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Bio-orthogonal Red and Far-Red Fluorogenic Probes for Wash-Free Live-Cell and Super-resolution Microscopy

Philipp Werther,^{##} Klaus Yserentant,^{##} Felix Braun, Kristin Grußmayer, Vytautas Navikas, Miao Yu, Zhibin Zhang, Michael J. Ziegler, Christoph Mayer, Antoni J. Gralak, Marvin Busch, Weijie Chi, Frank Rominger, Aleksandra Radenovic, Xiaogang Liu, Edward A. Lemke, Tiago Buckup, Dirk-Peter Herten,^{*} and Richard Wombacher^{*}



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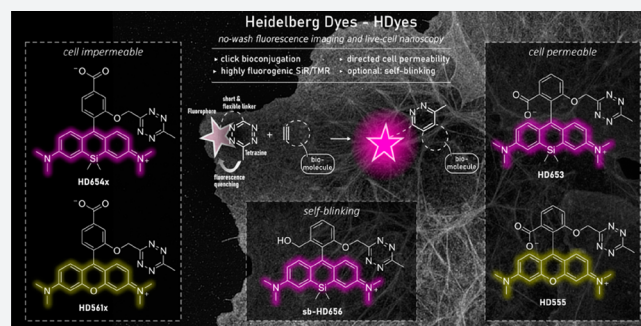


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ABSTRACT: Small-molecule fluorophores enable the observation of biomolecules in their native context with fluorescence microscopy. Specific labeling via bio-orthogonal tetrazine chemistry combines minimal label size with rapid labeling kinetics. At the same time, fluorogenic tetrazine–dye conjugates exhibit efficient quenching of dyes prior to target binding. However, live-cell compatible long-wavelength fluorophores with strong fluorogenicity have been difficult to realize. Here, we report close proximity tetrazine–dye conjugates with minimal distance between tetrazine and the fluorophore. Two synthetic routes give access to a series of cell-permeable and -impermeable dyes including highly fluorogenic far-red emitting derivatives with electron exchange as the dominant excited-state quenching mechanism. We demonstrate their potential for live-cell imaging in combination with unnatural amino acids, wash-free multicolor and super-resolution STED, and SOFI imaging. These dyes pave the way for advanced fluorescence imaging of biomolecules with minimal label size.



INTRODUCTION

Over the last decades, fluorescence microscopy has become an indispensable tool to study biomolecules in their native cellular environment. A fundamental challenge is to achieve specific fluorescent labeling of biomolecules while minimizing the size of the label. This is crucial to reduce the perturbation of the biomolecule's behavior as well as to avoid distortions by linkage errors in super-resolution microscopy.¹ In recent years, bio-orthogonal chemistry has become an important approach to selectively introduce small organic fluorophores. Various strategies have been developed to label proteins,^{2,3} nucleic acids,^{4,5} sugars,^{6,7} or lipids.^{8,9} With the advancement of bio-orthogonal chemistries, probe designs have been reported where bond formation is accompanied by fluorescence enhancement.^{10,11} This so-called fluorogenic effect can substantially improve the signal-to-background ratio in fluorescence microscopy and can allow no-wash experiments.^{12–15} In particular, quenched tetrazine dyes that react in inverse electron demand Diels–Alder (DA_{inv}) reactions bear great potential as fluorogenic bio-orthogonal labels due to fast kinetics and high biocompatibility.¹⁶ Previous work has shown that the quenching efficiency in tetrazine dyes determines the fluorescence turn-on of the probe upon reaction and is strongly dependent on the interchromophore distance between tetrazine and fluorophore.^{17–19} Tetrazine–dye designs with

short interchromophore distances, however, show only very low quenching efficiencies for far-red-shifted fluorophores compared to their blue-shifted analogues.^{14,20,21}

Beyond fluorogenicity and bioconjugation functionality, fluorescent labels need to exhibit high water solubility and should not aggregate or bind unspecifically to cellular structures. Moreover, cell permeability, high fluorescence brightness, and high photostability are crucial for long-term observation of intracellular targets. Rhodamines and silicon rhodamines (SiRs), both xanthene-type dyes, meet these demands and are widely used in super-resolution fluorescence microscopy techniques like single-molecule localization microscopy (SMLM)^{22,23} or stimulated emission depletion microscopy (STED).^{24,25} Different structural designs for fluorogenic tetrazine conjugates have been developed for both dye classes.^{14,26,27} For rhodamines, a fluorescence enhancement up to 76-fold has been reported.²⁶ However,

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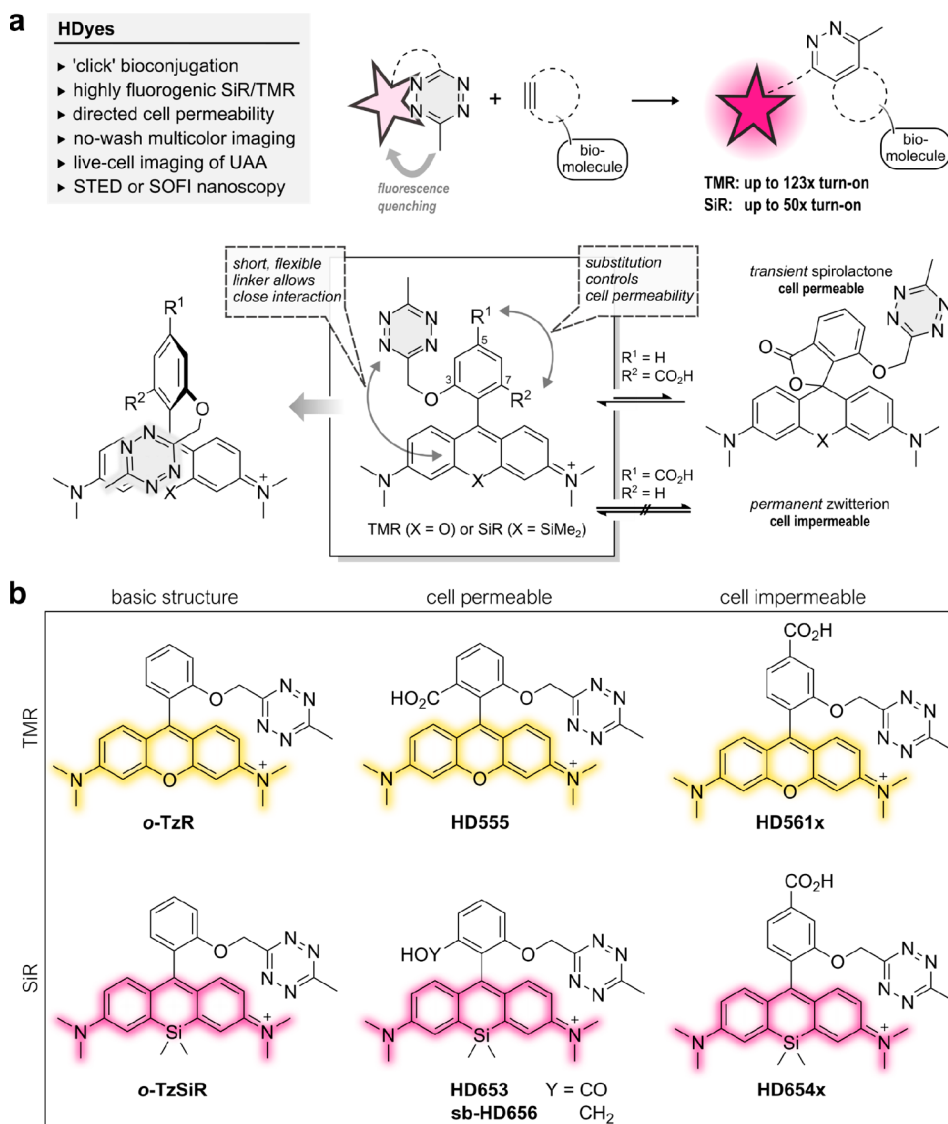


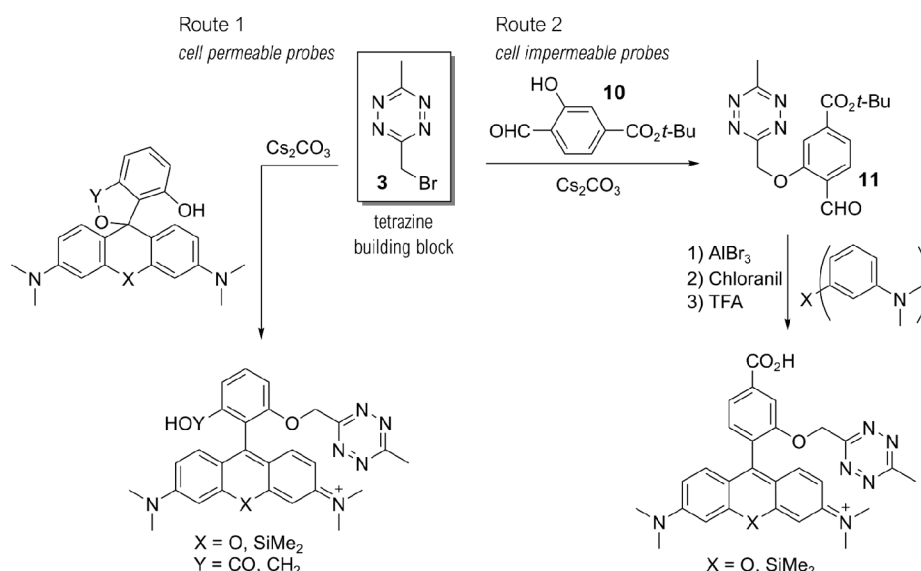
Figure 1. Design rationale for close proximity quenched fluorogenic probes. (a) Tetrazine dyes can be conjugated to dienophile-modified biomolecules accompanied by an increase in fluorescence intensity. The new design allows a close fluorophore–tetrazine interaction, resulting in efficient quenching and high fluorescence turn-on. Cell permeability is controlled by varying the position of a CO₂H substituent. (b) Overview of fluorogenic TMR and SiR tetrazine probes (HDyes) with distinct cell permeability.

so far, quenched SiR–tetrazine conjugates suffer from moderate fluorescence enhancements¹⁴ or do not allow live-cell imaging.²⁷

Here, we report a novel design concept for tetrazine–dye conjugates that enables highly efficient quenching for rhodamines and SiRs resulting in high fluorescence enhancement upon DA_{inv}. We have thoroughly characterized the spectroscopic properties of the tetrazine–dyes and found evidence of Dexter exchange as the underlying fluorescence quenching mechanism. The tetrazine–dye conjugates that we name HDyes (“Heidelberg Dyes”) exhibit excellent properties for a number of live-cell applications including the targeting of unnatural amino acids (UAAs) and allow extra- and intracellular multicolor wash-free labeling. Finally, we demonstrate that bio-orthogonal labeling with HDyes enables live-cell STED as well as super-resolution optical fluctuation imaging (SOFI).

RESULTS AND DISCUSSION

Design and Synthesis of Close Proximity Quenched Fluorogenic HDyes. The main challenge in the development of fluorogenic probes for bio-orthogonal chemistry is to accommodate both high fluorogenicity upon target binding and favorable properties for live-cell imaging in one molecular structure. Generally, an ideal concept enables further structural modification of the fluorophore to provide distinct molecular functionalities for specialized applications. In the case of fluorogenic tetrazine dyes, efficient fluorescence quenching is particularly difficult to achieve with red-shifted fluorophores.²⁰ To address this problem, we designed tetramethylrhodamine (TMR)– and SiR–tetrazine conjugates with minimized interchromophore distances. In this design, the tetrazine is placed in close proximity to the fluorophore by an unconjugated chemical linkage in the *ortho*-position of the phenyl ring pendant to the xanthene core (Figure 1a). We reasoned that the short and flexible oxymethyl linker would lead to a stacked conformation of the fluorophore and tetrazine

Scheme 1. Synthesis of HDyes^a

^aTwo complementary synthetic routes provide access to *ortho*-oxymethyl-linked SiR and TMR tetrazine probes with distinct cell permeability. The key compound for tetrazine derivatization in both routes is the bromomethyl tetrazine building block 3.

as was suggested in previous work from Beliu et al. on oxazine and SiR dyes.²⁸ As a consequence of this shortened bichromophoric linkage, the stacked conformation could become more likely, which in turn would improve fluorescence quenching. Furthermore, building on literature reports, we aimed to control cell permeability by variation of additional phenyl ring substitutions. It has been shown that a carboxylic acid substituent in the *ortho*-position results in high cell permeability owing to the formation of a transient uncharged spirolactone.^{12,13} We anticipated that a carboxylic acid substituent in the *para*-position lacking the possibility of spirolactonization yields a permanent zwitterionic, thus cell-impermeable dye.

To realize such cell-permeable and -impermeable (Si)-rhodamine-tetrazines, we developed synthetic routes using bromomethyl tetrazine building block 3 (Scheme 1, see Schemes S1–4 and Table S1 for detailed description). We synthesized tetrazine benzaldehyde 11 from 3 via Williamson ether synthesis and used them in Friedel–Crafts-reactions¹⁴ to create 3,5-substituted (Si)-rhodamines (Route 2). Subsequent *tert*-butyl ester cleavage afforded cell-impermeable HDyes, HD561x and HD654x (Figure 1b). However, corresponding cell-permeable 3,7-substituted derivatives were not accessible through this route presumably due to inactivation of the benzaldehyde intermediate by hemiacetal formation and problematic ester cleavages. Yet, we were able to obtain phenolic (Si)-rhodamines by addition of bisaryllithium intermediates to phthalic anhydrides²⁹ or phthalides and subsequent MOM deprotection (Route 1, Schemes 1 and S4). These were reacted with 3 in Williamson ether syntheses to furnish the respective 7-CO₂H cell-permeable HDyes HD555, HD653, and sb-HD656 (Figure 1b).

Photophysical Characterization and Investigation of the Quenching Mechanism. Understanding the quenching mechanism of fluorogenic probes is of particular importance for the experimental application and future developments in the field. Therefore, we studied the photophysical properties and the quenching mechanism of *ortho*-oxymethyl-linked tetrazine rhodamines using time-resolved spectroscopy. We

synthesized the basic structure *o*-TzR and found that it is highly quenched, with a fluorescence quantum yield (Φ_F) of 0.3%. In addition to *o*-TzR, we synthesized regioisomers *m*- and *p*-TzR where the oxymethyl-linked tetrazine is in the *meta*- and *para*-positions, respectively (Figure 2a, Scheme S3, Table S2). Further, we reacted *o*-TzR with (bicyclo[6.1.0]non-4-yn-9-yl)methanol (BCN) to isolate the cycloadduct 26 and synthesized the structurally related methoxy-substituted 25, both serving as unquenched reference dyes (Figure 2a). We obtained the intramolecular tetrazine–xanthene distances (*R*) for *o*/*m*/*p*-TzR from geometry-optimized structures using DFT calculations (Figure 2b). These computational results confirmed our hypothesis that *o*-TzR affords a stacked tetrazine–fluorophore conformation with a smaller intramolecular distance between tetrazine and the fluorophore than those of *m*-TzR and *p*-TzR.

We then studied the distance-dependence of the quenching by recording femtosecond transient absorption (TA) spectra with a variable probe delay after a 20 fs excitation pulse at 575 nm. We found that all compounds show ground-state bleach (GSB) and stimulated emission (SE) bands with a maximum at 560 nm (Figures S1, S2 and Note S1). The absorbance at 560 nm in dependence of the probe delay shows the different decays of all five compounds (Figure 2c). Reference compounds 25 and 26 lacking a quenching group showed a minor signal recovery within 1.3 ps followed by a very slow decay compatible with their emission characteristics. In contrast, the TA signal recovery was strongly accelerated for tetrazine regioisomers *o*/*m*/*p*-TzR: While in the case of *m*/*p*-TzR, the TA signal decayed almost to half of its initial amplitude after 16 ps, it was quenched to below 20% of the initial amplitude for *o*-TzR (Figures S1, S2). Accordingly, *m*- and *p*-TzR exhibited substantially lower turn-on in DA_{inv} of 6.9- and 8.5-fold, respectively (Figure S4, Table S1). For a quantitative assessment of the TA data, we performed global target analysis with a sequential model to the full data set (Figure S3 and Note S2). The fast recovery with a time constant τ_2 of 1.56(±0.06) ps for *o*-TzR confirmed an efficient quenching compared to *m*-TzR and *p*-TzR with τ_2 of

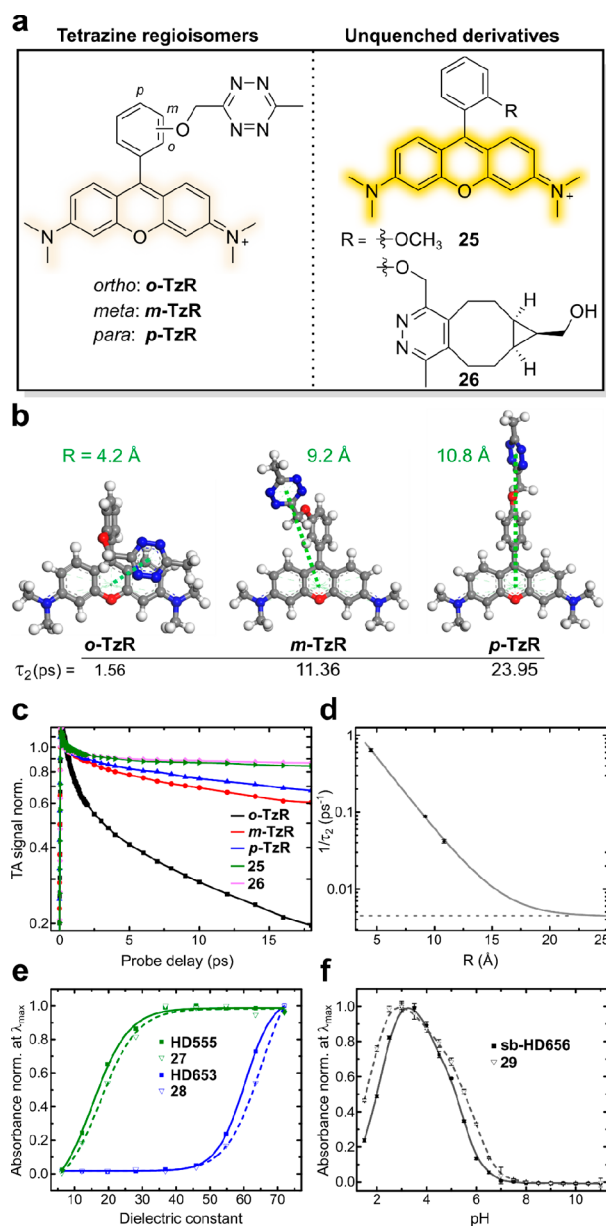


Figure 2. Fluorescence quenching and physicochemical properties of HDyes. (a–d) Time-resolved spectroscopy to study fluorescence quenching: (a) Structures of tetrazine rhodamine regioisomers and unquenched reference dyes. (b) DFT-optimized structures of *o/m/p*-TzR, respective tetrazine–xanthene intramolecular distances (*R*), and transient absorption decay times τ_2 . (c) Selected transient absorption traces at 560 nm (max. of GSB/SE). Data was normalized at a probe delay of 0.5 ps to avoid contributions of the coherence spike and vibration coherence at earlier delay times. (d) Experimental decay rates ($1/\tau_2$) obtained from global fitting of the transient absorption data and its dependence on *R*. Single-exponential fitting was performed assuming a constant offset (dashed horizontal line) obtained from the average lifetimes τ_2 (see Table S3) of nonquenched derivatives **25** and **26**. (e,f) Solvatochromic spirocyclization properties: (e) Normalized absorbance at λ_{max} (zwitterionic form) of 7-CO₂H tetrazine dyes and respective cycloadducts as a function of the dielectric constant of water/dioxane mixtures (v/v; 10/90 to 90/10). (f) Normalized absorbance at λ_{max} (measured in triplicates) of 7-CH₂OH tetrazine dye **sb-HD656** and respective cycloadduct **29** as a function of pH.

11.36(±0.30) ps and 23.95(±1.30) ps, respectively. **25** and **26**, lacking the quenching group, yielded time constants of 258(±10) ps and 196(±3) ps (Table S3). The interchromophore distance-dependence of the decay rate τ_2^{-1} (Figure 2d) for *o/m/p*-TzR could be well described with an exponential fit using an offset from the unquenched reference dyes **25** and **26**. These findings rule out resonance energy transfer (RET) ($\tau_2^{-1} \propto R^{-6}$) and point toward an electron-transfer-based quenching mechanism, like Dexter-type electron exchange or photo-induced electron transfer (both $\tau_2^{-1} \propto e^{-R}$).³⁰ Additionally, the spectral features of the TA signals of *o/m/p*-TzR serve to further differentiate the mechanism (see Note S2). Essentially, for *o*-TzR, GSB, SE, and excited-state absorption (ESA) signals are recovered simultaneously, which we interpret as an indication for a solely Dexter-type electron exchange mechanism. In contrast, for *m/p*-TzR, the GSB/ESA recovery is delayed compared to the SE recovery. Therefore, we conclude that at these higher interchromophore distances, another quenching mechanism begins to contribute.

In TA measurements, we observed that fluorescence quenching occurs from an *ortho*-tetrazine substituent with high efficiency, yielding a low Φ_F and a fast decay time for *o*-TzR. This led to a high fluorescence turn-on of 95-fold in kinetic measurements upon reaction with BCN (Table 1, Figure S4). Importantly, the red-shifted analogue *o*-TzSiR exhibited comparable fluorogenicity with a Φ_F of 0.7% and 45-fold turn-on. Likewise, CO₂H-substituted HDyes showed turn-ons of 123-, 45-, 50-, and 33-fold for **HD555**, **HD561x**, **HD653**, and **HD654x**, respectively (Table 1, Figures S5–7), with the expected fast reaction kinetics upon DA_{inv} product formation (Figure S8).

With regards to live-cell labeling of intracellular targets, we examined the solvatochromic spirocyclization behavior that determines the cell permeability of TMR and SiR dyes. As mentioned above, 3,7-substituted **HD555** and **HD653** can transiently form an uncharged and nonchromophoric spiro-lactone from the zwitterionic and chromophoric open form. Therefore, we studied the solvent-polarity-dependent spiro-lactone–zwitterion equilibrium of the 7-CO₂H dyes. We measured the absorbance of **HD555** and **HD653** along with their respective cycloadducts **27** and **28** in dioxane–water mixtures of varying composition (Figure 2e, Figure S11) to determine $D_{0.5}$, the dielectric constant at half-maximum absorption^{25,31} (Table 1). While **HD555** ($D_{0.5} \approx 17$) resided predominantly in the zwitterionic form even in low-polarity media, **HD653** ($D_{0.5} \approx 60$) had a distinct propensity to adopt to the colorless spiro-lactone, which is in good accordance with reports for other TMR and SiR fluorophores.^{31,32} Of note, **27** ($D_{0.5} \approx 19$) and **28** ($D_{0.5} \approx 63$) had slightly higher propensities to adopt to lactone forms than their respective tetrazine parent dyes. The formation of a nonchromophoric, spirocyclic species is also a central feature of self-blinking hydroxymethyl-substituted SiR dyes. It was shown that the reversible transition from a fluorescent zwitterion to a nonfluorescent spiroether results in fluorophore blinking that enables stochastic, switching-based super-resolution microscopy such as SMLM³³ or SOFI.³⁴ Recently, we showed that the combination of fluorogenic labeling with self-blinking (sb) dyes decreases background localizations in SMLM,³⁵ and environment-sensitive sb-tetrazine probes were used for long-term SMLM.³⁶ Our set of HDyes includes a self-blinking SiR, **sb-HD656**, which was synthesized from 7-hydroxy-phthalide via route 1 (Schemes 1, S4). We evaluated the pH-dependence

Table 1. Spectral Properties of HDyes^a

name	X	R ¹	R ²	λ_{abs} [nm]	λ_{em} [nm]	ϵ [10^4 (M cm) ⁻¹]	Φ_F	D _{0.5}	turn-on	intra- extra- cellular application	
<i>o</i> -TzR (26)	O	H	H	557 (557)	583 (582)	5.52 (7.41)	0.003 (0.447)	-	95	-	-
<i>o</i> -TzSiR	SiMe ₂	H	H	652	671	2.16	0.007	-	45	-	-
HD555 (27)	O	H	CO ₂ H	555 (556)	581 (583)	5.10 (10.0)	0.004 (0.571)	17 (19)	113/123 ^b	+	+
HD55Lx	O	CO ₂ H	H	561	588	7.70	0.007	-	45	-	+
HD653 (28)	SiMe ₂	H	CO ₂ H	653 ^d (652 ^d)	676 ^d (674 ^d)	4.86 ^d (5.92 ^d)	0.022 ^d (0.477 ^d)	60 (63)	50 ^b	+	+
HD654x	SiMe ₂	CO ₂ H	H	654	673	12.5	0.007	-	33	-	+
sb-HD656 (29)	SiMe ₂	H	CH ₂ OH	656 ^c (656 ^c)	676 ^c (677 ^c)	11.4 ^c (10.4 ^c)	0.026 ^c (0.310 ^c)	-	9.0 ^c	+	+

^aAll measurements were performed in phosphate-buffered saline (PBS pH 7.4) and turn-on experiments with 15 eq BCN unless noted otherwise. Properties of respective BCN cycloadducts of the dyes listed in parentheses (see Figure S9 for all structures and Figure S10 for spectra). The assignment of the substituents refers to the general structure in Figure 1a. ^b10 eq of EGFP–HaloTag–BCN. ^cSodium phosphate buffer pH 3.5. ^d50 mM SDS in PBS pH 7.4.

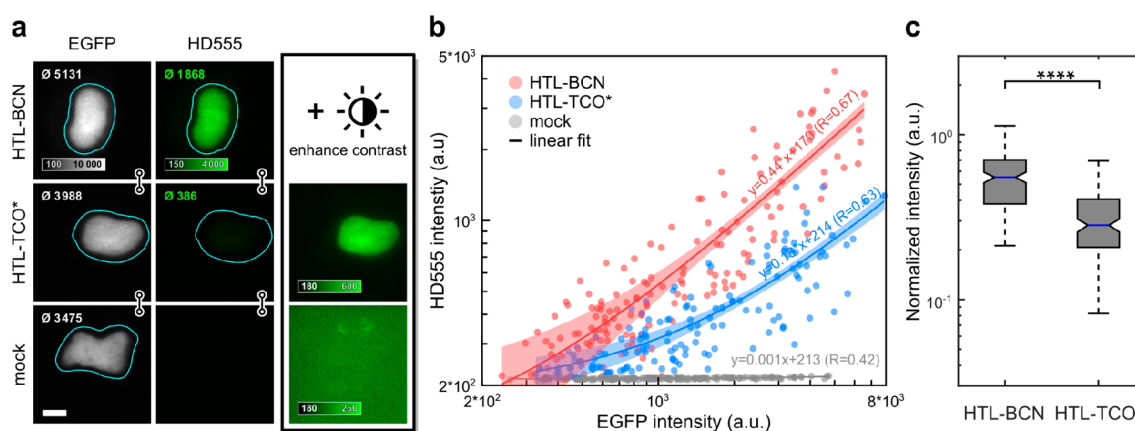


Figure 3. Fluorescence intensity quantification of **HD555** in live cells. COS-7 cells expressing H2B-EGFP–HaloTag labeled with **HD555** via HTL–BCN or HTL–TCO* and a mock control omitting any HaloTag linker. (a) Representative epifluorescence microscopy images of cells with comparable expression levels as indicated by mean intensities in segmented nuclei (top left corner of each image). The **HD555** channel is shown at identical display settings (indicated by linked images) and with enhanced contrast. (b) Mean nucleus intensities of 151 (HTL–BCN), 145 (HTL–TCO*), and 147 (mock) cells pooled from three independent experiments in EGFP and the **HD555** channel. Linear regression (line) with 95% confidence intervals. R: Kendall's tau correlation coefficient. (c) Box plot of **HD555** intensities normalized to EGFP are significantly higher for HTL–BCN compared to HTL–TCO* (Wilcoxon rank-sum test, $p < 10^{-23}$).

of the spiroether–zwitterion equilibrium of **sb-HD656** and its cycloadduct **29** (Figures 2f, S12). In both cases, we observed low absorbance at physiological pH, which indicates the predominance of the closed, nonfluorescent spiroether isomer (pK_{cycl} 5.2 and 5.5 for **sb-HD656** and **29**, respectively). The on-fraction of ~5% at physiological pH opens the possibility of SOFI, which does not rely on single-emitter localization and is therefore compatible with a wide range of illumination and blinking conditions.³⁷

Live-Cell Bioconjugation – Influence of Dienophiles on Fluorescence Intensities. Having confirmed generally favorable spectral and physicochemical properties of the dyes, we next evaluated their suitability for bioconjugation. Unreacted excess of the fluorophore in bioconjugation experiments is particularly difficult to remove in live-cell and in vivo experiments, leading to background signal and undesired artifacts. Here, fluorogenic probes can unfold their full potential given they maintain fluorogenicity in live cells. Therefore, after having shown successful in vitro labeling of BCN-modified EGFP (Figure S13), we evaluated the

fluorogenicity of HDyes in live-cell labeling and imaging. While we used BCN as a dienophile for in vitro characterization due to its formation of a single product, we tested both, BCN and *trans*-cyclooctene (TCO), as dienophiles in live cells. TCO derivatives have been reported to possess superior kinetics over BCN,^{38–40} and it is further known that the choice of the DA_{inv} dienophile can influence the fluorescence turn-on of tetrazine dyes.^{26,41,42} We therefore compared the brightness of cell-permeable **HD555** upon conjugation with a BCN- or (*E*)-cyclooct-2-en-1-ol (TCO*)-modified intracellular target. First, COS-7 cells transiently expressing the histone H2B fusion protein H2B–EGFP–HaloTag were loaded with respective HaloTag ligand–dienophile (HTL–dienophile), a bifunctional linker, to provide specific binding sites for fluorogenic labeling with **HD555**. With HTL–BCN, we observed specific staining with excellent contrast after dye incubation at 1 μ M over 2 h and mere buffer replacement (Figure 3a, top row). Second, we tested TCO* as a DA_{inv} dienophile under identical conditions. Here, we observed a specific signal but substantially lower contrast (Figure 3a,

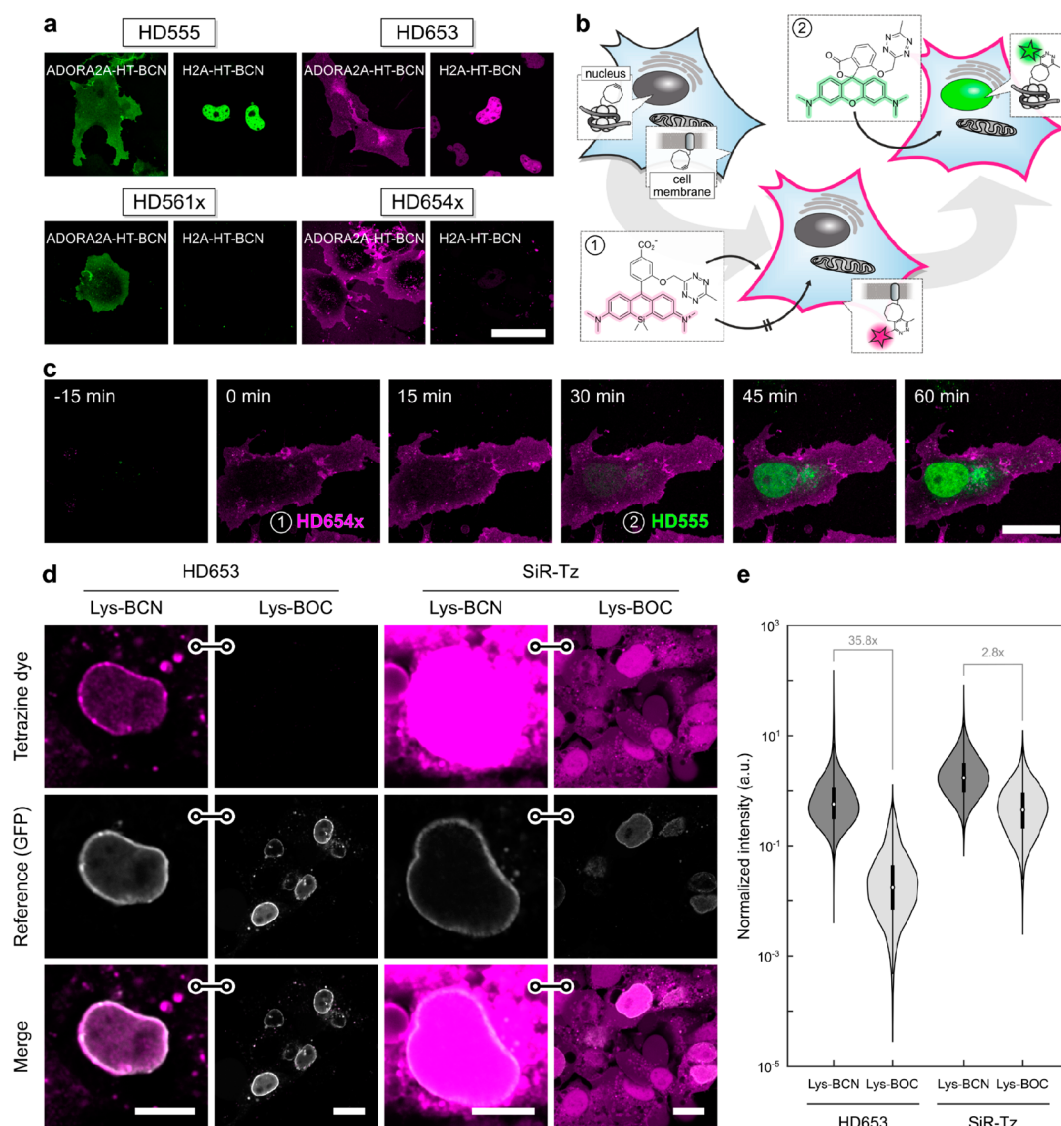


Figure 4. Wash-free multicolor and unnatural amino acid imaging in live cells. (a) Wash-free labeling of intra- and extracellular targets in COS-7 cells. Cells were transiently transfected with ADORA2A–HaloTag (plasma membrane) or H2A–HaloTag (nucleus), loaded with HTL–BCN, and incubated with HDyes at 1 μ M for 30 min. Scale bar 50 μ m. For reference and controls, see Figures S16–19. (b) Dual-color labeling with a single chemistry. Intra- and extracellular proteins H2A and ADORA2A are targeted via HaloTag and HTL–BCN and then incubated with cell-impermeable HD654x for 30 min followed by addition of HD555. (c) Time-lapse confocal microscopy shows labeling of intra- and extracellular targets with spectrally distinct fluorophores can be achieved without any washing. Scale bar 20 μ m. For individual color channels and reference staining, see (Figure S23). (d) Un-natural amino acid labeling of pEGFP^{N149TAG}–Nup153 with Lys–BCN or Lys–BOC (control) in COS-7 cells. Live-cell confocal microscopy after incubation with tetrazine dyes HD653 and SiR–Tz at 500 nM for 30 min and mere buffer replacement. Scale bars: 10 μ m (Lys–BCN) or 20 μ m (Lys–BOC). (e) Quantitative comparison of target and background labeling with flow cytometry analysis. pEGFP^{N149TAG}–Nup153 COS-7 cells were loaded with either Lys–BCN for specific or Lys–BOC for unspecific labeling. Intensity of HD653 and SiR–Tz was normalized to EGFP and signal-to-background ratios calculated from population medians.

middle row). Accordingly, mean intensities (see inset in nuclei images) of the nucleus were significantly lower for HTL–TCO* compared to HTL–BCN. A mock control omitting HTL–dienophiles (Figure 3a, bottom row) showed no unspecific staining from unreacted dye with intensity values at background level. For a quantitative evaluation of fluorescence intensities with HTL–BCN or HTL–TCO*, we compared intranuclear HD555 intensities against EGFP intensity, which served as an expression control (Figure 3b). For both dienophiles, we observed a linear correlation of HD555 and EGFP signal despite considerable cell-to-cell variation (Figure S14) and a 3.4-fold higher intensity obtained using HTL–BCN compared to HTL–TCO*. Considering the

observed variability, we additionally compared HD555 intensities normalized to EGFP, confirming the significantly reduced emission intensity obtained with TCO* in live cells (Figure 3c).

This was in good accordance with the different emission intensities obtained after reaction of HD555 with BCN and TCO* in vitro (Figure S15) and recent work by others that reported a strong dependency of fluorescence enhancement in DA_{inv} on the used dienophile.^{42,43} Due to the higher obtainable fluorescence brightness, we employed BCN as a DA_{inv} dienophile for cellular labeling in all following experiments.

Intra- and Extracellular Wash-Free Protein Imaging in Living Cells. Cell permeability of synthetic small-molecule

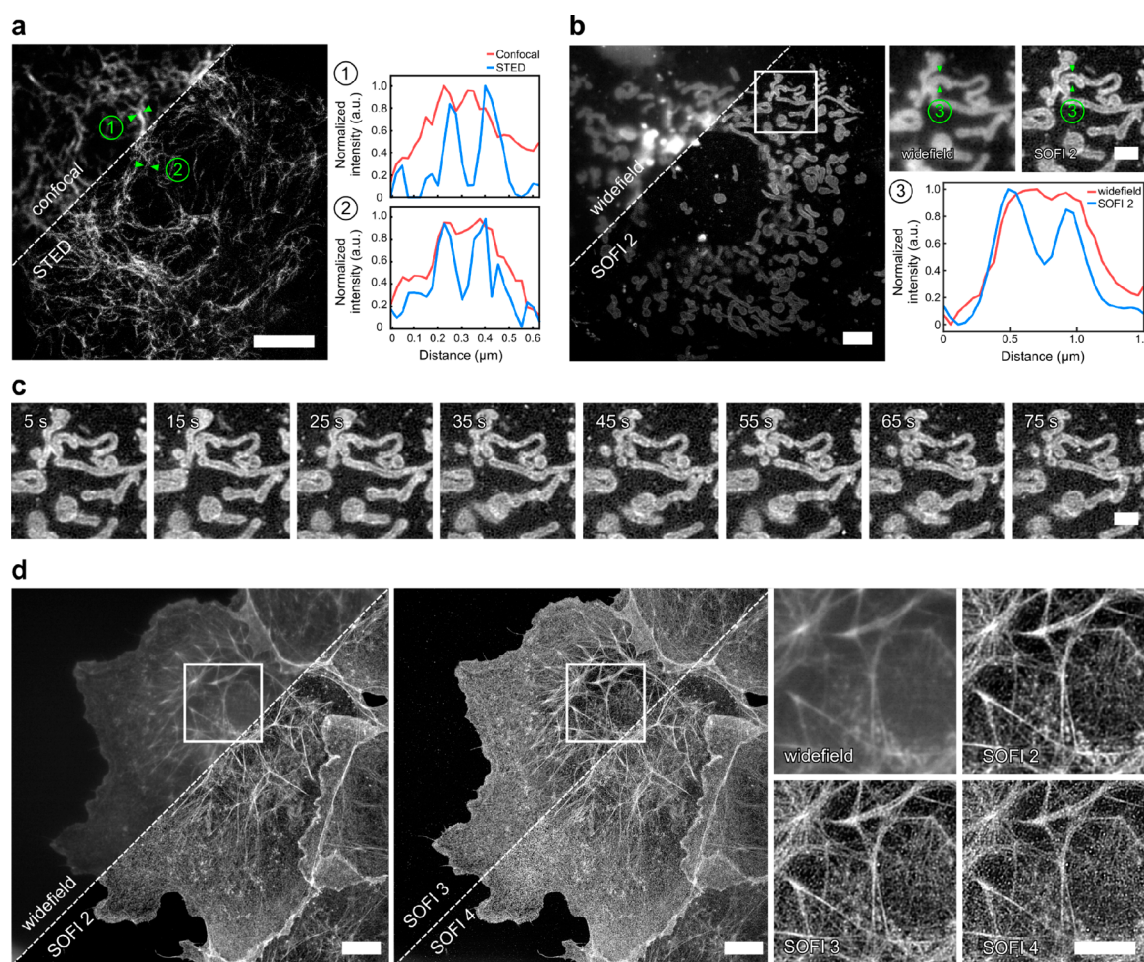


Figure 5. Super-resolution microscopy with HDyes. (a) Live-cell confocal and STED microscopy of COS-7 cells expressing pVimentin^{N116TAG→BCN} labeled with HD653 with intensity line profiles showing the resolution improvement. Scale bar 5 μm . (b) Live-cell widefield (temporal average of series) and second-order SOFI of mitochondria in COS-7 cells transiently expressing TOMM20-mCherry-HaloTag labeled with sb-HD656 via HTL-BCN. Scale bar 5 μm . The close-ups correspond to the ROI indicated in the overview image. The cross-sectional profile of the averaged widefield image and second-order SOFI shows the improved image contrast and resolution. Scale bar 2 μm . (c) Time course of second-order SOFI corresponding to the ROI indicated in (b) showing the movement of mitochondria with a time resolution of 10 s (see Figure S28 for the full sequence). Image acquisition: 500 frames per time point, 20 ms exposure time, 635 nm laser, 140 W/cm². The images are representative of more than five cells from two independent experiments. (d) High-order SOFI imaging of sb-HD656 labeled f-actin in fixed COS-7 cells showing SOFI analysis up to fourth cumulant order. Image acquisition: 20 000 frames, 50 ms exposure time, 635 nm laser, 275 W/cm². The images are representative of six cells from two independent experiments. Scale bars 10 μm .

fluorophores is a crucial factor in live-cell imaging. Hence, after having identified a suitable DA_{inv} dienophile for live-cell conditions, we next evaluated the cell permeability of the newly developed tetrazine dye series, including the potentially cell-impermeable 3,5-substituted derivatives. This was tested using HTL-BCN conjugated to HaloTag fusion proteins with intra- or extracellular localization. In transiently transfected COS-7 cells, all four HDyes showed specific extracellular labeling of adenosine A_{2A} receptor (ADORA2A) via HaloTag-BCN. Incubation with 1 μM dye for 30 min without further washing yielded uniform membrane staining with negligible background and excellent overlap with a reference label (Figures 4a, S16–19). To address an intracellular target, we incubated COS-7 cells transiently expressing histone H2A-HaloTag loaded with HTL-BCN for 30 min with 1 μM dye. Imaging under wash-free conditions showed that only HD555 and HD653, dyes capable of forming transient spirolactones, were able to pass through the plasma membrane and specifically label the nuclear target. Using HD654x and HD561x under

identical labeling and imaging conditions showed no nuclear signal (Figures 4a, S16–19). Controls without HTL-BCN or HaloTag fusion proteins yielded negligible signals for all four dyes (Figures S16–19, S22). In contrast to these zwitterionic and net neutral dyes, the positively charged dyes o-TzR and o-TzSiR showed significant unspecific mitochondrial accumulation (Figures S20, S21). To confirm the improved fluorogenicity and live-cell applicability of the new red-emitting dyes, we compared the SiR derivative HD653 with previously reported tetrazine-SiR conjugates in live-cell imaging. Under no-wash conditions, HD653 showed highly specific labeling and no visible off-target staining, while all other SiR derivatives tested exhibited varying degrees of off-target staining (Figure S22).

We then used complementary dye pairs with distinct cell permeability and absorption/emission wavelengths to perform dual-target labeling for two-color microscopy. First, we labeled an extracellular target at saturating concentrations with cell-impermeable HD654x, followed by labeling of an intracellular

target with cell-permeable **HD555** (Figure 4b). This concept allows to exploit the high efficiency of labeling via DA_{inv} with fluorogenic tetrazines to consecutively label two targets and overcomes the need for orthogonal labeling reactions. To demonstrate this, we coexpressed ADORA2A–HaloTag and H2A–HaloTag in COS7 cells and treated the cells with HTL–BCN. After washing to remove excess HTL–BCN, we incubated the cells with **HD654x** at 1 μ M for 30 min, followed by a second 30 min incubation with **HD555** at 1 μ M. Image acquisition was performed over time without additional washing steps after incubation with HDyes (Figure 4c). To achieve orthogonal labeling, complete saturation of extracellular BCN-modified proteins with the impermeable dye is crucial, to avoid cross reactivity of the second dye. This is achievable due to high reaction rates of the DA_{inv} reaction and elevated dye concentrations facilitated by effective quenching of unreacted dyes. Accordingly, unspecific background and cross-labeling of remaining extracellular target with the second added dye was not observed (for individual color channel images, see Figure S23). After the stepwise incubation with **HD655x** and **HD555** for a total of 60 min without further washing, we observed high contrast specific target labeling (Figure 4c), which could be confirmed by reference chase staining with HTL–Rhodamine110 (Figure S23). Spectrally interchanged labeling using **HD561x** for extra- and **HD653** for intracellular labeling yielded the expected comparable results (Figure S24). Therefore, our dyes enable live-cell two-color microscopy of an intra- and extracellular target under wash-free conditions with full spectral flexibility.

Encouraged by these results for intra- and extracellular labeling using protein tags, we next moved on to apply HDyes for live-cell protein labeling using UAAs. The labeling of UAA-modified proteins with small-molecule fluorophores via DA_{inv} results in a minimal size modification, which reduces unwanted perturbation of the target protein.^{28,44} Due to low cellular autofluorescence in the far-red spectral region, we anticipated the highest contrast from SiR dyes. We therefore used **HD653** for site-specific labeling of the nuclear pore complex via the Amber (TAG) mutant nucleoporin 153 construct (EGFP^{N149TAG}–Nup153) and performed a side-by-side comparison with commercial SiR–Tz (for structure, see Figure S25). To account for an additional fluorogenic effect due to environmental influence on the zwitterion–spirolactone equilibrium, we determined fluorescence turn-on values with a purified BCN-tagged protein of 55-fold for **HD653** and 3.1-fold for SiR–Tz (Figure S7). COS-7 cells were transiently transfected with the Nup153 construct in the presence of the respective genetic code expansion machinery and Lys–BCN, washed, and then labeled with **HD653** or SiR–Tz. Confocal microscopy (Figure 4d) showed that **HD653** yields comparable specific signal but substantially lower off-target background than SiR–Tz. We then used nonreactive Lys–BOC to further assess the labeling specificity and observed unspecific background signal from SiR–Tz in contrast to barely detectable background from **HD653** (Figure 4d). To quantify the labeling specificity, we performed flow cytometry analysis of cells expressing the NUP153 construct in presence (Lys–BCN) or absence (Lys–BOC) of specific reaction sites for **HD653** or SiR–Tz, respectively (Figures 4e and S26). Due to efficient quenching, the difference in signal of **HD653** (35.8 \times) was substantially higher compared to SiR–Tz (2.8 \times). This shows that the high fluorogenicity of **HD653** allows for UAA

live-cell labeling with minimal washing and enables high signal-to-background ratio for fluorescence imaging applications.

STED and SOFI Super-resolution Microscopy. With improving spatial resolution of imaging techniques, the size of fluorescent labels becomes increasingly important. Having shown that specific labeling of UAA with improved contrast could be achieved with our dyes, we evaluated their use in super-resolution microscopy. In this context, we used **HD653** in combination with UAAs as minimal size labels. Based on previous reports using structurally akin SiRs,^{12,24,32} we evaluated the performance of **HD653** in STED microscopy. To test its applicability, we labeled the Amber mutant vimentin^{N116TAG}–mOrange construct with Lys–BCN and **HD653** in live COS-7 cells. STED imaging using a 775 nm continuous wave laser for depletion resolved vimentin structures and clearly showed enhanced resolution compared to confocal microscopy (Figure 5a). Of note, after 50 cycles of STED imaging, bleaching for **HD653**-labeled vimentin was lower compared to commercial SiR–Tz (Figure S27).

Another approach for super-resolution microscopy uses stochastic temporal emission fluctuations of single fluorophores. Based on our own previous work,³⁵ we used a self-blinking fluorogenic tetrazine–SiR for minimally invasive live-cell super-resolution microscopy. While SMLM can achieve resolutions down to a few nanometers,⁴⁵ it requires sparse signals to allow for localization of individual emitters. In contrast, SOFI exploits spatially and temporally correlated intensity fluctuations for resolution improvement without the need of isolated emitters.⁴⁶ This is especially beneficial for live-cell microscopy of highly abundant targets at low excitation intensities. We used the self-blinking derivative **sb-HD656** in combination with SOFI for subdiffraction imaging in living and fixed cells. First, we performed live-cell imaging of mitochondrial import receptor protein TOMM20 labeled via HaloTag and HTL–BCN with **sb-HD656** at low illumination intensities in the near-infrared (~ 140 W/cm² at 635 nm). Due to low phototoxicity, it was possible to perform time-lapse imaging of the rearrangement of the mitochondrial network in COS-7 cells (Figure 5c, for longer time course see Figure S28). Second-order cumulant analysis required only a few hundred frames and resolved the mitochondrial outer membrane (see close-up and line profiles in Figure 5b). In contrast to SMLM, SOFI was also compatible with higher active emitter densities as encountered with highly abundant histone protein H2B in nuclei of live COS-7 cells (Figure S29).

In order to demonstrate higher-order SOFI imaging enabled by the high photostability of HDyes, **sb-HD656** was used to stain the cytoskeletal actin network. We exploited the versatility offered by DA_{inv} tetrazine labeling and used a small-molecule labeling approach with phalloidin, a natural toxin with a high affinity to f-actin to introduce BCN as DA_{inv} reaction partner. To this end, we targeted f-actin with phalloidin–BCN in fixed COS-7 cells and subsequently labeled it with **sb-HD656**. The pH of the imaging buffer was chosen for a reduced on/off ratio for high-order SOFI analysis (Figure S30). **sb-HD656** was found to be photostable at relatively low illumination intensities (275 W/cm²), thus allowing the acquisition of up to 20 000 frames with moderately low photobleaching (Figure S31). SOFI analysis was performed up to fourth cumulant order while maintaining sufficiently high signal-to-noise ratio (Figure 5d). Compared to widefield imaging, second-order SOFI provided a significant contrast enhancement, while higher-order (third–fourth) SOFI images

allowed to resolve individual actin filaments that were not distinguishable in widefield images. Additionally, we compared **sb-HD656** to a previously published self-blinking tetrazine probe.³⁵ The use of **sb-HD656** resulted in higher on/off ratios and on times which makes it well suited for SOFI (Figure S32).

CONCLUSION

We developed a series of red and far-red fluorogenic tetrazine dyes (HDyes) for live-cell bio-orthogonal labeling of intra- and extracellular targets. The design rationale with tetrazines flexibly linked to the fluorophore at minimal distance led to efficient fluorescence quenching by Dexter exchange. HDyes exhibited high fluorogenicity and were well-suited for live-cell imaging under wash-free conditions as demonstrated in two-color labeling experiments and STED super-resolution imaging of UAA-labeled target proteins. The modular synthetic route provided a self-blinking HDye, which was successfully exploited in long-term live-cell SOFI imaging. We expect the HDyes to leverage applications of bio-orthogonal chemistry in the field of super-resolution microscopy. Their excellent photophysical properties, improved signal-to-background ratios, and small labeling size are ideal prerequisites for the direct labeling of UAA-modified proteins, which has great potential of reducing the linkage error. We expect the herein presented tetrazine–xanthene scaffold to serve as general motif in the development of numerous other fluorogenic probes in the future.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acscentsci.1c00703>.

Syntheses, compound characterization data, and details of the computational and time-resolved spectroscopy studies as well as of imaging experiments (PDF)

Structural data of HD555 as a Crystallographic Information File (CIF)

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Notes

The authors declare no competing financial interest. Crystallographic data has been deposited at the Cambridge Crystallographic Data Centre (CCDC) under the CCDC deposition number 2021104. The data can be obtained free of charge via <https://www.ccdc.cam.ac.uk/structures>.

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