclear if alterations to the microbiota occur to certain pre-existing immunomodulatory bacterial strains that are enriched post-FMT, or if they are solely donor derived and engrafted after treatment. It is clear however, that the gut microbiota is fast becoming a pivotal therapeutic target which holds considerable potential.

Contributors

NPR and MNQ performed the search and data extraction. NPR wrote the first draft with critical feedback and edits from MNQ. All authors (NPR, WS, CQ, CT, RDH, NS, ADB, THI, MNQ) provided feedback and approved the final version of the draft.

Data sharing statement

The authors confirm that the data supporting the findings of this study are available within the article [and/or] its supplementary materials.

Declaration of interests

All authors declare no relevant conflict of interests.

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Supplementary materials

Supplementary material associated with this article can be found in the online version at doi:10.1016/j.ebiom.2022.104088.

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